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Role of salicylic acid in postharvest management of peach and plum fruits: A review

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Abstract

Salicylic acid (SA) is a naturally occurring phenolic compound that is widely distributed in plants and it is considered as hormone due to its regulatory role in plants. SA has attracted considerable interest because of its role in modulating plant responses to biotic and abiotic stresses during postharvest storage of fruits. SA has been shown to exhibit a high potential in delaying ripening, maintaining fruit quality, and controlling postharvest losses. Use of SA could be a promising technology for enhancing postharvest storability and retaining the nutritional quality of peach and plum fruits after harvest.

Keywords: Postharvest management, peach, plum, salicylic acid

Introduction

Most fruits are perishable due to their physiological properties and cause significant postharvest loss in developing countries with limited storage options. According to Kader (2005) ^[1], one-third of all fresh fruits wasted before they reach to customers. The pre-harvest factors such as crop growing environment (weather and climate), tree conditions, light penetration, training pruning, irrigation, nutrient management, insect pest and disease management, use of growth regulators etc. influence overall fruit quality and suitability for storage by modifying physiology, chemical composition, and morphology of fruits. Besides preharvest factors, the main causes associated with the postharvest losses of fruits are poor management of handling and packaging, and storage environment conditions (Kitinoja, et al., 2011)^[2]. Due to soft texture of fruits, they are easily bruised or wounded as a result of harvesting, and other postharvest handling operation such as packaging, transportation and during storage. Mechanically damaged fruits are more susceptible to pathogen attacks thus fruit decay during long distance transport, storage and retail environment causes high losses besides deterioration in fruit quality. The quality of fruits after harvest is largely governed by water content, respiratory rate, ethylene production, endogenous hormones in fruit and exogenous factors such as microbial growth, temperature, relative humidity and atmospheric compositions. Therefore, post-harvest loss of fruits needs to be reduced and their shelf life increased considerably by careful manipulation of pre and postharvest factors associated with the losses. The loss can be reduced by adopting suitable growing environment, implementing the adequate cultural methods, careful harvesting, handling, packaging, transportation and storage. The use of appropriate chemicals at pre and postharvest stage may prolong the availability of fresh produce for a long period of time by protecting them from pathogens and other harmful environmental factors. Also controlled atmosphere storage has been found to be effective in minimising rate of fruit quality deterioration.

Various synthetic chemical compounds have been used for extending the postharvest life and retaining the quality of fruit after harvest. However, concerns about chemicals pesticides being used for postharvest management of fruits are arising among consumers during recent past decades because of their negative consequences on human health as well as environment and also prompted the researchers to look for better fungicide alternatives (Babalar, *et al.*, 2007) ^[3]. The use of natural compounds in postharvest quality management of fruits has been gained attention during the past few decades. Among others, salicylic acid is one of the natural compounds reported to be beneficial in maintaining the quality of several fruits during storage (Romani, *et al.*, 1989; Zeng *et al.*, 2006; Wang *et al.*, 2006; Mo *et al.*, 2008; Shafiee *et al.*, 2009)^[4-8].

Salicylic acid: A Plant hormone with diversified role in plants

Salicylic acid and its derivatives, as one of the plant hormones produced by the plant, belong to the phenolic acids group and are made up of a ring attached to the hydroxyl and carboxyl groups. It is cosmically spread in plant kingdom (Raskin et al., 1990)^[9] that has been considered as a powerful plant hormone and plays a variety of regulatory actions in plant metabolism (Raskin, 1992)^[10]. Because of the key role of salicylic acid in photosynthesis, plant water relations, various enzyme activities, and its effect on plants exposed to various biotic and abiotic stresses, exogenous application of salicylic acid shown to be beneficial to crop growth and biological productivity under constantly changing environmental conditions (Senaratna et al., 2000, Horvath et al., 2007; Hayat et al., 2010) [11-13]. Salicylic acid has recently attained a lot of attention because it regulates the expression of different modalities in plant responses to biological activities (Malamy and Klessig, 1992; Ryals et al., 1996; Shah et al., 2005) [14-16] and abiotic stressors (Hayat et al., 2010; Ding et al., 2001, Ding at al., 2003) [13, 17-18].

