



ISSN (E): 2277- 7695
ISSN (P): 2349-8242
NAAS Rating: 5.23
TPI 2021; 10(11): 1483-1488
© 2021 TPI
www.thepharmajournal.com
Received: 11-08-2021
Accepted: 30-10-2021

G Raghavi
PG Scholar, Department of Plant Breeding and Genetics, Agricultural College and Research Institute, Tamil Nadu Agricultural University, Killikulam, Thoothukudi, Tamil Nadu, India

S Merina Prem Kumari
Associate Professor, Department of Plant Breeding and Genetics, Agricultural College and Research Institute, Tamil Nadu Agricultural University, Killikulam, Thoothukudi, Tamil Nadu, India

M Arumugam Pillai
Professor and Head, Department of Plant Breeding and Genetics, Agricultural College and Research Institute, Tamil Nadu Agricultural University, Killikulam, Thoothukudi, Tamil Nadu, India

S Saravanan
Assistant Professor, Department of Plant Breeding and Genetics, Agricultural College and Research Institute, Tamil Nadu Agricultural University, Killikulam, Thoothukudi, Tamil Nadu, India

A Kavitha Pushpam
Associate Professor, Soil science, Department of Soil Science and Agricultural Chemistry, Agricultural College and Research Institute, Tamil Nadu Agricultural University, Killikulam, Thoothukudi, Tamil Nadu, India

Corresponding Author:
G Raghavi
PG Scholar, Department of Plant Breeding and Genetics, Agricultural College and Research Institute, Tamil Nadu Agricultural University, Killikulam, Thoothukudi, Tamil Nadu, India

Improvement of rice cultivar Kavuni for semi dwarf character by *in vitro* chemical mutation using Ethyl methyl sulphonate

G Raghavi, S Merina Prem Kumari, M Arumugam Pillai, S Saravanan and A Kavitha Pushpam

Abstract

The present study investigated the effect of an EMS treatment on the embryonic calli of Kavuni for the characters *viz.*, late flowering, lodging, and long duration and improvement for these aspects will promote black rice cultivation. The experiment was conducted using Kavuni embryonic calli that showed 86.5% callus induction percentage and it was treated with EMS at 0.05%, 0.1%, 0.15% and 0.2% level along with control for mutagenesis to create dwarfism. The LC₅₀ for EMS treatment based on callus survival was 0.10%. The maximum regeneration percentage of EMS treatment was 53.6% in the control and 0.1% EMS treatment recorded 40%. The lowest plant height of 20.2 cm in 0.05% EMS treatment. Nutritional analysis recorded the maximum protein content of 5.50% at 0.05% EMS and carbohydrate content was maximum of 54.56 at 0.05% EMS. The pigment *viz.*, β-carotene, chlorophyll and anthocyanin were maximum of 0.02 µg/g, 2.25mg/g and 1.34 mg/g respectively at 0.05% EMS treatment. Total phenolics with the highest level of 28.5 mg/g was recorded at 0.05% EMS. The *in vitro* mutated rice Kavuni plantlet at 0.05% EMS concentration has improved characters and selection will be made in the subsequent generation for dwarf trait.

Keywords: Kavuni, *in vitro* mutation, LC₅₀ value, regeneration, morphological and biochemical characterisation

Introduction

Rice is one of the most important cereal crop of the world meeting the dietary requirement of people living in tropics and subtropics. Asia accounts 90% of production and consumption of rice (Khush 2005) [18]. India has the world's largest area of rice cultivation in 42.5 million hectares and is the second largest producer next to China. Kavuni, a traditional brownish black rice variety is known for its anti-diabetic properties and is grown under limited area due to its long duration, poor tillering and photosensitive period. Kavuni grains are rich source of vitamins and antioxidants and its dark purple colour is primarily due to its anthocyanin content (Purwanto *et al.*, 2019) [31]. It is suitable for making porridge, dessert, traditional Chinese black rice cake, bread and noodles.

