



ISSN (E): 2277-7695
ISSN (P): 2349-8242
NAAS Rating: 5.23
TPI 2022; 11(11): 1949-1961
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www.thepharmajournal.com

Received: 26-08-2022
Accepted: 29-09-2022

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Efficacy of different eco-friendly methods against late blight of potato, *Phytophthora infestans*: A review

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Abstract

One of the most dramatic episodes induced by a plant pathogen in human history was the late blight of the potato caused by the fungus *Phytophthora infestans*, which was responsible for the famine in Ireland (1845). Famine in Ireland claimed the lives of one million people. Therefore, it is essential to achieve eco-friendly methods potato late blight disease management. Due to the rapid spread of late blight outbreaks in recent years, management efforts have solely focused on the application of fungicides. Fungicide use, however, poses a major risk to both the environment and public health when used carelessly. Additionally, it causes the pathogen to develop a resistance and has a negative impact on helpful species including nitrogen fixers, resident antagonists and mycorrhizal fungi. Therefore, environmentally friendly methods for managing late blight are needed as a top priority in order to reduce the usage of fungicides. Most recently, Late blight fungal isolates have undergone considerable modifications, including variations in how aggressive they are toward the crop. Since late blight affects the entire community, all producers, farmers, gardeners and growers must implement an efficient, environmentally friendly management strategy with the assistance of governmental organizations, extension specialists, crop consultants, etc. The first line of defense in this situation is disease management through cultural practices and the efficient, effective and environmentally friendly management of late blight of potatoes depends on forecasting system, physiological strategies, biological control, host plant resistance and bio-technological approach.

Keywords: *Phytophthora infestans*, pathogen, biological control, antagonists and eco-friendly

1. Introduction

Late blight caused by the oomycete pathogen *Phytophthora infestans*, disease of major concern all over the world is costing huge losses every year in potato (Kumaon *et al.*, 2009) [84]. Reports of complete field destruction due to late blight epidemics are relatively common. The fungus is responsible for global annual crop loss of US \$ 12 billion. Yield loss due to late blight in India varies from year to year and range from 20 to 75% (Sundaresha *et al.*, 2015) [145]. The pathogen *Phytophthora infestans* is highly variable and has sophisticated weaponry including effectors molecules coded by avirulence genes that allow rapid infection and host tissue colonization (Kamoun and Smart, 2005) [85]. Once inside host tissue, a complex set of compounds such as metalloproteinase, cutinase and other protein with no identification function required for cell killing and nutrient uptake are promptly activated (Lee *et al.*, 2006) [100]. There are ten avirulence proteins are known to be involved in pathogenesis and act as effectors that are delivered inside the plant cell (Haldar *et al.*, 2006) [72]. These protease inhibitors produce by the pathogen to disturb the defense mechanism of the host plant. New class of protease inhibitors were described in *Phytophthora infestans* (Tian *et al.*, 2007) [147]. For more than a decade, controlling late blight has become increasingly demanding because of the emergence of new strains of the pathogen, some new strains are known to be more aggressive and resistant to the fungicide metalaxyl (Daayf *et al.*, 2001) [38]. Moreover, indiscriminate use of systemic fungicides especially metalaxyl (ridomil) provides chance to develop resistant strain of the fungus has been reported from home and abroad (Singh, 2000) [138]. The term alternative can have multiple meanings and its interpretation varies accordingly. In this context, Cultural control is the first line of defense which includes change in date of planting, intercropping, nutrient management, irrigation management *etc.* For eco-friendly disease management, forecasting system plays important role so that preventive measures can be taken up if there is likelihood of disease appearance. In the last decades, growers and researchers have made impressive strides in implementing successful local epidemic

forecasting systems and guided chemical pesticide management regimes for efficient PLB control in the field (Poudel *et al.* 2020) ^[115]. Physiological strategies *i.e.* escape and resistance is very important to cope up with disease. In general, due to the different modes of actions (*i.e.* antagonistic effects or induction of plant defense mechanisms), the use of microorganisms as biological control agents has a definite potential. Currently potato varieties with fully late blight resistance are under serious study in breeding programme. A new sexually reproducing potato late blight (*Phytophthora infestans*) population characterised by early oospore-derived epidemics is one of the most severe threats to organic potato production (Lehtinen and Hannukkala, 2004) ^[101]. In some growth areas, up to eight pesticide applications per growth season is required for sufficient control, presenting both heavy economic and environmental costs associated with potato production (Naumann *et al.*, 2020) ^[108]. Innovative and effective control measures are needed if fungicide use is to be reduced or eliminated as in case of organic potato production. These so-called effector molecules could function either within or outside of host plant cells, participating in diverse physiological processes such as nutrient uptake, plant cell wall degradation and host defense signaling interference (He *et al.* 2018; Du *et al.* 2021) ^[75, 44]. Protectants are capable of preventing *P. infestans* infection by interfering with spore germination and/or initial penetration of plant surface processes (Lamichhane *et al.* 2018) ^[98].

