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Variation in andrographolide content among different accessions of *Andrographis paniculata*

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Abstract

Andrographis paniculata (Burm. f.) Wall. ex Nees, commonly known as Kalmegh is used both in Ayurvedic and Unani system of medicines for a number of ailments related to digestion, liver (hepatic), fever, malaria and sore throat. It has an imperative place in the Indian Pharmacopoeia and is being prominently used in at least 26 Ayurvedic formulations. It is a hardy and erect plant which grows mainly as under shrub in tropical, moist deciduous forest. It is widely distributed southwards through Thailand and Peninsular Malaysia to Indonesia and in India it is found in the states of Madhya Pradesh, Chhattisgarh, Orissa, Jharkhand, Maharashtra, Assam, Bihar, West Bengal, Uttar Pradesh, Andhra Pradesh, Tamil Nadu, Karnataka and Kerala. Andrographolide, is the major constituent extracted from the plant and exhibits several biological activities. The quality of the herb depends on andrographolide content. The andrographolide content was determined by HPTLC. The average andrographolide content varied from 1.38 to 3.12 % on dry weight basis. The differences in andrographolide content among Kalmegh collected from different locations were statistically significant. The result indicated that the andrographolide being secondary metabolite may be influenced by the environmental, seasonal factors and soil characteristics. The results indicated that Kalmegh populations with highest andrographolide content may be potential source of quality raw material and ultimately more efficacious drugs.

Keywords: andrographolide, *Andrographis paniculata*, Accessions, HPTLC

Introduction

The history of herbs is as long as the history of mankind. Herbs are plants valued for their medicinal and aromatic properties and are often grown and harvested for these unique properties. Medicinal plants play an important role in disease management and livelihoods of people worldwide. There are about 15,000 to 20,000 plant species reported to have medicinal value with 30% considered as endemic to India. Among these 7,000-8,000 are reported to be used in unregulated informal systems of medicine and 1,200-2,000 in the regulated AYUSH^[1]. Their active phytoconstituents are mainly responsible for these potential medicinal effects. In recent years, the growing demand for medicinal plants has accelerated over exploitation of valuable resources by unscientific and destructive manner without considering supportability and quality issues. Distinctive ecological factors, for example, temperature, humidity, altitude, rainfall and genetic makeup affect the qualitative and quantitative nature of secondary metabolites present in plants.

Andrographis paniculata (Burm. f.) Wall. ex Nees, commonly known as “king of bitters”, traditionally known as ‘Kalmegh’, green chirayta is an annual herb widely used in tropical Asia. It is distributed southwards through Thailand and Peninsular Malaysia to Indonesia and in India it is found in the states of Madhya Pradesh, Chhattisgarh, Orissa, Jharkhand, Maharashtra, Assam, Bihar, West Bengal, Uttar Pradesh, Andhra Pradesh, Tamil Nadu, Karnataka and Kerala^[2]. The plant holds an imperative place in the Indian Pharmacopoeia and is being prominently used in at least 26 Ayurvedic formulas^[3]. Panchang (stem, leaf, flowers, seed and root) of the plant is being used in various formulation of Indian system of medicine. The leaves and aerial parts of the plant are used in Indian traditional medicine for the treatment of fever, malaria and sore throat^[4]. Clinical information shows the adequacy of the plant for the treatment and prevention of the common cold, tonsillitis and diarrhea. The plant is additionally announced powerful against intestinal sickness^[5]. The whole plant has variety of therapeutic values. It has immunosuppressive and alexipharine properties and is useful in wounds, ulcers, leprosy, sore throat, tonsillitis, osteodynea, menstrual and post partum haematometra, hypertension etc^[6]. Decoction of the plant is a blood purifier and is used to cure liver disorder, jaundice and dermatological disease^[7].