Black rice possesses significantly higher levels of β-carotene (46.2 µg/100g), lutein (221.6 µg/100g), total phenolics (28.8 µg/100g) and anti-oxidants activity. The health benefits of black rice are diverse and it prevents cancer and diabetes, reduces inflammation and body weight, improves liver and kidney health and builds immunity (Valarmathi *et al.*, 2015) [33]. A high antioxidant activity is contributed by phenolic compounds comprising of vanilic acid, caffeic acid, ferulic acid, p-coumaric acid, protocatechuic acid and 4-OH benzoic acid. In black rice the anthocyanin content ranged from 109.5 to 256.6 mg/100g.

Dwarfism is an agronomically important trait in breeding for resistance to damage by wind, rain and for stable high yields that increases the harvest index. Some of the varieties are very tall (>160cm) in stature and exhibit very late maturity (>150 days). Black rice and cultivars are hard to find currently and even nearing extinction. Black rice has the good flavour and aroma and is delicious. Due to the problem of tall stature, it is prone to lodging and late maturity with more fertilizer requirement.

Tissue culture technique offers a great potential for crop improvement through *in-vitro* mutagenesis by chemical mutagens. Mutation induction has become an established tool in crop improvement to improve cultivars for certain specific traits (Penna *et al.*, 2012) [30].

Radiation mutagenesis combined with *in vitro* culture has proven to be an useful method for the induction and selection of novel genetic variabilities (Jain 2010).

Brassinosteroids are involved in many morphological and physiological processes of rice, including the elongation and unrolling of leaves, development of tillers, root differentiation and reproductive growth (Mori *et al.*, 2007) [25]. The reduction in plant height is enhanced from EMS concentration of 0.05% to 0.2%. The objective of this research is to develop early maturing Kavuni mutant with shorter plant crown and higher productivity by chemical mutation using ethyl methane sulphonate (EMS). Also, identification of Kavuni dwarf mutant lines with good combination of nutritional qualities will be used for future breeding programmes.

Materials and Methods

The research was conducted at Agricultural College and Research Institute Killikulam. Healthy, pure, uniform and dry seeds of Kavuni rice were inoculated for callus induction in MS media (Murashige and Skoog, 1962) [26] as described by (Amaravel 2019) [2]. Also the embryogenic calli formed was treated with different EMS concentrations of 0.05%, 0.1%, 0.15% and 0.2% as described by Venkateshan, 2021 [34]. After two weeks of inoculation callus induction percentage was calculated and survival percentage was recorded after EMS treatment. Relative growth rate and RDR were calculated after three times of sub culturing.

$$\text{Callus induction percentage} = \frac{\text{Number of seeds that produced callus}}{\text{Total seeds cultured (100)}} \times 100$$

$$\text{Survival percentage} = \frac{\text{Number of calli that survived}}{\text{Total number of gamma irradiated calli}} \times 100$$

$$\text{Relative growth rate (RGR \%)} = (W_n - W_o / W_o) \times 100$$

Where

W_n Weight of the calli at the end of the experiment

W_o Weight of the calli at the start of the experiment

$$\text{Relative differentiation rate (RDR \%)} = \frac{\text{No. of survived calli in treatment}}{\text{No. of survived calli in control}} \times 100$$

The irradiated calli after sub culturing were inoculated in regeneration media consisting of BAP 3mg/l, NAA 1mg/l, maltose 30g/l and clerigel 3g/l (Amaravel *et al.*, 2019) [2]. The regeneration percentage of calli at various EMS treatments was calculated.

$$\text{Regeneration percentage} = \frac{\text{Number of calli that produced green shoot}}{\text{Total number of calli transferred to regeneration media}} \times 100$$

The *in vitro* regenerated shoots were transferred for *in vitro* rooting to half MS medium without hormones for two weeks and the plantlets were transferred to greenhouse condition for two weeks and subsequent to shade net house for hardening. After three weeks of hardening, biochemical analysis was done in Kavuni rice mutant plantlets. Anthocyanin pigment content (Pedro *et al.*, 2016) [29], chlorophyll content (Khalil *et al.*, 2018) [17], β -carotene, protein content (Lowry *et al.*, 1951) [23] and carbohydrate content (Hodge *et al.*, 1962) [13] were analysed. The LC_{50} values were calculated based on the survival percentage and relative differentiation rate of calli using curve expert 1.4 programme. The experimental data sets observed were analysed descriptively using the analysis of variance at 5% confidence level using DMRT analysis.