Role of salicylic acid in postharvest management of fruits

Salicylic acid and its derivatives are widely in use to enhance postharvest life of fruits by modifying physiological processes and alleviating the deleterious effects of various biotic and abiotic stresses in fruits. Fruits are exposed to various pathogen attacks after harvest too. To defend themselves from infections plants activating defence mechanisms such as locally acquired resistance (LAR) and systemic acquired resistance (SAR) (Vlot et al., 2009) [19]. One of the processes that protect the host from pathogens is the formation of huge levels of salicylic acid. Salicylic acid and its derivatives are considered an important regulator of disease resistance in plants (Klessig and Malamy, 1994) [20]. Salicylic acid is involved in the signal transduction pathway that leads to systemic acquired resistance (Ryals et al., 1996; Murphy et al., 1999) [15,21], stimulates a number of defence genes, most notably those encoding pathogenesis-related (PR) proteins (Carr and Klessig, 1989; Bowles, 1990)^[22,23], and several of these PR proteins have antimicrobial properties, viz. chitinase or 1,3-glucanase activity (Schlumbaum et al., 1986; Mauch et al., 1988; Salzman, et al., 1998) [24-26], and have been demonstrated in vitro to suppress the growth of oomycete pathogens and true fungi (Niderman, et al., 1995)^[27]. Thus, the exogenous administration of salicylic acid and its derivatives plays a significant role in plant signal transduction and disease resistance, as acetylsalicylic acid promotes the expression of pathogenic genes and confers pathogen resistance (Morris et al., 1998; Pasquer et al., 2005; Makandar et al., 2006) [28-30].

It has been demonstrated that salicylic acid maintain flesh firmness of harvested fruits (Wang *et al.*, 2006; Li and Han, 1999; Yan *et al.*, 1998) ^[6, 31-32] thereby reducing the fruit quality deterioration during storage. Exogenous salicylic acid's action is influenced by a variety of parameters, including plant type and developmental stage, salicylic acid application method and concentration, and endogenous salicylic acid content in plants under specific environmental conditions (Horvath *et al.*, 2007) ^[12].

Salicylic acid in postharvest management of peach fruits

Peaches are perishable fruits; they ripen rapidly at ambient temperature and require careful and adequate handling after harvest to avoid serious losses. The shelf life of peaches is often limited due to the damage caused by the cold. Susceptible varieties after refrigerated storage (<10 °C for 2-3 weeks) showing damage symptoms of textural changes and discoloration of the flesh besides rotting especially brown rot and transit rot infections; suitable management practices needed for minimizing the postharvest losses. Salicylic acid considerable role in regulating physiological activities that associated with postharvest losses of fruit. Postharvest Salicylic acid treatment of 'Okuba' peaches @ 0.10 g L⁻¹ inhibit the respiration rate and delay the ethylene production peak of ripening (Han et al., 2002) [33]. Although, SAtreatment exhibit higher cell membrane electrolyte leakage of fruit during the early days, but lower at the subsequent days (Han et al., 2002)^[33]. In an immersion treatment with @ SA 0, 0.35, 0.7 and 1 mM to the peach fruits (cv. Beijing 24) fruit at commercial maturity for 5 min and stored at 0 °C for 28 days, then moved to 20 °C for 3 days; Wang et al. (2006) [6] observed that only 1 mM SA significantly maintained higher firmness and lower chilling injury index, decay index, and thiobarbituric acid-reactive-substance content of fruit compared with the control. Further, they also studied whether 1 mM SA alleviated chilling injury by influencing antioxidant systems and/or heat shock proteins of the fruit wherein they observed reduced-to-oxidized ascorbate ratio (AsA/DHAsA) in 1 mM SA treated fruit and alleviated the cold storage induced reduction of a number of volatiles and sugars, and thereby maintained flavor quality of fruit during shelf life after cold storage. According to Awad (2013) [34], SA treatment of 'Flordaprince' peaches maintained a significantly lower enzymes activity (peroxidase and polyphenol oxidase) with increasing SA concentrations. SA treatment also increases phenols, flavonoids, and ascorbic acid contents leading to fruits with higher DPPH and FRAP radicals scavenging capacity with higher antioxidant enzymes catalase, ascorbate peroxidase and superoxide dismutase activity (Razavi et al., 2017) [35]. Peach cv. 'Flordaking' fruit treated with 4 and 6 mM SA exhibited delayed and reduced ethylene production as compared to untreated fruit during fruit ripening, reduced activities of fruit softening enzymes esterase, endo-1-4 β-Glucanase, endoincluding pectin and exo-polygalacturonase and reduced ethylene production as compared to untreated fruit during ripening at ambient conditions (Ullah et al., 2020)^[36].