A. paniculata has been phytochemically investigated for a number of bioactive compounds including andrographolide, neoandrographolide, panaculoside, flavonoids, andrographonin, panicalin, apigenin-7, 4'-di-O-methyl ether. The plant contains diterpenoids: 14-deoxy-11-oxo-andrographolide; 14-deoxy-11, 12-dehydroandrographolide; 14-deoxyandrographolide; neoandrographolide and andrographolide [8]. The major bioactive constituent 'Andrographolide' constitutes a group of diterpene lactones mainly found in leaves whereas stems contain the compound in traces [9]. In accordance with bare minimum standard of acceptability specified in pharmacopoeia, herbal industries prefer to receive Kalmegh (aerial parts) with not less than 40% leaves and total andrographolides content not less than 1.8% w/w. For leaves the industry expects not less than 2.8% w/w of total andrographolides which would contain at least 2.5% w/w of pure andrographolide.

The concentration of these active ingredients varies within plant parts and with the geographical distributions of the species. The andrographolide being secondary metabolites are often influenced by the environmental, seasonal factors and its distribution in between leaves and whole plant. It has been reported that there is wide variation in the andrographolide content in leaves and whole plant. There is also significant variation in andrographolide content in Kalmegh collected from different geographical areas [10]. Phytochemical marker compound (andrographolide) showed quantitative variations among the plants of different locations [11]. Chemical composition of plants varies not only in different parts but also with respect to other factors like growing regions, agroclimatic conditions, genetic makeup etc. [12, 13] The study therefore was aimed to calculate the percentage of andrographolide among different accessions of *A. paniculata* collected from different locations.

Materials and Methods

Plant Material

Germplasm of *Andrographis paniculata* (Kalmegh) was collected from Jharkhand, Madhya Pradesh, Uttar Pradesh, Karnataka, Gujarat, Haryana, Odisha and Uttarakhand. and also from research institutions like CIMAP Lucknow; NBRI Lucknow; SFRI Jabalpur; Anand Agricultural University, Anand; IIHR Bangalore; FRLHT Bangalore; Natural Remedies Bangalore; Dabur India Ltd. and Patanjali Ayurved Haridwar. Plant cuttings/poly potted plants of five to fifty (randomly sampled individual plants) from each location were brought to Forest Research Institute, Dehradun and planted together in polybags/beds filled with soil and farmyard manure. These plant cuttings/germplasms were maintained in the nursery of the institute as mother plants. The plants were watered regularly and allowed to grow and proliferate. The plants were harvested, washed, shade dried and finally ground to powder for further analysis.

Chemicals and analytical equipment

All chemical used for extraction and analysis were of analytical grade and referred to Emerck. For quantitative assessment of kalmegh accessions by HPTLC, solvents including methanol, toluene, chloroform, acetone, vanillin, sulphuric acid and water used were of HPLC grade procured from Merck Life Science Private Limited (Mumbai, India).

TLC Aluminium pre-coated plates with Silica gel 60 F₂₅₄ (20X20 cm²) used were obtained from E. Merck Ltd. (Mumbai, India). Andrographolide standard 99.8% was procured from Natural Remedies, Bangalore and was used as standard biomarker to confirm the results and to find correct percentage of andrographolide. A CAMAG (Switzerland) HPTLC system equipped with a sample Linomat V, Twin trough Glass Chamber (20 x 10) with SS lid, TLC Scanner III and Wincats an integrated Software 4.02(Switzerland).

Extraction and Sample preparation for HPTLC Analysis

Powdered plant material (1 gm) of each accession were extracted with methanol (15ml x 3) through refluxing on water bath for 20 min and filtered. The extracts obtained were subjected to estimation of andrographolide content using HPTLC. Extracts obtained from all accessions were makeup upto 50ml with methanol in volumetric flask.

Estimation of Andrographolide

For HPTLC analysis conditions were as follows- application volume- 10 µL, as 5-10 mm bands; developing solvent system- chloroform, acetone and toluene (2:2:1) and spray reagent- a mixture containing 1% vanillin in alcohol and 10% sulfuric acid in alcohol (1:1) and wavelength 223nm.