Results and Discussion

Callus induction percentage

In rice tissue culture, the callus induction mainly depends on the media composition, genotype of the mother plant, explants type and plant growth regulators (Khaleda and Forkan, 2006). The maximum callus induction was observed in MS Medium (Murashige and Skoog, 1962) [26] supplemented with 2,4-D 2mg/l, kinetin 0.5mg/l and sucrose 30g/l. The callus induction percentage was observed as 86.5% in the cultivar, Kavuni (Table 1.). According to Biswas and Mandal (2007) [4], MS Medium supplemented with 2,4-D and kinetin had a significant effect on callus induction percentage.

Paul and Roychoudhry (2019) [8] observed callus induction percentage in aromatic rice cultivars that ranged from 63.19% to 98.33% in eight rice cultivars and similar results by Bolivar *et al.* (2018) [5] in coffee. Luan *et al.* (2007) [24] in potato found mutagenic effect in embryonic calli when treated with different concentrations of EMS

In vitro mutagenesis of rice calli for dwarfism

Survival percentage, Relative Growth Rate and Relative Differentiation Rate of EMS treated calli

A gradual decrease in the survival percentage of calli was observed when exposed to an increasing concentration of EMS from 0.05% to 0.2% when compared to that of control (Table 2.). In the EMS treated calli, the maximum survival percentage was observed as 86.6% in control followed by 50%, 36.67%, 26.68% and 13.33% in 0.05%, 0.1%, 0.15% and 0.2% EMS concentrations of respectively. Acanda *et al.* (2014) reported a reduction in the embryonic calli survival rate at 10mM EMS concentration when compared with 1mM and 3mM EMS treatments in grapevine and similar decreasing survival percentage of calli was reported by Kumar *et al.* (2010) in rough lemon, Ekanayaka *et al.* (2016) in rice and Khalil *et al.* (2018) [17] in sugarcane. According to Das *et al.* (2010), a reduction in survival percentage depends on the EMS concentration and duration of the treatment in *Withania*.

The relative growth rate of the callus showed a linear decrease in the growth of the callus as the EMS concentration increases from 0.05% to 0.2% EMS and significant difference was observed among the various EMS concentrations. The highest relative growth rate of calli in the variety Kavuni was observed as 1.64 in control without EMS treatment followed by 0.86 at 0.05% EMS, 0.79 at 0.1% EMS, 0.65 at 0.15% EMS and 0.55 at 0.2% EMS. Similarly, Kumar *et al.* (2011) [20] observed striking differences not only in the callus induction but also in the weight of the calli. Pauli *et al.* (2019) [8] stated that EMS mutagens showed a significant impact on the callus growth by increasing EMS concentrations when compared to the untreated calli and resulted in callus necrosis at high EMS concentrations (Koch *et al.*, 2012) [19].

The relative differentiation rate is calculated for fixation of optimal dose for maximum heritable variation with minimum lethality. Among the various EMS treatments, the Kavuni embryogenic calli showed the relative differentiation rate of 100% in control followed by 78.57%, 45.71%, 21.42% and 15.51% at 0.05%, 0.1%, 0.15% and 0.2% EMS concentrations respectively.

Determination of LC_{50} of EMS treated calli

Newly proliferating embryonic calli was sensitive to the EMS treatment. The survival percentage of EMS treated embryonic calli decreased with increasing EMS concentrations. The LC_{50} value based on survival percentage was 0.10% (Table 2.) Ge

et al. (2015) reported the lethal concentration of embryonic calli of sweet orange as 0.15% EMS. Gadakh *et al.* (2015)^[10] reported LD₅₀ of sugarcane varieties CO 99004 and CO94012 embryonic calli as 0.05% EMS. The LD₅₀ in rice cell suspension culture was 0.4% (Chen *et al.*, 2013)^[6]. Dong *et al.* (2017) reported the optimal EMS concentration for rice varieties IR 231 and MR 219 as 0.05% and 0.06% respectively and showed high sensitivity to EMS.

