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Prevalence of major pathogenic bacteria from mastitis buffaloes

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Abstract

The study was carried out at the animal farms of the ICAR- Central Institute of Research on Buffaloes (ICAR-CIRB), Hisar across two breeds during winter season to compare the efficacy of infrared thermography with different diagnostic techniques for early detection of mastitis. The two organized herds included the Murrah breed animals (n=104) at the main campus located at Hisar, Haryana and the Nilli-Ravi herd (n=100) at ICAR-CIRB sub campus located at Nabha, Punjab. CMT was used as initial classifier for animal as normal/non-mastitis, subclinical/clinical mastitis. Milk samples from thirty-eight mastitis cases were examined for the isolation and identification of bacterial agents. Among bacterial agents isolated from thirty-eight mastitis cases, prevalence of Coagulase-negative Staphylococci was 65.79%, mixed infection was 21.05%, S. aureus and Escherichia coli was 5.26% each and Streptococcus spp. was 2.63%. Prevalence of subclinical mastitis in herd of CIRB was 10.29% and Clinical mastitis was 8.33%.

Keywords: Infrared thermography, CMT, mastitis, coagulase-negative staphylococci, S. aureus

Introduction

Animal husbandry is an integral component of agriculture sector and plays a multifaceted role in uplifting Indian economy. It provides employment and empowerment to rural population and acts as moving bank during financial crisis. Livestock provides sustained income to small and landless farmers throughout the year, thus contributing in their socio-economic development. Livestock sector contributes 4.4 percent to GDP and 25.6 percent of total agricultural GDP (*National Accounts Statistics-2016; Central Statistical Organisation; GoI*) ^[15]. India possesses world's largest livestock population and largest producer of milk but still not self-sufficient in milk. Milk production during 2017-18 and 2018-19 is 176.3 million tonnes and 187.7 million tonnes respectively showing an annual growth of 6.47%. (Annual report 2019-20, DAHD & F, GOI) ^[1] This growth is mostly the result of an increase in numbers of producing animals rather than a rise in productivity per head. Low productivity is due to poor animal health, inferior management practices and ineffective diagnostic techniques. Contagious disease affects animal health and productivity.

Worldwide most frequently occurring calamitous disease of dairy herd is mastitis (Ojo *et al.* 2009) ^[16] causing major economic losses to the farmers. Disease causes inflammation of affected mammary gland. The major pathogens of both contagious and environmental mastitis are; *Staphylococcus aureus*, *Streptococcus agalactiae*, *Streptococcus uberis*, *E. coli*, and *Klebsiella*, causes fever, inflammation, swelling, milk composition and color changes, and presence of somatic cells etc. Intensity of the inflammation can be classified into sub-clinical, clinical and chronic forms. Inflammation might differ in severity due to many factors such as pathogen type, animal health status, age and lactation cycle of the animal. In addition, mastitis being a potent zoonotic disease, poses serious hazard to human health (Blum *et al.* 2008)^[2].

Impacts of mastitis

Reduction or complete loss in milk production affects farmer's income. Milk has to be discarded for 3 days during treatment and 3 days post treatment for withholding period (Huijps, Lam and Hogeveen, 2008)^[10]. The quality of milk is deteriorated due to elevated somatic cell count and decreased milk fat. Antibiotic treatment and veterinary care impose additional financial burden on livestock holders. The minimised organoleptic properties affects the selling price of milk and milk products. The meat of mastitis affected animals fetch lower prices due to reduced quality and carcass yield. Mastitis affected animals are culled and replaced with healthy stock.

The premature replacement of stock largely contributes to economic losses (Halasa *et al.*, 2007; de Graves and Featrow, 1993; Hortet and Seegers, 1998)^[8, 5, 9].

Experimental setup

The study was carried out at the animal farms of the ICAR-Central Institute of Research on Buffaloes (ICAR-CIRB), Hisar across two breeds during winter season. The two organized herds included the Murrah breed animals (n=104) at the main campus located at Hisar, Haryana and the Nilli-Ravi herd (n=100) at ICAR-CIRB sub campus located at Nabha, Punjab. All animals studied were maintained as per standard practices of ICAR minimum feeding standards with ad-libtum drinking water availability round the clock. All buffaloes studied were milked twice daily.