Postharvest application of salicylic acid has been found effective in maintaining fruits quality during storage. A decreased weight loss of 'Maria Delicia' peaches and lower total soluble solids but a higher titrable acidity and ascorbic acid reported with increased SA concentrations from 0.5 mM to 2 mM compared to untreated control (Abbasi *et al.*, 2010)^[37]. In 'Flordaking' peaches, Tareen *et al.* (2012)^[38] observed that SA @ 2.0 mmol L⁻¹ significantly less weight loss compared to other SA doses (0, 0.5, 1.0 and 1.5 mmol L⁻¹). Awad (2013)^[34] recorded highest values of fruit losses in weight decreasing SA concentrations from 1.5 to 0.5 mM, and fruits receiving SA treatments produced the firmest fruits.

Salicylic acid application effectively reduce fungal decay (Khademi *et al.*, 2013) ^[39], rate of spoilage and loss of physiological weight total and helpful in obtaining higher fruit firmness, maintenance of fruit TSS and titrable acidity during storage (Awad, 2013; Razavi *et al.*, 2014, Razavi *et al.*, 2017) ^[34, 40, 35]. Among different postharvest application of SA (0, 2, 4 or 6 mm) to Flordaking peaches fruits at ambient conditions (25 ± 2 ⁰C and 60-65% RH), 6 mM SA exhibited lowest weight

loss of fruits, TSS and highest fruit firmness (Ullah *et al.*, 2020) ^[36]. SA treatment (1 mM) also effective in alleviation of the development of flesh browning, storage-induced reduction of a number of volatiles and sugars and softening thereby maintained fruit flavor quality of 'Hujingmilu' peach during shelf-life after cold storage Yang *et al.* (2020) ^[41].

Pre-harvest SA treatments @ 2 mM in 'Cresthaven' peaches found beneficial for better fruit characteristics (fruit weight, fruit flesh firmness, total antioxidant content, total phenol content and acidity) and decreasing the loss of fruit firmness at harvest and after storage at 2 °C for 8 days and 2 days post storage shelf life at 20 °C (Deniz and Imail, 2020)^[42]. Ali at al. (2021)^[43] reported that the pre-harvest application of SA @ 3 mM at fruit development stages can improve the harvest quality and storability of 'Flordaking' peaches during low temperature storage. They used different concentrations of SA (0, 1, 2 and 3 mM) sprayed on the plants at three growth stages of fruit, i.e., the cell division, cell enlargement and pithardening stages and observed that all the SA treatments improved the fruit quality at harvest and maintained higher levels of flesh firmness, titratable acidity and ascorbic acid during storage; however, fruit weight loss, soluble solid contents, membrane leakage, chilling injury, colour development, disease and decay incidence and the climacteric peak of ethylene were lowered by SA treatment after six weeks of low-temperature storage.