Regeneration response of EMS treated calli

The regeneration percentage decreased with increase in EMS concentration. In EMS treated calli, the maximum regeneration percentage observed was 53.6% in control and minimum percentage was 6.6% at 0.2% EMS followed by 20% at 0.15% EMS, 26% at 0.1% EMS and 40% at 0.05% EMS as shown in Table 3. Similarly, Joong and Lee (2002) stated that at high EMS concentration the regeneration frequency was reduced. The abnormal plants with chlorophyll deficiency, weak culm and albino plants with no root formation were observed. Similar studies by Rakshana *et al.* (2019)^[32] reported the maximum regeneration frequency of 80% with average of 12 shoots per calli in NB media of 3mg/l 6-BA, 0.5mg/l kinetin, 3% maltose and 0.3 g/l glutamine. Venkateshan (2021)^[34] reported a reduced regeneration frequency of 5% at 0.2% EMS in Kalinga and 2% at 0.3% EMS in Kuliyadichan.

Plant height of *in vitro* EMS mutated Kavuni plantlets

Plant height is an important character in plant breeding improvement because it is closely associated with effective utilisation of assimilates to result in the formation of improved plant products. Mutants produce dwarf stature, semi dwarf stature, early maturity, increased tillering, high grain yield and other desirable traits.

In EMS treatment of embryogenic calli and subsequent regeneration, the plant height was observed as 22.5cm in control followed by 22.4cm in 0.15% EMS, 21.8cm in 0.2% EMS, 21.4cm in 0.1% EMS and 20.2cm in 0.05% EMS. Similar findings were reported by Nasri *et al.* (2021)^[27] in chrysanthemum cultivars treated with ethyl methane sulphonate of 0%, 0.125%, 0.25%, 0.5% that showed a novel variation in leaf colour and shape, plant height, days to flowering and inflorescence head size. Arici *et al.* (2018)^[3]

stated that *in vitro* culture of Alanson potatoes when treated with a different EMS concentrations of 20 mM, 50 mM, 75 mM and 100 mM resulted in the plant height of 7.65cm at 100 mM followed by 7.17 cm at 20mM.

Biochemical and nutritional evaluation of *in vitro* mutated plantlets of Kavuni

Protein and Carbohydrate content

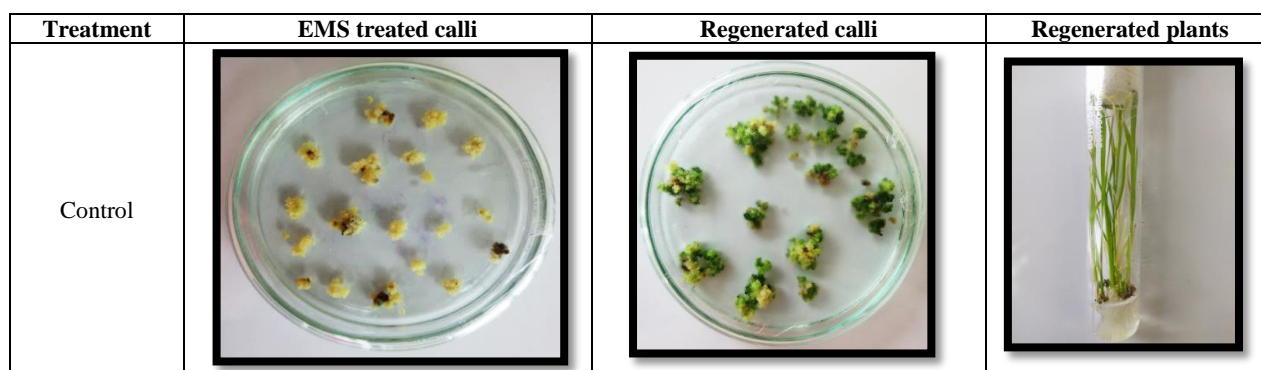
Among all the EMS treatments, the highest protein content was observed as 5.50% at 0.05% EMS followed by 5.14% at 0.2%, 5.02% at 0.1%, 4.56% in 0.15% and 4.22% in control. The carbohydrate content ranged from the maximum of 54.56% at 0.05% EMS and minimum of 48.22% in control followed by 49.06%, 51.94% and 52.78% at 0.15%, 0.1% and 0.2% EMS respectively. Gunasekaran *et al.* (2015) observed a reduced protein content of 32% at 0.2% EMS treatment when compared with control with a protein content of 34.58%.