Milk sample collection

CMT was used as initial classifier for animal as normal/nonmastitis, subclinical/clinical mastitis. Composite and quarterwise (10ml each) milk samples in sterile polypropylene tubes of all animals were collected and preserved for bacterial isolation and identification. During collection, udder of each animal was properly disinfected with 70% alcohol and dried before sample collection. First few streaks of milk were discarded and milk sample from each teat was collected in sterile test tubes and properly labelled with buffalo's identification number and teat position (i.e. fore-right, fore-left, hind-right and hind-left).

Bacteriological Culture Examination

Milk sample from each quarter (approx. 0.1 ml of milk) was inoculated using sterile cotton swab in a zigzag pattern onto the surface of blood agar, MacConkey Lactose agar, Mannitol salt agar and incubated at 37^oC for 24 hours under aerobic conditions. The isolates were identified by their cultural characteristics, microscopic examination in their Gram stained slides, catalase test, and oxidase test. Identification and characterization of bacteria was performed as per the method described by Cowan and Steel, 1970.

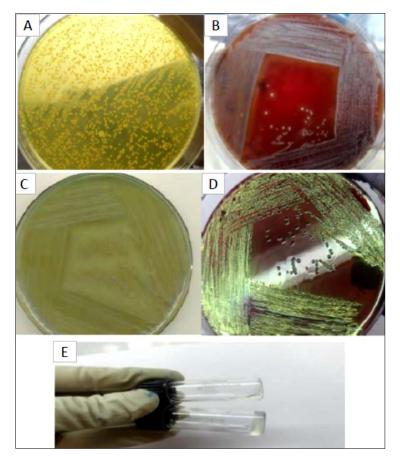


Fig 1: A. Primary isolation of *Staphylococcus* on Mannitol salt agar. Presumptive *S. aureus* ferments mannitol and produces yellow colored colonies B. Presumptive isolation of *Streptococcus spp.* on 5% bovine blood agar C. Green pigmentation of presumptive *Pseudomonas* colonies on nutrient agar D. Metallic sheen of *E. Coli* on Eosin Methylene Blue agar E. Tube coagulase test using rabbit plasma to confirm coagulase-positive *S. aureus*.

Results and Discussion

Milk samples from thirty-eight mastitis mastitis cases were examined for the isolation and identification of bacterial agents. Out of Milk samples from 38 mastitis cases, 08 samples were found containing mixed infection (*Coagulase-negative Staphylococci + Escherichia coli -2, Coagulase-negative Staphylococci + pseudomonas spp.-2, Coagulase-negative Staphylococci + Streptococcus spp.-2, S. aureus + Escherichia coli -1, S. aureus + Streptococcus spp.-1) and the rest 30 samples were associated with single infection.*

Mastitis can be epidemiologically categorized into contagious and environmental mastitis caused by a wide spectrum of pathogens. Increase in the level of humidity and pollution in the environment of the barn also increases the load of bacterial pathogens in the environment. In present study among bacterial agents isolated from 38 mastitis cases, prevalence of *Coagulase negative Staphylococcus spp.* was 65.79%, mixed infection was 21.05%, *S. aureus* and *E. coli* was 5.26% each and *Streptococcus spp.* was 2.63%. Prevalence of *Coagulase negative Staphylococcus spp.* was found highest in present study. The results were alike with Zeryehun and Abera, (2017)^[20] as their study revealed bacteriological examination of milk samples where Coagulase negative Staphylococcus species (CNS) (34.2%) was the predominant species while Streptococcus faecalis (2.1%) was identified as the least bacteria. Prevalence of mastitis particularly the subclinical mastitis was major problem likewise Coagulase negative Staphylococcus species was observed as a main causative agent in mastitis (Moroni et al., 2006; Saber *et al.*, 2017; Dhakal and Nagahata 2018; Boireau *et al.*, 2018)^[13, 17, 6, 3]. Prevalence of subclinical mastitis was found more in present study as compared to clinical mastitis which was in agreement with the results of Isaea and Kurtu (2018) ^[11]. Present study results were in close resemblance with Mostafa et al. (2018)^[14] who observed comparatively higher prevalence of Coagulase negative Staphylococci (CNS) (19.5%) followed by Escherichia coli (8.3%) and Streptococcus spp. (5.6%) in mastitis milk samples similarly very less prevalence of E. coli (6.7%) in mastitis was reported by Ferreira et al. (2018). Very less prevalence of Streptococcus spp. (2.4%) was also detected by Kasa et al. (2020) ^[12] in bovine clinical mastitis. The results were differing with the findings of Waseem et al. (2020)^[19] as they observed highest prevalence of Staphylococcus spp. in clinical cases of Bovine mastitis. Correspondingly Staphylococcus aureus was the most frequently isolated bacterial species as per the findings of Srinivasan et al. (2013) followed by Streptococcus agalactiae and Coagulase negative Staphylococcus species.