Salicylic acid in postharvest management of plum fruits

Plum fruits are perishable and fruit softens quickly after harvest, resulting in a shorter shelf life at ambient temperatures (Menniti et al., 2004) [44]. Refrigeration effectively slows fruit ripening and extends the life of plums after harvest, but the beneficial effects may be limited by the development of hypothermia-related diseases, including internal blackening, translucent pulp and/or redness (Crisosto et al., 2004; Manganaris, et al., 2007) [45, 46]. Chilling injury, disease incidence, electrolyte leakage, MDA content, respiration and ethylene production of control fruit increases during cold storage, therefore, delaying or reducing pulp softening and spoilage at low temperatures should be an important strategy for extending shelf life and maintaining plum fruit quality (Luo et al., 2011) [47]. According to Shaaban et al. (2020) [48], Fruit weight loss, total soluble solids, total carotenoids content and total phenolics content) increases with advances in storage period while the fruit firmness, lightness (L*), hue angle (h°) of colour, titratable acidity, total flavonoids content and antioxidant activity decreases. Salicylic said to augment the activities of SOD and CAT and maintain lower levels of cell membrane permeability and MDA; thereby, it inhibitory effect on the occurrence of cell membrane lipid peroxidation (Wang et al. 2020) [40].

In a preharvest SA spray treatments, Martínez-Esplá *et al.*, 2017) ^[49] observed enhanced activity of the antioxidant enzymes catalase (CAT), peroxidase (POX), superoxide dismutase (SOD), and ascorbate peroxidase (APX) and higher levels of antioxidant enzymes at the time of harvest in salicylate-treated fruits. Davarynejad *et al.* (2015) ^[50] evaluated SA dip (0, 1, 2, 3 and 4 mmol/L for 5 min) to the 'Santa Rosa' plum fruits at the mature ripe stage, packed in boxes with polyethylene covers and stored at 4 ^oC with 95% relative humidity for 25 days. They observed that weight loss and softening of the plum fruits were decreased significantly by the use of salicylic acid and found to be effective in

maintaining titratable acidity, ascorbic acid, total phenolics and antioxidant activity in plum fruits during storage. Martínez-Esplá, *et al.* (2017) ^[49] also applied foliar spray of SA, acetylsalicylic acid (ASA), and methylsalicylate (MeSA) treatments and advocated that preharvest treatment with salicylates could be a safe, eco-friendly, and new tool to improve and maintain plum quality attributes and especially their content of antioxidant compounds, with an additional effect on delaying the postharvest ripening process through increasing the levels of antioxidant compounds and the activity of the antioxidant enzymes.

In 'Qadri' plum, salicylic acid (0, 3, and 6 mmols.1-1) treatment increased titratable acidity, hardness, and moisture content, while water loss, significantly anthocyanin, and total phenols were reduced in fruits stored in a cold room at $2 \pm 1^{\circ}C$ and 90% relative humidity for 60 days. 6 mmols 1⁻¹ salicylic acid (Kasnazany et al., 2017) ^[51]. In a foliar sprays combinations of SA (3 and 4 mmol/L), putrescine (3 and 4 mmol/L) and moringa leaf extract (5 and 10%) and harvested fruits storage at 0 °C with 85-90% RH for 8 week, the SA @ 3mmol/ L, putrescine @ 4 3mmol/ L and moringa leaf extract @ 10% treatment found to be more effective than other treatments in decreasing the weight loss, softening and maintaining titratable acidity, total carotenoids, total phenolics, total flavonoids and antioxidant activity in plum fruits during storage at 0 °C; hence, preharvest treatment of plum fruits with salicylic acid, putrescine and moringa leaf extract can be effective in delaying the ripening processes and of commercial use to extend the storage life of postharvest plum fruits with acceptable fruit quality (Shaaban *et el.* 2020)

Salicylic acid treatment of 'Qingnai' plum fruit said to be useful in suppression of chilling injury and delayed onset of the climacteric peak of respiration besides inhibition of ethylene production during cold storage (Lau et al., 2011)^[47]. In aqueous dip SA treatments (0.5, 1.0, 1.5 and 2.0 mM) of Santa Rosa plum for 10 min at 20°C, air-dried and packed in plastic punnets followed by storage at 2°C and 90±5% RH, SA (2.0 mM) to be the best for maintaining the fruit quality and overall acceptability, fruit colour; hence, SA dip can be integrated as an effective postharvest practice in the supply chain management to extend their storage life while maintaining fruit quality during storage for 36 days (Sharma and Sharma, 2017) ^[52]. Salicylic acid also play vital role in cold resistance of postharvest plum fruits and its relationship with ethylene signal as the appropriate concentration of exogenous SA treatment of 'Bulang' plum (1.0 mmol L-1) flesh browning and mitigate chilling injury symptoms in fruits stored at 4 °C (Wang et al., 2020)^[53].