β - Carotene, Chlorophyll and Anthocyanin content

The maximum β Carotene was observed as 0.02μg/g at 0.05% EMS and the minimum of 0.012 μg/g in 0.15% EMS followed by 0.014 μg/g in control, 0.015 μg/g at 0.1% and 0.017 μg/g at 0.2% EMS concentrations respectively. The maximum chlorophyll content was observed as 2.25 mg/g at 0.05% EMS and minimum of 1.54 mg/g at 0.15% EMS followed by 1.60 mg/g in control, 1.88 mg/g at 0.1% EMS and 2.02 mg/g at 0.2% EMS concentration. The maximum anthocyanin content was observed as 1.34 mg/g at 0.05% EMS and minimum of 1.21 mg/g at 0.15%EMS followed by 1.25 mg/g in control, 1.26 mg/g at 0.1% EMS and 1.30 mg/g at 0.2% EMS concentration respectively. Valarmathi *et al.* (2017)^[32] reported that EMS concentration of 0.3% increased chlorophyll and carbohydrate content and decreased in other EMS concentrations. According to Kumar *et al.* (2018)^[21] the photosynthetic pigments showed a significant variation in the EMS treated and control set of coriander.

Phenolics

The maximum phenolics observed was 28.5 mg/g at 0.05% EMS and minimum of 18.9 mg/g at 0.15% EMS followed by 20.2mg/g in control, 21.6 mg/g at 0.1% EMS and 24.4 mg/g at 0.2% EMS concentration. It is hold up with the results of Valarmathi *et al.* (2017)^[32].



