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Variability in cultural and morphological characters of different isolates of *Helminthosporium sativum*

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Abstract

Leaf blight symptom collected from different location on various wheat cultivars showed remarkable variation. Symptoms incited by *Helminthosporium sativum* on wheat cultivars varied from light yellow to light orange. The maximum colony diameter was recorded in case of isolate H_5 (66.56mm) and minimum in case of H_3 (48.38mm).Regarding dry mycelial weight maximum dry weight was recorded in isolate H_4 (160.60 mg.) and minimum in H₃ (128.25mg.). The maximum number of spores were recorded in isolate H_5 (45.33000/ml.) and minimum in H₃. The purpose of this study to see the variations among the isolates, collected from different locations.

Keywords: cultural, morphological characters, Helminthosporium sativum

Introduction

Wheat is one of the most cereal crop grown worldwide and liking staples of nearly 2.5 billion of world population. India is the second largest producer of wheat worldwide (29.58mh and global area 14% and production 99.70mt.) reported by Sharma and Sendhil (2015 and 2016)^[8, 9]. Leaf blight of wheat caused by *Helminthosporium sativum* is a serious problem for wheat growers. The leaf blight, which used to pose a serious problem in the eastern regions of the country under warm and humid weather conditions, have become far and wide in the country. A number of pathogens causing leaf spot, blight and blotch on wheat crop has been reported in India. Among them *H. sativum* Pammel, King and Bakke is more important than *Alternaria triticina* (Singh *et al.* 1992) ^[10]. Variability in cultural characteristics was observed in morphology and growth rate. However, there was no relationship between morphological variability and virulence amongst isolates (Oliveira *et al.*1998) ^[6]. Mitra (1931) ^[5] described the variations in the growth features of two strain of *Bipolaris sorokiniana* obtained from wheat and barley on four types of media.

Materials and Methods

The leaves collected from infected wheat plant show blight symptoms from CRC, GBPUA&T, Pantnagar and from NDUA&T, Faizabad, Uttar Pradesh. Samples were done on Potato Dextrose Agar (PDA) medium. The pathogen was grown on PDA medium. The composition of PDA was as: Peeled potato- 200 gram, Dextrose 20gram, Agar-agar - 20gram and Distilled Water - 1000ml. The blighted spots were cut into small pieces of half a centimeter in size. These pieces were sterilized with mercuricm chloride solution (1:1000) for one minute followed by 2-3 washing with sterilized distilled water. The pieces were later dried using sterilized blotters and plated on the PDA on sterilized plastic Petri dishes (90 mm diameter) @ 5 pieces per plate at equal distance.

These petridishes were incubated at 25 ± 1 ⁰C in BOD incubator. After 48 hours of incubation, the growing mycelium from the margins of the apparently distinct colonies were sub cultured on fresh PDA slant.

Purification and Maintenance of the Culture: The culture of the fungus isolates were purified by Single spore isolation technique and maintained on PDA at low temperature $(5\pm1^{\circ}C)$ in the refrigerator. To keep the culture bible, sub culturing was done at an internal of 15-20 days (Duveiller and Altamirano, 2000)^[4]. The re-culture on as per need.

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Result and Discussion Cultural characters Growth on PDA

H. sativum isolates were grown on PDA for six days and growth was recorded on 3^{rd} and 6^{th} days of inoculation. The growth of isolates after 3^{rd} and 6^{th} days of inoculation was 39.69 and 74.99 mm, respectively and these were significantly different (Table-1). Comparison among

individual isolates of H. sativum indicated that isolates H_5 had highest growth of 66.56 mm and was significantly superior over others. It was followed by isolate H_4 (60.33 mm), H_2 (57.33 mm) and H. (54.16 mm). The lowest radial growth was recorded in isolate H_3 (48.38 mm) which was significantly lower than all the other isolates. Isolates H_3 , H_4 and H_5 showed significant differences among themselves however, H, and H2 were statistically at par.