References

- 1. Kader AA. Increasing food availability by reducing postharvest losses of fresh produce. Acta Horticulturae. 2005;682;2169–2175.
- 2. Kitinoja L, Saran S, Roy SK, Kader AA. Postharvest technology for developing countries: challenges and opportunities in research, outreach and advocacy. Journal of Science of Food of Agriculture. 2011;9:597-603.
- 3. Babalar M, Asghari M, Talaei A, Khosroshahi A. Effect of pre and postharvest salicylic acid treatment on ethylene production, fungal decay and overall quality of Selva strawberry fruit. Food Chemistry. 2007;105:449-453.
- 4. Romani RJ, Hess BM, Leslie CA. Salicylic acid

inhibition of ethylene production by apple discs and other plant tissues. Journal of Plant Growth Regulator. 1989;8:63-69.

- Zeng KF, Cao JK, Jiang WB. Enhancing disease resistance in harvested mango (*Mangifera indica* L. cv. Matisu) fruit by salicylic acid. Journal of the Science of Food and Agriculture. 2006;86:694-698. DOI:
- Wang L, Chen S, Kong W, Li S, Archbold DD. Salicylic acid pre-treatment alleviates chilling injury and affects the antioxidant system and heat shock proteins of peaches during cold storage. Postharvest Biology and Technology. 2006;41:244-251.
- Mo Y, Gong D, Liang G, Han R, Xie J, Li W. Enhanced preservation effects of sugar apple fruits by salicylic acid treatment during postharvest storage. Journal of the Science of Food and Agriculture. 2008;88;2693-2699.
- 8. Shafiee M, Taghavi TS, Babalar M. Addition of salicylic acid to nutrient solution combined with postharvest treatments (hot water, salicylic acid, and calcium dipping) improved postharvest fruit quality of strawberry. Scientia Horticulturae. 2009;124:40-45.
- 9. Raskin I. Salicylate, a new plant hormone. Plant Physiology. 1992;99: 799-803.
- Raskin IH, Skubatz, W, Tang, Meeuse BJD. Salicylic acid levels in thermogenic and non-thermogenic plants. Annals of Botany. 1990;66:376-383.
- 11. Senaratna T, Touchell D, Bunn E, Dixon K. Acetyl salicylic acid (aspirin) and salicylic acid induce multiple stress tolerance in bean and tomato plants. Plant Growth Regulators. 2000;30:157-161.
- 12. Horvath E, Szalai S, Janda T. Induction of abiotic stress tolerance by salicylic acid signaling. Journal of Plant Growth Regulators. 2007;26:290-300.
- Hayat Q, Hayat H, Irfan M, Ahmad A. Effect of exogenous salicylic acid under changing environment: A review. Environmental and Experimental Botany. 2010;68:14-25.
- 14. Malamy J, Klessig DF. Salicylic acid and plant disease resistance. Plant Journal. 1992;2:643-654.
- 15. Ryals JA, Neuenschwander UH, Willits MG, Molina A, Steiner HY, Hunt MD. Systemic acquired resistance. Plant Cell. 1996;8:1809-1819.
- 16. Shah J. Lipids, lipases and lipid-modifying enzymes in plant disease resistance. Annual Reviews of Phytopathology. 2005;43:229-260.
- Ding CK, Wang CY, Gross KC, Smith DL. Reduction of chilling injury and transcript accumulation of heat shock protein genes in tomatoes by methyl jasmonate and methyl salicylate. Plant Science. 2001;161:1153-1159. DOI:https://doi.org/10.1016/S0168-9452(01)00521-0.
- Ding C, K, Wang C Y. The dual effects of methyl salicylate on ripening and expression of ethylene biosynthetic genes in tomato fruit. Plant Science. 2003;164:589-596.
- 19. Vlot AC, Dempsey DA, Klessig DF. Salicylic acid, a multifaceted hormone to combat disease. Annual Review of Phytopathology. 2009;47:177-206..
- 20. Klessig DF, Malamy J. The salicylic acid signal in plants. Plant Molecular Biology. 1994;26:1439-1458.
- 21. Murphy IS, Chi vasa S, Singh DP, Car JP. Salicylic acidinduced resistance to viruses and other pathogens: a parting of the ways? Trends in Plant Science. 1999;4:155-160.
- 22. Carr JP, Klessig DF. The pathogenesis-related proteins of

plants. In: Setlow J, Editors. Genetic engineering: principles and methods. New York: Plenum Press; 1989.