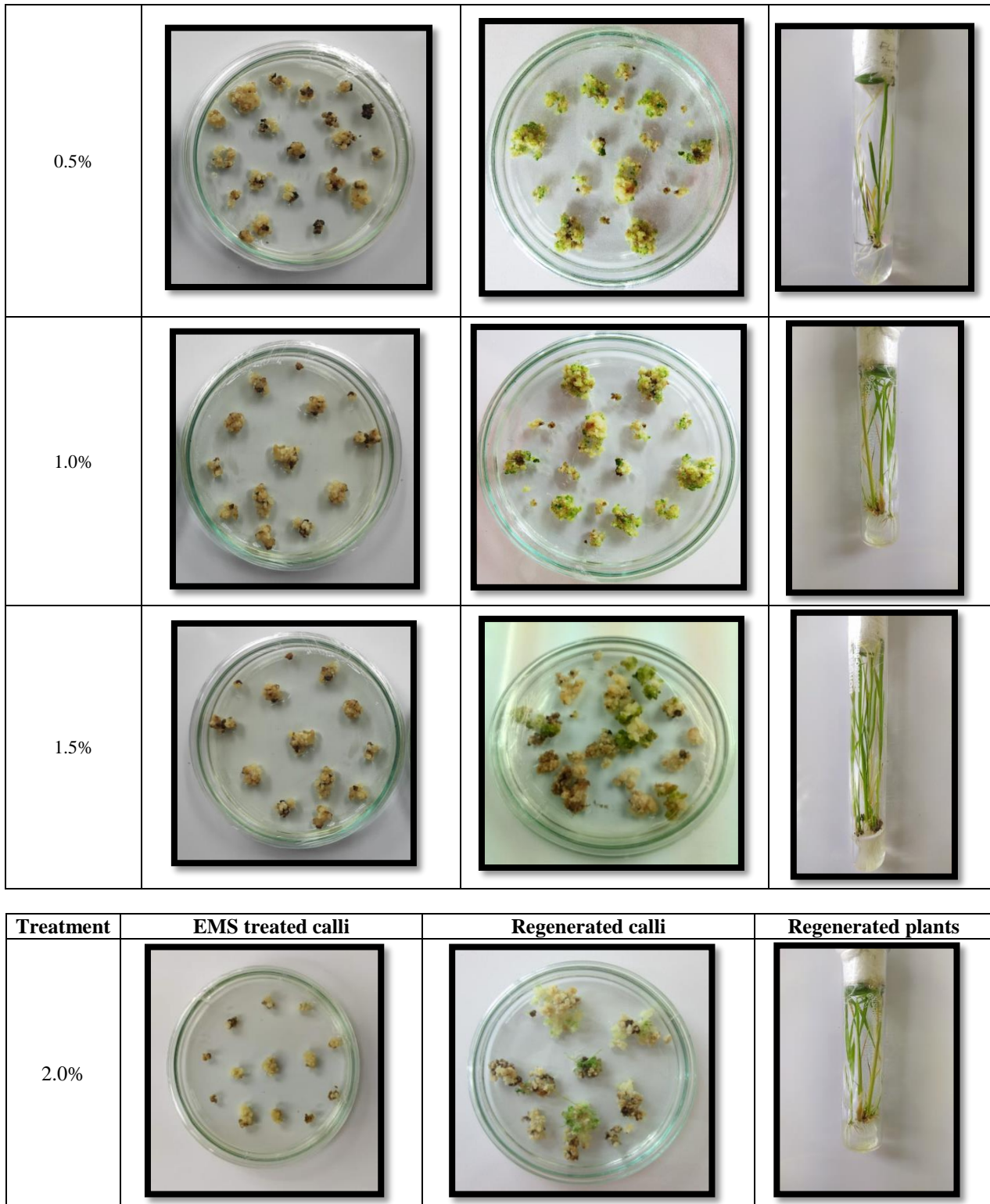


Table 1: Callus induction percentage in the rice cultivar Kavuni

Variety	No of seeds inoculated	Embryonic callus from seeds	Callus induction (%)
Kavuni	400	346	86.5

Table 2: Survival percentage, Relative growth rate, Relative differentiation rate and LC₅₀ value of EMS treated embryogenic calli.

EMS Concentration (%)	Survival percentage (%)	Relative growth rate RGR	Relative differentiation rate RDR (%)	LC ₅₀
Control	86.66	1.64±0.04 _a	100	0.10%
0.05%	50.00	0.86 ± 0.24 _{bc}	78.57	
0.1%	36.67	0.79 ± 0.05 _{cd}	45.71	
0.15%	26.68	0.65 ± 0.05 _d	21.42	
0.2%	13.33	0.55 ± 0.07 _c	15.51	

Relative growth rate values are analysed by DMRT Analysis. Data represented by Mean ± SE.

Table 3: Regeneration percentage of *in vitro* survived calli after EMS treatment in rice variety Kavuni

EMS treatment (%)	No. of calli inoculated	No. of calli that turned green	No. of calli that regenerated	Regeneration percentage (%)
Control	30	28	15	53.6%
0.05%	30	20	12	40.0%
0.1%	30	14	8	26.0%
0.15%	30	10	6	20.0%
0.2%	30	5	2	6.6%

Table 4: Morphological and biochemical parameters of *in vitro* EMS mutated Kavuni plantlets.

EMS treatments (%)	<i>In-vitro</i> Plantlet Height (cm)	Protein content (%)	Carbohydrate content (%)	β – carotene ($\mu\text{g/g}$)	Chlorophyll (mg/g)	Anthocyanin (mg/g)	Phenolics (mg/g)
Control	22.5	4.22	48.22	0.014	1.60	1.25	20.2
0.05%	20.2	5.50	54.56	0.020	2.25	1.34	28.5
0.1%	21.4	5.02	51.94	0.015	1.88	1.26	21.6
0.15%	22.4	4.56	49.06	0.012	1.54	1.21	18.9
0.2%	21.8	5.14	52.78	0.017	2.02	1.30	24.4

Conclusion

The overall study on the effect of EMS treatment on the embryonic calli decreased the survival percentage, relative growth rate and relative differentiation rate was decreased linearly with increasing concentration of EMS from 0.05% to 0.2%. The highest regeneration percentage was observed in at 0.05% EMS concentration. Further generation will be studied in order to select the stable mutant for dwarfism in rice variety, Kavuni.