Results and Discussion

The germplasm of *A. paniculata* were collected from different locations of India and was phytochemically evaluated for the key therapeutically important compound, andrographolide. In HPTLC, andrographolide band of plant extract was detected at R_f value 0.31± 0.03 in Chloroform: Acetone: Toluene (2:2:1) mobile phase. The spectra of andrographolide showed maximum absorption at 223nm (Fig.1). The chromatogram of andrographolide from plant samples were obtained and compared with chromatogram of andrographolide standard on the basis of retention factor and peak area (Fig. 2 and Fig. 3). Twenty eight accessions of *A. paniculata* were analysed for active ingredient andrographolide by HPTLC.

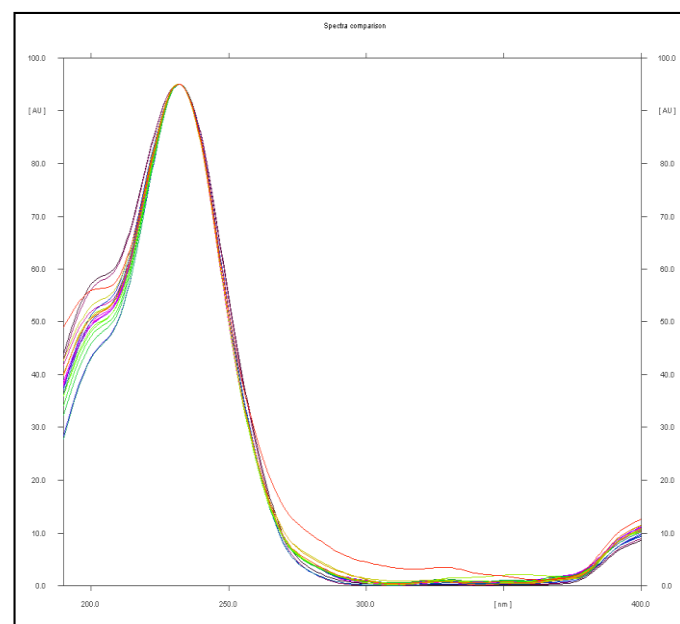


Fig 1: Andrographolide HPTLC Spectrum

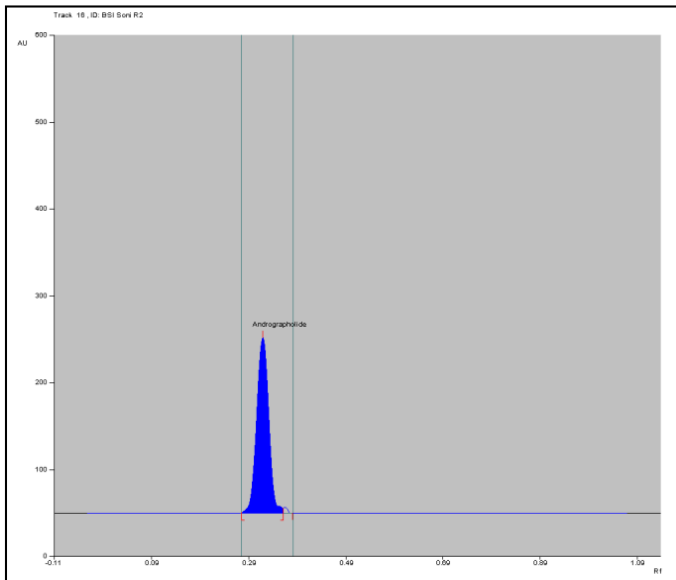


Fig 2: Andrographolide Standard Peak

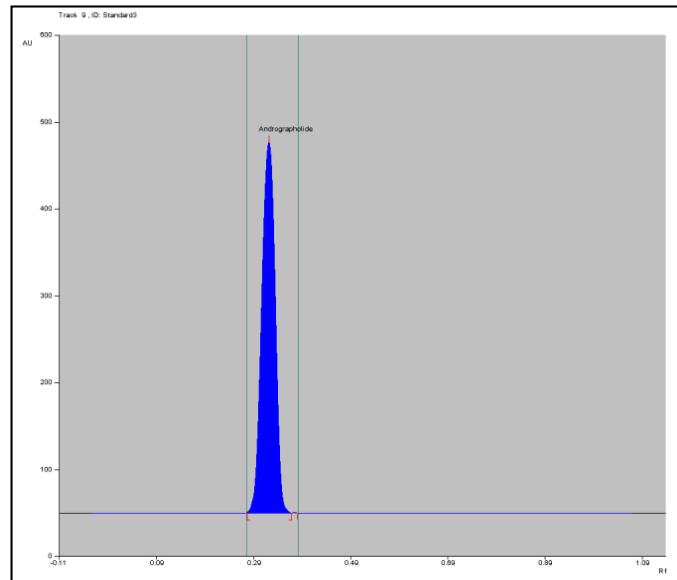


Fig 3: *Andrographis paniculata* Sample peak

Table 1: Percentage of Andrographolide among different accessions of *Andrographis paniculata*