 Table 1: Prevalence of bacterial agents isolated from thirty-eight mastitis cases

Pathogen	N=38	Prevalence Rate	
Coagulase-negative Staphylococci	25	65.79%	
Staphylococcus aureus	2	5.26%	
Streptococcus spp.	1	2.63%	
Escherichia coli	2	5.26%	
Mixed infection	8	21.05%	
Table 2. Details of heatenial agents acusing mixed infections			

Table 2: Details of bacterial agents causing mixed infections

Mixed Infections	
Coagulase-negative Staphylococci + Escherichia coli	2
Coagulase-negative Staphylococci + Pseudomonas spp.	2
Coagulase-negative Staphylococci + Streptococcus spp.	2
Staphylococcus aureus + Escherichia coli	1
Staphylococcus aureus + Streptococcus spp.	1

 Table 3: Prevalence of bacterial agents isolated from mastitis cases of Nilli-Ravi buffaloes

Pathogen	N=16	Prevalence Rate
Coagulase-negative Staphylococci	10	62.50%
Staphylococcus aureus	01	6.25%
Streptococcus spp.	01	6.25%
Escherichia coli	01	6.25%
Mixed infection	03	18.75%

 Table 4: Details of bacterial agents causing mixed infection in mastitis cases of Nilli-Ravi buffaloes

	Mixed Infections	
	Coagulase-negative Staphylococci + Escherichia coli	02
C	Coagulase-negative Staphylococci + Streptococcus spp.	01

 Table 5: Prevalence of bacterial agents isolated from mastitis cases of Murrah buffaloes

Pathogen	N=22 Prevalence Rate	
Coagulase-negative Staphylococci	15	68.18%
Staphylococcus aureus	01	4.54%
Escherichia coli	01	4.54%
Mixed infection	05	22.72%

 Table 6: Details of bacterial agents causing mixed infections in mastitis cases of Murrah buffaloes

Mixed Infections		
Coagulase-negative Staphylococci + Pseudomonas spp.	02	
Coagulase-negative Staphylococci + Streptococcus spp.	01	
Staphylococcus aureus + Escherichia coli	01	
Staphylococcus aureus + Streptococcus spp.	01	

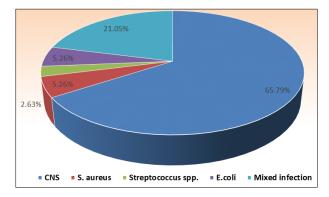


Fig 2: Prevalence rate of bacterial agents isolated from 38 mastitis cases

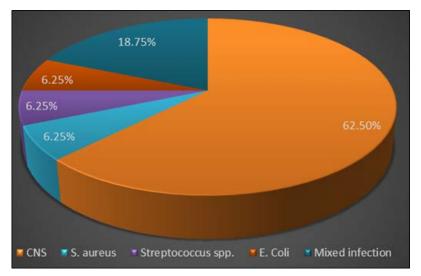


Fig 3: Prevalence rate of bacterial agents isolated from mastitis cases of Nilli-Ravi buffaloes

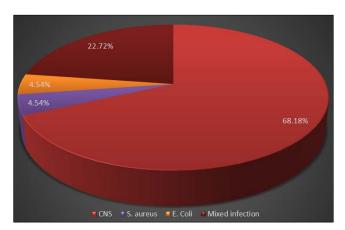


Fig 4: Prevalence rate of bacterial agents isolated from mastitis cases of Murrah buffaloes