References

- Acanda Y, Martínez Ó, Prado MJ, González MV, Rey M. EMS mutagenesis and PCR-HRM prescreening for point mutations in an embryogenic cell suspension of grapevine. *Plant cell reports* 2014;33(3):471-81.
- Amaravel M, Kumari SMP, Pillai MA, Saravanan S, Mini M, Binodh AK. Mass screening for salinity tolerance in rice (*Oryza sativa* L.) genotypes at early seedling stage by hydroponics. *Electronic Journal of Plant Breeding* 2019;10(1):137-42.
- Arici S, Tuncel Z, Zara A, Caltılı O. *in vitro* mutagenesis and selection of potato mutants resistance to fusarium dry root.
- Biswas A, Mandal AB. Plant regeneration in different genotypes of indica rice. 2007.
- Bolivar-González A, Valdez-Melara M, Gatica-Arias A. Responses of Arabica coffee (*Coffea arabica* L. var. Catuaí) cell suspensions to chemically induced mutagenesis and salinity stress under *in vitro* culture conditions. *in vitro Cellular & Developmental Biology-Plant* 2018;54(6):576-89.
- Chen YL, Liang HL, Ma XL, Lou SL, Xie YY, Liu ZL *et al.* An Efficient Rice Mutagenesis System Based on Suspension-Cultured Cells. *Journal of integrative plant biology* 2013;55(2):122-30.
- Das A, Datta AK, Bhattacharya A, Bhattacharyya A, Ghose S. EMS induced mutagenesis in Poshita and Jawahar 22 of *Withania somnifera* (L.) Dunal (Solanaceae). *Cytologia* 2010;75(3):305-11.
- Di Pauli V, Fontana P, Lewi D, Felipe A, Erazzú L. Somatic embryogenesis response in Argentinian sugarcane genotypes for *in vitro* mutagenesis application. Paper presented at the Proceedings of the International Society of Sugar Cane Technologists, 2019.
- Ekanyaka E, Weerakoon S, Silva T, Somaratne S. Induction of herbicide resistance via seed-derived rice (*Oryzasativa*) Calli. *IRA-International Journal of Applied Sciences* 2016;3(3):2455-4499.
- Gadakh S, Patel D, Patil A. Evaluation of sugarcane (*Saccharum spp. complex*) mutants for yield, yield contributing traits and quality parameters. *International Journal of Advanced Biological Research* 2015;5:220-28.
- Ge H, Li Y, Fu H, Long G, Luo L, Li R. Production of sweet orange somaclones tolerant to citrus canker disease by *in vitro* mutagenesis with EMS. *Plant Cell, Tissue and Organ Culture (PCTOC)* 2015;123(1):29-38.
- Gunasekaran A, Pavadai P. Studies on induced physical and chemical mutagenesis in groundnut (*Arachis hypogea*). *International Letters of Natural Sciences* 2015.
- Hodge J, Hofreiter B. *Methods in Carbohydrate Chemistry* (eds Whistler, RL and Be Miller, JN) Academic Press New York, 1962.
- Jain SM. Mutagenesis in crop improvement under the climate change. *Romanian biotechnological letters* 2010;15(2):88-106.
- Juturu V, Mekala GK, Garladinne M, Reddy P, Sekhar A. Optimization of *in vitro* regeneration protocol for a popular indica rice (*Oryza sativa* L. cv Swarna). *Ann. Plant Sci* 2016;2:1395-401.
- Khaleda L, Al-Forkan M. Stimulatory effects of casein hydrolysate and proline in *in vitro* callus induction and plant regeneration from five deep water rice (*Oryza sativa* L.). *Biotechnology* 2006;5(3):379-84.
- Khalil F, Naiyan X, Tayyab M, Pinghua C. "Screening of EMS-induced drought-tolerant sugarcane mutants employing physiological, molecular and enzymatic approaches. *Agronomy* 2018;8(10):226.
- Khush GS. What it will take to feed 5.0 billion rice consumers in 2030. *Plant molecular biology* 2005;59(1):1-6.
- Koch AC, Ramgareeb S, Rutherford RS, Snyman SJ, Watt MP. An *in vitro* mutagenesis protocol for the production of sugarcane tolerant to the herbicide imazapyr. *in vitro Cellular & Developmental Biology-Plant* 2012;48(4):417-27.
- Kumar B, Mistry N, Singh B, Gandhi CP. Indian horticulture database. National Horticulture Board. Ministry of Agriculture, Government of India, 2011.
- Kumar G, Pandey A. Ethyl methane sulphonate induced changes in cyto-morphological and biochemical aspects of *Coriandrum sativum* L. *Journal of the Saudi Society of Agricultural Sciences* 2018;18(4):469-75.
- Lee JH, Lee SY. Selection of stable mutants from cultured rice anthers treated with ethyl methane sulfonic