Fig 1: Variability in colony character and conidia of different isolates of Helminthosporium sativum

 Table 1: Colony diameter of different isolates of Helminthosporium

 sativum

| Colony diameter (mm)* after | | | | |
|-----------------------------|-------|--------|-------|--|
| Isolates | 3 day | 6 days | Mean | |
| H_1 | 35.00 | 73.33 | 54.16 | |
| H_2 | 44.33 | 70.33 | 57.33 | |
| H ₃ | 30.11 | 66.66 | 48.38 | |
| H_4 | 40.66 | 80.00 | 60.33 | |
| H ₅ | 48.33 | 84.00 | 66.56 | |
| Mean | 39.69 | 74.99 | 57.34 | |

Cd₁ at 5% 3.26 (interval), Cd₂ at 5% 1.16 (Isolates), Cd₃ at 5% 7.30 (D x I) CV = 7.47

Growth in liquid medium

Isolates were grown in liquid medium (PD broth) and dry mycelial weight was recorded after 8,16 and 24 days of inoculation. Average mycelial dry weight at different intervals showed that the growth of isolates increased with the time, recording 111.98, 142.12 and 171.50 mg, respectively. These values were statistically significant from each other. The results are given in Table 2.

 Table 2: Mycelial dry weight of different isolates of *H. sativum* in liquid medium

| | Dry weight (mg) | | | | | |
|----------------|-----------------|---------|---------|--------|--|--|
| Isolates | 8 day | 16 days | 24 Days | Mean | | |
| H_1 | 100.63 | 172.35 | 159.76 | 144.25 | | |
| H ₂ | 103.75 | 133.73 | 163.03 | 133.50 | | |
| H ₃ | 135.05 | 111.64 | 138.06 | 128.25 | | |
| H_4 | 129.49 | 161.84 | 190.46 | 160.60 | | |
| H ₅ | 90.98 | 131.03 | 206.20 | 142.74 | | |
| Mean | 111.98 | 142.12 | 171.50 | 141.87 | | |

 CD_1 (Days intervals) at 5% 2.37, CD_2 (Isolates) at 5% 3.06 CD_3 (DXI) at 5% 5.31

CV 8.24 *Mean of three replications.

Dry weight of mycelium ranged from 160.60 mg in isolate H_4 to 128.25 mg in H_3 . Isolate H_4 had significantly higher mycelial dry weight than all the other isolates which was followed by H_1 and H_5 giving mycelial dry weight yield of 144.25 and 142.74 mg, respectively. Isolates H_1 , H_2 and H_3 with mycelial dry weight of 144.25, 133.50 and 128.25 mg,

respectively had significant differences among themselves whereas isolate H_1 (144.25) and H_5 (142.74) were at par. The isolate H_2 H_3 and H_4 showed significant differences among themselves.

The growth of isolates recorded at first observation i.e. after 8 days of inoculation indicated that the highest dry weight was in isolate H_3 (135.05 mg) and the lowest (90.98mg) in isolate H_5 . All the isolates had significantly different growth except isolate1 and H_2 which were at par.

At the second observation i.e. after 16 days of inoculation the highest mycelial dry weight of 172.35 mg was recorded in isolate H_1 and the lowest (111.64 mg) in isolate H_3 . All the isolates had statistically significant differences among themselves except H_2 and H_5 which were statistically at par.

Third observation was taken after 24 days of inoculation, the isolates showing significant differences among themselves except H (159.76 mg) and H₂ (163.03 mg). Isolate H₁ was found to record highest (206.20mg) mycelial yield followed by the isolate H₄ (190.46 mg). The lowest (138.06mg) mycelial dry weight was recorded by H₃ isolate.

Sporulation

Isolates were grown on PDA and number of spores were counted after 5 and 10 days of inoculation. The observations are presented in Table 3.

The average sporulation of the isolates on 5^{th} and 10^{th} day of inoculation was found to be 29×10^3 and 49.53×10^3 spores/ml which were significantly different.