References

- 1. Annual report 2019-20, DAHD & F, GOI.
- Blum S, Heller ED, Krifucks O. Identification of a bovine mastitis Escherichia coli. Vet. Microbiol 2008;132:135-48.
- Boireau C, Cazeau G, Jarrige N, Calavas D, Madec JY, Leblond A *et al.* Antimicrobial resistance in bacteria isolated from mastitis in dairy cattle in France, 2006-2016. J Dairy Sci 2018;101(10):9451-9462.
- 4. Cowan and Steel's Manual for the Identification of Medical Bacteria. J Clin. Pathol 1993;46(10):975.
- De Graves FJ, Featrow J. Economics of mastitis and mastitis control. Vet. Clin. North Am. Food Anim. Pract 1993;9(3):421-433.
- 6. Dhakal IP, Nagahata H. Evaluation of Mastitis Related Measures & Their Applications to Classify Buffalo Milk in Chitwan, Nepal. Sci. Technol 2018;8:99-111.
- Ferreira JC, Gomes MS, Bonsaglia ECR, Canisso IF, Garrett EF, Stewart JL. Comparative analysis of four commercial on-farm culture methods to identify bacteria associated with clinical mastitis in dairy cattle. PLoS ONE 2018;13(3):e0194211.
- Halasa T, Huijps K, Osteras O, Hogeveen H. Economic effects of bovine mastitis and mastitis management: A review. Veterinary Quarterly 2007;29(1):18-31.
- Hortet P, Seegers H. Calculated milk production losses associated with elevated somatic cell counts in dairy cows: review and critical discussion. Vet. Res 1998;29:497-510.
- 10. Huijps K, Lam TJGM, Hogeveen H. Cost of mastitis: facts and perception. J Dairy Res 2008;75:113-120.
- Isaea AA, Kurtu MY. Mastitis and its Effect on Chemical Composition of Milk in and around Worabe Town, Am. Sci. Res. J Eng., Technol., Sci 2018;42(1):210-220.
- 12. Kasa G, Tegegne B, Tadesse B. Isolation and Identification of Major Pathogenic Bacteria from Clinical Mastitis Cows in Asella Town, Ethiopia. Vet. Med. Int 2020;1:6.
- Moroni P, Sgoifo RC, Pisoni G, Castiglioni B, Boettcher PJ. Relationships between somatic cell count and intramammary infection in buffaloes. J Dairy Sci 2006;89:998-1003.
- 14. Mostafa A, Gamil A, Zeedan SG, Abdoula H, Zeina A. Isolation and identification of bacteria causing mastitis in small ruminants and their susceptibility to antibiotics, honey, essential oils, and plant extracts. Vet. World 2018;11(3):355-362.

- 15. National Accounts Statistics-2016. Central Statistical Organisation; GoI.
- Ojo OE, Oyekunle MA, Ogunleye AO, Otesile EB. Escherichia coli, O157:H7 in food animals in part of South-Western Nigeria: Prevalence and *in vitro* antimicrobial susceptibility. Trop. Vet 2009;26(3):23-30.
- 17. Saber AA, Hassan AM, Nabtiti ASE, Hassan AM, Mansour SR. Evaluation of field techniques to diagnose early subclinical mastitis in relation to hygiene score in a buffalo farm. CATRINA 2017;16(1):53-60.
- Srinivasan P, Jagadeswaran D, Manoharan R, Giri T, Balasubramaniam GA, Balachandran P. Prevalence and Etiology of Subclinical Mastitis among Buffaloes (Bubalus bubalus) in Namakkal, India. Pak. J Biol. Sci 2013;16:1776-1780.
- Waseem R, Muhee A, Malik HU, Akhoon ZA, Munir K, Nabi SU, Taifa S. Isolation and Identification of Major Mastitis Causing Bacteria from Clinical Cases of Bovine Mastitis in Kashmir Valley. Indian J Anim. Res 2020;54:1428-1432.
- 20. Zeryehun T, Abera G. Prevalence and Bacterial Isolates of Mastitis in Dairy Farms in Selected Districts of Eastern Harrarghe Zone, Eastern Ethiopia. J Vet. Med 2017;1:7.