Accession	Location	Andrographolide content % on DWB
1.	IIHR Bangalore	3.12
2.	Ranchi	1.94
3.	Patanjali Haridwar	2.17
4.	CIMAP, Lucknow	2.59
5.	Rajeshwari Nursery, Haridwar	1.84
6.	F.R.I, Dehradun	2.58
7.	Gujarat Agriculture University-1	2.92
8.	Gujarat Agriculture University -2	2.00
9.	Anand Gujarat 1	2.50
10.	Anand Gujarat 2	1.84
11.	Anand Gujarat 3	2.49
12.	Anand Gujarat 4	1.92
13.	Anand Gujarat 5	2.50
14.	FRLHT, Bangalore	2.31
15.	Natural Remedies, Bangalore	1.75
16.	Ayurved	2.11
17.	Dabur	2.05
18.	Bhopal	2.35
19.	R.K.Mission, Kolkata	2.25
20.	BSI, Kolkata	2.17
21.	NBPGR, Delhi	1.56
22.	RPRC, Bhuvneshwar	1.91
23.	SMPB, Bhuvneshwar	2.16
24.	Shantikunj, Haridwar	1.78
25.	Dr. Sushila Devi Herbal Garden, Rishikesh	2.04
➤	CD value at 5 % level	0.226
➤	CD value at 1 % level	0.313

*DWB- Dry Weight Bases

The studies revealed the variation in andrographolide content ranged from 1.38 to 3.12 % on dry weight basis (Table 1). The highest andrographolide content was found in IIHR

Bangalore accession (3.12%), followed by 2.92% in GAU 1 and 2.59% in CIMAP (Fig. 4)