This technology has been recently applied to clone *R* genes from wild wheat species (Arora *et al.* 2019) ^[10], as well as genome-wide curation of *R* genes across 18 *Solanum* species (Seong *et al.* 2020) ^[121] and diverse *A. thaliana* accessions (Van de Weyer *et al.* 2019) ^[153].

Thus, they may represent an interesting tool for the development of novel concepts in disease management. Even biotechnology is also being employed in the pursuit of late blight resistance. Fully resistant genetically engineered strains are expected soon. In this review, we will try to use eco-friendly approaches that include non-fungicidal strategies such as cultural method, forecasting system, physiological strategies, biological control, host plant resistance and biotechnological approach for management of late blight disease.

2. Pathogen

In 1861, Anton de Bary experimentally established that the fungus was the cause of plant disease known as late blight of potato, a disease that closely resembles the downy mildew. In 1875, de Bary studied it in detail and gave name the pathogen to *Phytophthora infestans* which means infectious plant destroyer (Turner, 2005; Widmark, 2010) ^[151, 164]. *Phytophthora* belongs to the kingdom Stramenopila, a group of micro-organism that is closely related to brown algae, golden brown algae and diatoms. This statement supported the references (Forster *et al.*, 1990; Dick, 2001; Kamoun and Nusbaum, 2009; De Bary, 1876; Widmark, 2010) ^[54, 43, 84, 40, 164]. Anton de Bary also showed that late disease could easily appear on fungal spore dusted on potato plants.

2.1 Pathogen variability

Important aspects of monitoring population structure of *Phytophthora infestans* should be considered for thorough analysis and proper development of alternative procedures aiming at controlling the late blight in potato. Even it has been on the agenda of scientific community and techniques have

become available. Biological markers including mating types, race pattern, metalaxyl sensitivity and ploidy are the potential means for monitoring variability in *Phytophthora infestans*. The occurrence of sexual reproduction also affects the population dynamics of the pathogen. Two contrasting scenarios for genetic structure of population of *Phytophthora infestans* can be envisioned: (1) A clonal population structure with limited number of clonal lineages (Eduardo *et al.*, 2007) ^[47]. High genotypic diversity of pathogen can provide greater potential for local adaptation to the changing environment which makes disease management more difficult (Sujkowski *et al.*, 1994) ^[144]. (2) A panmictic population is comprised of a large number of distinct genotypes of *P. infestans*. In areas where population is clonal, the risk of new diversity being introduced grows as seed trade becomes more globalized.

2.2 Physiological races

Fungus *Phytophthora infestans* is considered as highly variable due to the frequent appearance of its new pathogenic virulent types in field and often observed in the laboratory. Giddings and Berg, 1919; Berg, 1926) ^[65, 14] were pioneers in detecting variations in the *Phytophthora infestans* populations. After almost seven years of introduction of resistant hybrids/cultivars having *R* genes, pathological specializations (races) within potato isolates were reported by Schick (1932) ^[120]. In 1958, race 0 and race 1 were prevalent, the former being predominant in North-western hills. In 1965, race 3 and 4 appeared for first time (Dutt, 1965) ^[46]. In 1969, race 0 was still common race (58%) but frequency of race 1 increased considerably (42%) over the years. Two gene complex races recorded in 1971 attacked the late blight resistant varieties *i.e.* Kufri Jyoti, K. Jeevan (Phadtare *et al.*, 1973) ^[113]. Thirteen new races were identified by 1975 in Shimla hills. By 1978, the complex races started appearing more frequently. Absence of race 0 and predominance of race 3, 4, 7, 8, 10, 11 indicated the shift in the virulence of *Phytophthora infestans* was almost complete. This statement supported the references (Bhattacharyya and Singh, 1986; CPRI, 1981 and 1987; Rivera-Pena, 1995; Cristinzio *et al.*, 1998; CPRI, 2013; CPRI, 1999; Shanmugam, 2001) ^[15, 30, 31, 117, 35, 33, 127]. However, universal appearance of races or, at least their detection, did not occur until resistance genes from *Solanum demissum* were transferred to commercial potato species, *S. tuberosum*. Since then, race spectrum in different countries/regions has been monitored regularly. Virulence to all major resistance genes was recorded (Guo *et al.*, 2009) ^[68].