- 23. Bowles DJ. Defense-related proteins in higher plants. Annual Review of Biochemistry. 1990;59:873-907.
- 24. Schlumbaum A, Mauch F, Vogeli U, Boller T. Plant chattiness are potent inhibitors of fungal growth. Nature. 324:365-367.
- 25. Mauch F, Hadwiger LA, Boller T. Antifungal hydrolases in pea tissue 1. Purification and characterization of 2 chitinases and 2 beta-1,3-glucanases differentially regulated during development and in response to fungal infection. Plant Physiology. 1988;87:325-333.
- 26. Salzman NT, Genetet I, Bruyere T, Gees R, Stintzi A, Legrand M, *et al.* Pathogenesis related PR1 proteins are antifungal isolation and characterization of 314kilodalton proteins of tomato and of a basic PR-1 of tobacco with inhibitory activity against Phytophthora infestans. Plant Physiology. 1995;108:17-27.
- 27. Salzman RA, Tikhonova I, Bordelon BP, Hasegawa PM, Bressan RA. Coordinate accumulation of antifungal proteins and hexoses constitutes a developmentally controlled defense response during fruit ripening in grapes. Plant Physiology. 1998;117:465-472.
- 28. Morris SW, Vernooij B, Titatarn S, *et al.* Induced resistance responses in maize. Molecular Plant-Microbe Interactions. 1998;11:643-658.
- 29. Pasquer F, Isidore E, Zarn J, Keller B. Specific patterns of changes in wheat gene expression after treatment with three antifungal compounds. Plant Molecular Biology. 2005;57:693-707.
- Makandar R, Essig JS, Schapaugh MA, Trick HN, Shah J. Genetically engineered resistance to Fusarium head blight in wheat by expression of Arabidopsis NPR1. Molecular Plant-Microbe Interactions. 2006;19:123-129.
- 31. Li LP, Han T. The effects of salicylic acid in the storage of peach. Food Science. 1999;7:61-63.
- 32. Yan T, Shen QG, Liu CD. Effects of salicylic acid (SA) on ripening fruits. Chinese Bulletin of Botany. 1998;15:61-64.
- 33. Han T, Wang Y, Li L, Ge X. Effect of exogenous salicylic acid on post-harvest physiology of peaches. In: XXVI International Horticultural Congress: Issues and Advances in Postharvest Horticulture. 2002;628: 583-589.
- 34. Awad RM. Effect of post-harvest salicylic acid treatments on fruit quality of peach cv. "Flordaprince" during cold storage. Australian Journal of Basic and Applied Sciences. 2013;7(7):920-927.
- 35. Razavi F, Hajilou J, Aghdam MS. Salicylic acid treatment of peach trees maintains nutritional quality of fruits during cold storage. Advances in Horticultural Science. 2017;32(1):33-40.
- 36. Ullah S, Malik MA, Khan AS, Akhtar G, Hafiz Faried N, Razzaq K, *et al.* Exogenous application of salicylic acid influences fruit softening and quality of 'flordaking' peach during ripening at ambient conditions. Pure and Applied Biology. 2020;9(1):1009-1024.
- Abbasi NA, Hafeez S, Tareen, MJ. Salicylic acid prolongs shelf life and improves quality of 'maria Delicia' peach fruit. Acta Horticulturae. 2010;880:191-198.
- Tareen MJ, Abbasi NA, Hafiz IA. Effect of salicylic acid treatments on storage life of peach fruits CV. 'Flordaking'. Pakistan Journal of Botany. 2012;44(1):119-124.