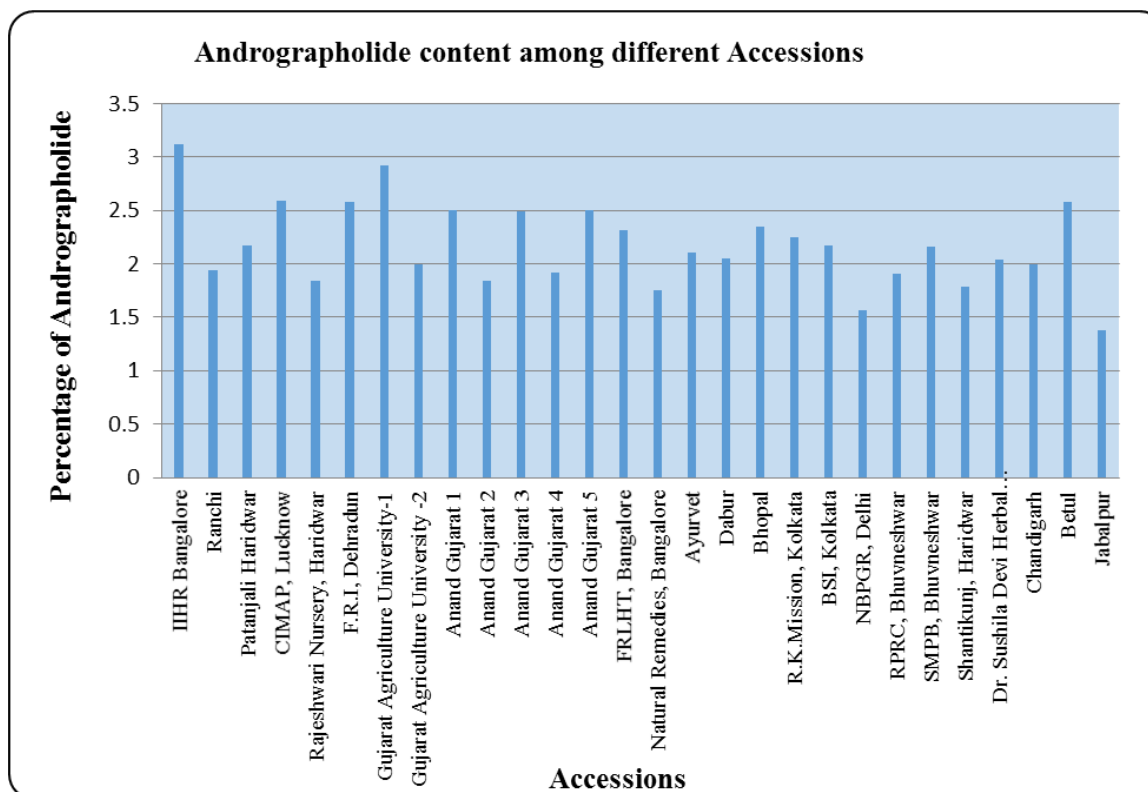


Fig 4: Graphical representation of Variation in Andrographolide content.

Variability in andrographolide content has previously been assessed by various workers. Mishra *et al.* 2001 has reported 0.67% to 1.82% of andrographolide content in leaves while 0.33% to 0.83% in complete plant of *A. paniculata* collected from Madhya Pradesh [10]. Another study carried out by Sabu *et al.* 2001 reported 0.73% to 1.47% andrographolide in leaves [14]. While Pandey and Mandal 2010 reported a variability of 1.07 to 2.24 % in andrographolide content in dried leaves collected from five locations of Madhya Pradesh and Chattisgarh [11]. Further Raina *et al.* 2007 has reported a variation of andrographolide content in dry leaves from 1.14% to 2.60% [15], Sharma *et al.* (2013) conducted a study on different harvesting time reporting the content vary from 0.81% to 1.86% before and after flowering [16]. Bhan *et al.* (2006) has concluded that the total andrographolide concentration in leaves increased from 6.23% to 6.96% from September to November [17].

Conclusion

Study was conducted with the objective to find out superior material of Kalmegh in terms of active ingredient (andrographolide) to obtain quality drug. The average andrographolide content varied from 1.38 to 3.12 % on dry weight basis. The differences in andrographolide content among Kalmegh collected from different locations were statistically significant. The highest andrographolide content was found in IIHR Bangalore accession (3.12%), followed by 2.92% in GAU 1 and 2.59% in CIMAP. The study revealed that the andrographolide content being secondary metabolite may be influenced by the environmental, seasonal factors and soil characteristics. Genetic makeup of the germplasm also contributed in the quality of raw material. The results indicated that populations/germplasm having highest andrographolide content may be potential source for good quality raw material vis-à-vis production of more efficacious drugs.

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Conflict of Interest

Conflict of interest declared none.