2.3 Mating types

Phytophthora infestans is heterothallic and requires two mating types for sexual reproduction. Prior to 1984 the A2 mating type was restricted to Mexico and Andean mountains. Singh *et al.*, (1994) ^[136] reported the presence of A2 mating type in India. Occurrence of A2 mating type in different parts of the world is considered to be due to a second migration of *Phytophthora infestans* from Mexico (Fry *et al.*, 1999) ^[58], the first being from Europe and America during the historical potato famine around the year 1845. In India, A2 mating type has stabilized in temperate hills while the A1 is dominating in sub-tropical plains (Singh *et al.*, 2005; CPRI, 2013) ^[137, 33]. Oospores germinate by means of a germ tube that produces a sporangium, although at times the germ tube grows directly into the mycelium. Sporangia germinate almost entirely by releasing three to eight zoospores at temperatures up to 12 or

15 °C, whereas above 15 °C temperature, sporangia may germinate directly by producing a germ tube (Agrios, 2005)^[2]. The new strains of the pathogen have been found to be more aggressive than the old population (Fry *et al.*, 1999)^[99]. First report of A2 mating type outside Mexico was from Switzerland (Hohl and Iselin, 1984)^[77]. Subsequently, A2 mating type was detected in other countries too. When the two mating types grow adjacently, the female hypha grows through the young antheridium (male reproductive cell) and develops into a globose oogonium (female reproductive cell) above the antheridium. The antheridium then fertilizes the oogonium, which develops into a thick walled and hardy oospore (Tsedaley, 2014)^[150]. Recent work has indicated that the new *Phytophthora infestans* clones, especially the US-8 and US-14 genotypes, are more aggressive (Kirk *et al.*, 2009)^[95]. Migration and sexual recombination can play an important role in enhancing genetic diversity in *P. infestans*. An increasing severity of late blight, a shift in pathogen population toward increased specific virulence and an increased tolerance to metalaxyl has been recorded in past three decades in the north western plains of India (Arora, 2008)^[9].

2.4 Metalaxyl resistance

Metalaxyl, a phenylamide group of systemic fungicide, acts by interference in RNA synthesis by inhibition of rRNA polymerase or both RNA and DNA synthesis. Due to its site specific nature, it becomes more prone to development of resistance in the pathogen. Fungicide resistant isolates were detected in oomycetous fungi soon after the introduction of these fungicides as single products on various crops including *Phytophthora infestans* in Europe, the Middle East and in the Moscow region at the end of 1980. In India, resistance to metalaxyl in *Phytophthora infestans* wild population was first observed in Nilgiri hills of South India in 1989.

Metalaxyl resistant strains appeared towards the end of summer crop season and their frequency increased to 13 per cent in autumn season. Since then the monitoring for metalaxyl resistance is being done regularly. A few strategies have been identified to manage the problem of resistance against metalaxyl in *Phytophthora infestans*. These include withdrawal of straight product and introduction of mixture with contact residual fungicides, regulation of number of sprays and their use early in the season *etc.* This statement supported the references (Arora *et al.*, 2014; Arora *et al.*, 1992)^[8, 7].

2.5 Disease symptoms

Disease symptoms are characterized as circular or irregular water soaked lesions on leaves. Initially, these lesions are localized on tips and margins of the leaves, sometimes surrounded by pale yellowish border. Lesions develop very fast into brown to black necrotic spots and cover entire leaf surface. These lesions also appear on the stem. Whole crop gives blighted appearance and may destroy under high disease severity. Under favourable weather condition (cool and high wetness period) white, fluffy fungal growth may be visible primarily on the underside of the leaf which is the main distinguishing feature of late blight disease. These lesions in affected tubers spread irregularly from the outer surface (3 mm) to deep (10 mm) in pulp. Disease appears on stem as grayish brown to black elongated lesions. Under high moisture and low temperature conditions, it sporulate

profusely and destroy entire green top of the plant. Fry *et al.*, (2001)^[59] also recorded reddish brown, dry, hard and granular lesions in potato tubers.