- acid. *Plant cell, tissue and organ culture* 2002;71(2):165-71.
23. Lowry OH, Rosebrough NJ, Farr AL, Randall RJ. "Protein measurement with the Folin phenol reagent. *Journal of biological chemistry* 1951;193:265-75.
 24. Luan YS, Zhang J, Gao XR, An LJ. Mutation induced by ethylmethanesulphonate (EMS), *in vitro* screening for salt tolerance and plant regeneration of sweet potato (*Ipomoea batatas* L.). *Plant cell, tissue and organ culture* 2007;88(1):77-81.
 25. Mori M, Tomita C, Sugimoto K, Hasegawa M, Hayashi N, Dubouzet JG *et al.* Isolation and molecular characterization of a Spotted leaf 18 mutant by modified activation-tagging in rice. *Plant molecular biology* 2007;63(6):847-60.
 26. Murashige T, Skoog F. A revised medium for rapid growth and bio assays with tobacco tissue cultures. *Physiologia plantarum* 1962;15(3):473-97.
 27. Nasri F, Zakizadeh H, Vafaei Y, Mozafari AA. *In vitro* mutagenesis of *Chrysanthemum morifolium* cultivars using ethylmethanesulphonate (EMS) and mutation assessment by ISSR and IRAP markers. *Plant Cell, Tissue and Organ*
 28. Paul S, Roychoudhury A. Comparative analyses of regeneration potentiality of eight Indigenous Aromatic Indica rice (*Oryza sativa* L.) Varieties. *Int. J Sci. Res. in Biological Sciences* 2019;6:1.
 29. Pedro AC, Granato D, Rosso ND. Extraction of anthocyanins and polyphenols from black rice (*Oryza sativa* L.) by modeling and assessing their reversibility and stability. *Food Chemistry* 2016;191:12-20.
 30. Penna S, Vitthal SB, Yadav PV. *In vitro* mutagenesis and selection in plant tissue cultures and their prospects for crop improvement. *Bioremediation, Biodiversity, Bioavailability* 2012;6:6-14.
 31. Purwanto E, Nahdhiana Z, Yunindanova M. Yield and anthocyanin content of M1 generation of black rice induced by gamma rays. Paper presented at the IOP Conference Series: Materials Science and Engineering, 2019.
 32. Rakshana P, Valarmathi R, Raveendran M. Optimization of tissue culture protocol for rapid regeneration of traditional therapeutic rice genotype 'Kavuni'. *Electronic Journal of Plant Breeding* 2017;10(2):334-40.
 33. Valarmathi R, Raveendran M, Robin S, Senthil N. Unraveling the nutritional and therapeutic properties of 'Kavuni' a traditional rice variety of Tamil Nadu. *Journal of Plant Biochemistry and Biotechnology* 2015;24(3):305-15.
 34. Venkateshan R. Improvement of rice cultivars, Kalinga and ADT 43 for salt tolerance by *in vitro* mutation 2021.