References

1. Qazi GN. Resources, technologies and knowledge-sharing on MAPs from India, In: Workshop on Strengthening Cooperation of MAPs National Focal Points, ICS-UNIDO, Trieste, Italy, 2003, 26-27.
2. Ghosh KB, Datta KA, Mandal A, Dubey PK, Halder S. An Overview on *Andrographis paniculata* (Burm. F.) Nees. International Journal of Research in Ayurveda and Pharmacy, 2012; 3(6):752-760.
3. Nadkarni AK. Nadkarni's Indian Materia Medica. 1st ed. Popular Book Depot. Bombay, 1954, I.
4. Chopra RN, Nayar SL, Chopra IC. Glossary of Indian medicinal plants. 1st ed. Publication and Information Directorate, CSIR, New Delhi, 1956.
5. Misra P, Pal NL, Guru PY, Katiyar JC, Srivastava V, Tandon JC. Antimalarial activity of *Andrographis paniculata* (Kalmegh) against Plasmodium berghei NK 65 in *Mastomys natalensis*. Int J Pharmacol. 1992; 30:263-274.
6. Puri AR, Saxena RP, Saxena KC, Saxena V, Srivastava V, Tandon JS. Immunostimulant agents from *Andrographis paniculata*. J Natural Products 1996; 56(7):995-999.
7. Prathanturug S, Soonthornchareonnon N, Chuakul W, Saralamp P. Variation in growth and diterpene lactones among field-cultivated *Andrographis paniculata*. Journal of Natural Medicines. 2007; 61(2):159-163.

8. Pawar RK, Sharma Shivani, Singh KC, Sharma Rajeev KR. Development and Validation of HPTLC Method for the determination of Andrographolide from *Andrographis paniculata* (Whole Plant). International Journal of Chemistry Research, 2010; 3(2):85-89.
9. Saxena S, Jain DC, Bhakuni RS, Sharma RP. Chemistry and pharmacology of *Andrographis* species. Indian Drugs, 1998; 35:458-467.
10. Misra HO, Sharma JR, Lal RK, Shukla N. Patterns of genetic variability for different traits in a collection of Kalmegh (*Andrographis paniculata*) genotypes. Journal of Medicinal and Aromatic Plant Sciences. 2001; 22(4A), 23(1A):348-51.
11. Pandey AK, Mandal AK. Variation in morphological characteristics and andrographolide content in *Andrographis paniculata* (Burm. f.) Nees of central India, Iranica Journal of Energy & Environment. 2010; 1(2):165-169.
12. Patarapanich C, Laungcholatan S, Mahaverawat N, Chaichantipayuth C, Pummangura S. HPLC determination of active diterpene lactones from *Andrographis paniculata* Nees planted in various seasons and regions in Thailand. Thailand Journal of Pharmaceutical Sciences. 2007; 31:91-99.
13. Mishra S, Tiwari SK, Kakkar A, Pandey AK. Chemoprofiling of *Andrographis paniculata* (Kalmegh) for its andrographolide content in Madhya Pradesh, India, International Journal of Pharma and Bio Sciences. 2010; 1(2):1-5
14. Sabu KK, Padmesh P, Seeni S. Intraspecific variation in active principle content and isozymes of *Andrographis paniculata* Nees (Kalmegh): a traditional hepatoprotective medicinal herb of India. Journal of Medicinal and Aromatic Plant Sciences. 2001; 23:637-647.
15. Raina AP, Kumar A, Pareek SK. HPTLC analysis of hepatoprotective diterpenoid andrographolide from *Andrographis paniculata* Nees (Kalmegh). Indian Journal of Pharmaceutical Science. 2007; 69:473-475.
16. Sharma M, Sharma GR. Identification, purification and quantification of Andrographolide from *Andrographis paniculata* (Burm. F.) Nees by HPTLC at different stages of life cycle of crop. J Curr Chem Pharm Sci. 2013; 3(1):23-32.
17. Bhan MK, Dhar AK, Khan S, Lattoo SK, Gupta KK, Choudhary DK. Screening and optimization of *Andrographis paniculata* (Burm. f) Nees for total andrographolide content, yield and its components. Scientia Horticulturae. 2006; 10:386-391.