2.6 Epidemiology

Infected seed tubers put into cold storage are responsible for perpetuation of fungal spores and mycelial growth from one season to another. 0.01% to 3.0% of tuber infection is sufficient to initiate and develop late blight epidemic in the next cropping season (Bhattacharyya *et al.*, 1990). Congenial conditions for appearance and build up of late blight disease includes 18^o to 22 °C temperature and 80 to 100% RH (Fry *et al.*, 2001)^[59]. Production of sporangia and their germination are an important key stages in the life cycle of pathogen which lead to infection, are mainly dependent on high humidity so humidity is always consider as key factor for development of disease. High relative humidity or free moisture is very important for prolonged survival of sporangia. Sporangia germinated by single germ tube at low temperature (13^o to 21 °C) and at high temperature (30 °C) fungus slow down or stop the growth in the field. Forbes and Landeo (2006)^[51] also reported that several wild solanaceous plants carry the pathogen *Phytophthora infestans*. Ideal condition for late blight infection and development are night temperature 10-16 °C with light rain, fog, next day temperature ranged from 13 °C to 16 °C with high relative humidity (Kirk, 2009; Kirk *et al.*, 2013)^[94, 93]. According to Agrios (2005)^[2] at high relative humidity (near 100% RH) fungus restarts the sporulation. The fungal spores are carried by air currents and rain from infected plant to healthy plants and disease cycle restarts. Several workers also reported that the pathogen *Phytophthora infestans* has been carried into field by air currents (Martin *et al.*, 1994; Kirk *et al.*, 2013; Tsedaley, 2014)^[106, 93, 150]. Whenever sporangia come in contact with tuber right from tuberization till harvesting, potato tubers may get infected. Generally, infection takes place when sporangia washed and fall down from lesion to soil and then through soil to tubers. During tuber enlargement period, contact between tubers and sporangia may be more because it develop cracks in the soil and sporangia become readily accessible and infection can be noticed on developing on mature tubers. Temperatures more than 18 °C suppress the infection in tubers. Fry (1998)^[61] observed that sporangia can survive for a weeks or few days in soil so tuber may infected for some period of time even after infection are no longer producing sporangia. The fungus can survive in living tissues, infected seed tubers, infected tubers in cull piles, unharvested and volunteer potatoes left in the ground (Shinners *et al.*, 2009; Kirk *et al.*, 2013)^[130, 93]. Wind, splashed rain, mechanical transport and animals etc are responsible for spread of sporangia from one field to another produced by fungus *Phytophthora infestans* (Martin *et al.*, 1994; Kirk *et al.*, 2013)^[106, 93]. After few days of infection, new sporangiophore emerges from stomata of leaves and gives rise to numerous sporangia. Sporangia germinate either directly by germ tube or indirectly by liberating zoospores. Germ tube enters through stomata and mycelium grows between the cell and send long curled haustoria into the cell. Agrios (2005)^[2] concluded that as disease develop under favourable condition, the already established lesions increase in size and cover the entire surface of leaves, new ones lesion also develop, kill the foliage and finally reduce the tuber yield. During wet weather when sporangia washed away from

leaves and reached into the soil, the second phase of disease starts in the field. Zoospores develop and penetrate the tubers through wounds, mechanical injury or through lenticels. Mycelium grows profusely between the cells and send haustoria into the cells of tubers. During harvesting, tubers got contaminated due to presence of living sporangia in the soil or on diseased foliage and become infected. The excessive humidity (near 100%) coupled with suitable temperature ranged from 15 °C to 25 °C important for disease development (Agrios, 2005) [2]. Generally, pathogen survives in infected tubers in areas where sexual reproduction does not take place. According to Fernandez- Pavia *et al.*, (2004) [49] fungus may survive in soil for many years in areas where viable oospore or sexual reproduction occurs and supported the other (Tiwari *et al.*, 2021) [148].

2.7 Disease management

The main obstacle associated with late blight management is emergence of new strain of *Phytophthora infestans* which always create a major challenge to potato growers. So management of late blight disease through integration of several options *i.e.* cultural control, disease forecasting system, physiological strategies, biological control, host plant resistance and bio-technological approaches can help to overcome this major challenge by avoiding environmental hazards by fungicides.

2.7.1 Cultural control

Garrett and Dendy, (2001) [63] observed that reduction in pathogen populations by lower down the survival, dispersal and reproduction of pathogen are main principle of cultural control which can be achieve by change in date of planting, intercropping, nutrient management and irrigation management.