Table 3: Sporulation of Different isolates of *H. sativum* on PDA

| Number of spores (000/ml)* after | | | | |
|----------------------------------|-------|---------|-------|--|
| Isolates | 5 day | 10 days | Mean | |
| H_1 | 31.66 | 43.33 | 37.50 | |
| H ₂ | 26.33 | 50.66 | 38.50 | |
| H_3 | 23.33 | 50.00 | 36.66 | |
| H_4 | 29.00 | 47.66 | 38.33 | |
| H_5 | 34.55 | 56.00 | 45.33 | |
| Mean | 29.00 | 49.53 | 39.26 | |
| | | | | |

CD at 5% 2.36 (interval), CD₂ at 5% 3.73 (isolates), CD₃ at 5% 5.28 (D XI) CV 7.90 * Mean of three replications

Comparison among isolates indicated that H_5 produced maximum spores (45.33x10³ spores/ml) which were significantly superior to all the other isolates. Number of spore produced by the isolates H_1 , H_2 , H_3 and H_5 were statistically at par.

Interaction between isolates and interval showed that at the first observation i.e. 5 days after inoculation, the highest number of spores (34.55 10^3) was produced by isolate H₅ which was significantly superior over that of all the other isolates. Isolates H₃, H₄ and H₁ had significant differences among themselves whereas H₂ and H₃ were statistically at par. At the time of second observation, the highest number of spores (56.00 x 10^3) was found in isolate H₅ and this was also significantly superior over others. The lowest number of spores (43.33 x 10^3) was produced by isolate H₁ which was at par with H₄ (47.66 x 10^3). However, isolate H₄ was at par with isolates H₃ and H₂ as well. Bidari and Govindu (1976) ^[2] also reported to marked differences in sporulation of the pathogen. Valim *et al.* (1997) ^[11] also studied the variation in cultural

characters that is growth rate, colour, presence of sectors and textures among ten wheat isolates of *Bipolaris sorokiniana*.

Morphological characters

Growth rate of the pathogen based on colony diameter was medium (H₁, H₂ H₃ and H₄) to fast (H₅) with growth ranging between 39.69 to 74.99 mm. Colony shape was irregular (H₁ and H₅) to circular (H₂, H₃ and H₄) with smooth margin (H₁ and H₅) to rough margin (H₁, H₂, and H₃) without zonation, texture was thick-fluffy in H₁ and thick to thick dense in all the isolates other than H₁ Light yellow pigment produced by H₁ H₂ and H4, Pale yellow by H₃ and light orange by isolate H₅. The colour of colony was blackish gray (H₁ and H₂), shooty black (H₃ and H₅) and grayish black (H₄). Conidia cylindrical in isolates H₅ to ellipsoid straight to curved in H₁, H₂, H₃ and H₄ measuring 45.62 x 56.15 - 17.70 x 19.90 µm (Table 4 and Plate A, B and C). The similar observation were also recorded by Chauhan *et al.* (2017) ^[3] and Mitra (1931) ^[5].

|--|

| Characters | \mathbf{H}_{1} | \mathbf{H}_2 | H_3 | H_4 | H_5 |
|-------------------|---------------------------------------|------------------------------|------------------------------|------------------------------|-----------------------------|
| Growth Characters | Medium | Medium | Medium | Medium | Fast |
| Shape | Irregular | Circular | Circular | Circular | Irregular |
| Margin | Scattered | Wavy | Smooth | Wavy | Smooth |
| Colour | Blackish Gray | Blackish Gray | Shooty black | Blackish Black | Shooty black |
| Texture | Thick Fluffy | Thick Dense | Thick dense | Thick Dense | Thick |
| Pigmentation | Light Yellow | Light Yellow | Pale Yellow | Light Yellow | Light Orange |
| Conidial Size | $45\text{-}62\times17.967\mu\text{m}$ | $46.98 \times 17.72 \ \mu m$ | $56.10 \times 19.89 \ \mu m$ | $52.30 \times 19.25 \ \mu m$ | $25.13 \times 8.62 \ \mu m$ |
| Conidial shape | Ellipsoid | Ellipsoid | Ellipsoid Straight | Ellipsoid Curved | Cylindrical |

The present study, variations in cultural characters agreement with the finding of Pascual and Raymundo (1995)^[7] where cultural characters were exhibited by 20 isolates grown on three media *viz*. PDA, V-8 juice agar and wheat extract agar. Such variations in morphological characters were also reported by Oliveira *et al.* (1998)^[6]. This experiments was resembles with findings of Ahmed *et al.* (1997)^[1] in colour

variation of isolates.

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