- Khademi Z, Ershadi A. Postharvest Application of Salicylic Acid Improves Storability of Peach (*Prunus persica* cv. Elberta) Fruits. International Journal of Agriculture and Crop Sciences. 2013;5(6): 651-655.
- 40. Razavi F. Hajilou J, Dehgan G, Nagshi R, Hassani B, Turch N. Enhancement of postharvest quality of peach fruit by salicylic acid treatment. International Journal of Biosciences. 2014;4(1):177-184.
- Yang C, Duan W, Xie K, Ren C, Zhu C, Chen K, Zhang. Effect of salicylic acid treatment on sensory quality, flavor-related chemicals and gene expression in peach fruit after cold storage. Postharvest Biology and Technology. 2020;161:111089. [DOI: https://doi.org/10.1016/j.postharvbio.2019.111089]
- 42. Deniz E, İsmail O. Effect of pre-harvest salicylic acid treatments on the quality and shelf life of the 'Cresthaven' peach cultivar Folia Horticulturae. 2020;32(2):221-227.
- 43. Ali I, Wang X, Tareen MJ, Wattoo FM, Qayyum A, Hassan MU, *et al.* Foliar Application of Salicylic Acid at Different Phenological Stages of Peach Fruit CV. 'Flordaking' Improves Harvest Quality and Reduces Chilling Injury during Low Temperature Storage. Plants 2021;10(10):1981. [DOI: https://doi.org/10.3390/plants10101981]
- 44. Menniti A, Gregori MR, and Donati I. 1-Methylcyclopropene retards postharvest softening of plums. Postharvest Biology and Technology. 2004;31:269-275.
- 45. Crisosto CH, Garner D, Crisosto GM, Bowerman E. Increasing 'Blackamber' plum (*Prunus salicina* Lindell) consumer acceptance. Postharvest Biology and Technology. 2004;34:237-244.
- 46. Manganaris GA, Vicente AR, Crisosto CH, Labavitch JM. Effect of dips in a 1-methylcyclopropene-generating solution on 'Harrow Sun' plums stored under different temperature regimes. Journal of Agricultural and Food Chemistry. 2007;55:7015-7020.
- 47. Luo Z, Chen C, Xie J. Effect of salicylic acid treatment on alleviating postharvest chilling injury of 'Qingnai' plum fruit. Postharvest Biology and Technology. 62(2):115-120.
- 48. Shaaban FKM, Gehan AM, El-Hadidy, Mahmoud TSM. Effects of salicylic acid, putrescine and moring leaf extract application on storability, quality attributes and bioactive compounds of plum cv. 'Golden Japan': Effects of salicylic acid, putrescine and moring leaf extract application. Future of Food: Journal on Food, Agriculture and Society. 2020;8(2):1-14.
- 49. Martínez-Esplá A, Serrano M, Valero D, Martínez-Romero D, Castillo S, Zapata PJ. Enhancement of antioxidant systems and storability of two plum cultivars by preharvest treatments with salicylates. International Journal of Molecular Sciences. 2017;6:18(9):1911. DOI: https://doi.org/10.3390/ijms18091911]
- Davarynejad GH, Zarei M, Nasrabadi ME, Ardakani E. Effects of salicylic acid and putrescine on storability, quality attributes and antioxidant activity of plum cv. 'Santa Rosa'. J Food Science and Technology 2015;52(4):2053-62.
- Kasnazany SAS, Fakhraddin M, Hama Salieh H, Aljabary AMO. Effect of salicylic acid and potassium metabisulfite on postharvest quality of plum cv. Qadri. Euphrates Journal of Agriculture Science. 2017;9(3):79-

91.

- Sharma S, Shamra RR. Effect of salicylic acid treatment on fruit quality of Japanese plum (*Prunus salicina*) cv. Santa Rosa. Indian Journal of Agricultural Sciences. 2017;87(9):1209-1213.
- 53. Wang Y, Zhang Y, Yang HY. Ethylene signals involved in salicylic acid to improve cold resistance of postharvest plum fruits (J). Science and Technology of Food Industry. 2020:41(5):262-267.