2.7.1.1 Change in date of planting

Change in normal planting time not only gave maximum tuber yield but also reduced 10-15% late blight disease (Deshraj *et al.*, 1997; Garg *et al.*, 1999; Shailbala and Pundhir, 2006) [42, 62, 122] less late blight disease incidence were noticed in early planted crop. This may be due to significantly higher population of phylloplane fungi in early planted crop and showed less disease pressure (Shailbala and Pundhir, 2007) [123]. Change in planting date affect the crop susceptibility towards disease and its potential to attack and infect adjacent crops (Hospers-Brands *et al.*, 2008) [78].

2.7.1.2 Intercropping

Intercropping of potato with garlic crop in 1:3 ratio gave the best results for management of late blight of potato crop and become as potential intercropped crop due to chemical secreted from root of the garlic affect the late blight disease Kassa and Sommartha, (2006) [86]. Shailbala and Pundhir, (2007) [123] reported that intercropping of potato cultivar Kufri Chipsona with mustard cultivar Divya showed less disease incidence, less infection rate, high tuber yield and finally more return. Bouws and Finckh (2008) [21] concluded that potato intercropped with cereals and clover grass may reduce the late blight disease pressure.

2.7.1.3 Nutrient management

To get high tuber yield and quality tubers, nutrient dose in balanced proportion is primary requirement for potato crop

(Imas and Bansal, 1999) [79]. Always avoid excessive use of nitrogen to the potato crop produce lush green, large canopy which maintain excessive moisture in between the crop and increase the risk of late blight disease. Even make crop more susceptible to infection. Marschner, (1995) [105] reported that optimum dose of potassium fertilizer lower down the late blight incidence.

2.7.1.4 Irrigation management

Potato tubers are sown in ridges to avoid the contact of potato tubers to excessive moisture which may lead to rotting *etc.* Irrigation plays very important role to manage the late blight disease. If soil moisture is more than field capacity for at least 24 hours followed by ≥ 8 mm rainfall results tuber infection by pathogen *Phytophthora infestans* (Adams and Stevensen, 1990) [1]. To minimize the duration of leaf wetness, irrigation management is very important. During morning hours foliage become wet due to presence of dew. Irrigate the crop during morning hours so that irrigation coincides with leaf wetness period just because of dew. Alternatively irrigate the crop during day time because foliage goes through drying period. Very long wetness period is responsible to increase late blight risk. If possible drip irrigation can also be practiced to minimize irrigation applied leaf wetness.

2.7.2 Disease forecasting system

More precise and timely environmental monitoring, weather forecasting, technology transfer network at state, zonal and national level are main factors for disease forecasting. Different late blight forecasting models utilized in many potato growing areas of world which adequately and efficiently forecast the first appearance of disease in particular area. Late blight forecasting models are area specific due to variation in weather condition. Van Everdingen, 1926; Bourke, 1953) [154, 19] developed Irish rules for Irish peoples to forecast the disease and concluded that atleast 12 hours of wetness period, air temperature not less than 10 °C and 90% RH are required by sporangia of *Phytophthora infestans* for germination and infection. Temperature, dew period, cloudiness and rainfall were used to predict initial appearance of late blight disease in Holland. According to Smith (1956) [139], minimum temperature 10 °C for two consecutive days and at least 11 hours of RH $\geq 90\%$ is favourable for disease development. Accurate forecasting can be possible to those diseases which are sporadic in nature (Bourke, 1970; Keane, 1995) [20, 88]. Daily maximum temperature 17^o to 24 °C, minimum temperature ≥ 10 °C, RH at noon $\geq 75\%$, daily rainfall ≥ 0.1 mm are important factor to assess the risk of potato late blight disease (Forsund, 1983) [55]. Henshall *et al.*, (2006) [76] developed Shtienberg model for late blight risk in which they combined inoculum index with infection index to develop 0 to 3 late blight risk index corresponding to nil (0), light (1), moderate (2) and severe (3) risk of disease. Sharma (2000) [128] developed late blight forecasting model for North Western region of India especially Jalandhar and concluded that disease development was positively correlated with maximum RH, rainfall, dew and cloudy days.

2.7.2.1 Favourable days model

Daily rainfall, maximum and minimum temperature are important features of favourable days model. For different potato growing areas, thermohydrograph were prepared by Dutt (1964) [45] which indicate blight periods of that particular

area. Singh *et al.*, (2000) ^[135] developed, a computerized forecasting system known as Jhulsacast against late blight of potato for Western U.P. for rainy as well as non-rainy years, in which model 1 for rainy year includes measurable rain (0.1-0.5 mm) for a minimum of 2 consecutive days, 5 day moving >85% relative humidity period ≥ 50 hours and 5 day moving congenial temperature (7.2-26.6 °C) period ≥ 120 hours while model 2 for non-rainy year includes 7 day moving >85% relative humidity period ≥ 60 hours and 7 day moving congenial temperature (7.2-26.6 °C) period ≥ 120 hours. If 7 days moving precipitation average 30 mm for Shimla, 28.9 mm for Ooctamound and 38.5 mm for Shillong with mean temperature ≤ 23.9 °C for 7 consecutive days or hourly temperature ranged from 10⁰ – 20 °C with $\geq 80\%$ RH for 18 hours for two consecutive days, late blight would appear within 3 weeks (Bhattacharya *et al.*, 1983) ^[16].

2.7.2.2 Severity value model

The first appearance of late blight and periods of late blight favorable weather can be predicted using relative humidity, rainfall and temperature data collected from an electronic weather monitor or a hygrothermograph. The weather data is converted into units called severity values (SV) for the purpose of predicting late blight outbreaks. Late blight is first expected to appear within 12 weeks after 18 SV have accumulated. Fungicide applications to protect potatoes should be initiated as soon as possible after 18 SV have accumulated. The Wallin system was developed by Wallin JR (1962) ^[161] in Midwestern United States which interprets the epidemiologic consequences of extended periods of high relative humidity and temperature during those humid periods. Late blight is forecast to occur 7 to 14 days after 18 severity value have accumulated.

2.7.2.3 Negative prognosis model

Ullrich and Schrodter (1966) ^[152] gave negative prognosis model uses measurement of temperature, relative humidity and rainfall to predict when late blight epidemics are not likely to occur. It has been used to predict the timing of the first prophylactic treatment in Germany. Daily and accumulated risk values over a week are calculated starting at crop emergence. Disease is expected when accumulated risk value has exceeded the threshold of 150.

2.7.2.4 Model with spatial dynamics

Minogue and Fry (1983) ^[107] modelled the spatial dynamics of potato late blight and found that blight foci expand with constant velocity determined by host resistance and fungicide use. Ferrandino, 1989; Paysour and Fry, 1983) ^[50, 112] used a model to calculate the level of inter plot interference in experiment with potato late blight.

2.7.2.4.1 Mathematical model on late blight disease

Johnson *et al.*, (1996) ^[82] developed two discriminant function and two logistic regression models to forecast outbreak of late blight in South Central area of Washington State. The first discriminant function uses the following variables: late blight out break during the preceding year ($Y_p=0$ =no; $Y_p=1$ = yes), number of days with rains greater than or equal to 0.25 mm during the month of April and May and total precipitation during May when daily minimum temperature was more or equal to 5 °C (P_m). The second discriminant function uses Y_p , R_{am} and number of days with rain more or equal to 0.25 mm

during July and August.

2.7.2.4.2 Simulation model

Waggoner, (1968) ^[160] presented the first computer simulation model for potato late blight in which the effect of environmental condition on the resistance component was extensively modelled. A procedure for timely fungicide application to suppress potato late blight was developed by using two simulation models. The first model stimulated the effect of environment, fungicide and host resistance on *Phytophthora infestans* in potato foliage (Bruhn and Fry, 1981; Bruhn and Fry, 1982 a, b) ^[22, 23, 24]. Fry *et al.* (1983) ^[56] described forecasting model which was derived from two simulation model. One model described weather effects on fungicide distribution and amount while second model described effects of host resistance and weather on development of *Phytophthora infestans*. Van Oijen, (1992) ^[155] used simulation model to evaluate the importance of different host resistance components. Andrare Piedra *et al.*, (2005) ^[6] discovered the domain over which a validated model may be properly used, was illustrated with lateblight, a mathematical model that stimulates the effect of weather, host growth, resistance and fungicide use on asexual development and growth of *Phytophthora infestans* on potato foliage. Forbes *et al.*, (2008) ^[52] developed most recent version LB2004 which has been validated in the high- lands tropics and several other countries.

2.7.3 Decision support system/expert system

Decision support system increases the efficacy of control strategies without increasing disease risk in potato plant. Krause and colleagues at Pennsylvania State University (Krause *et al.*, 1975) ^[96] developed a computerized forecast model known as BLITECAST. PhytoPRE is a computer based information and decision support system for potato late blight in Switzerland which consists of an epidemiological forecast model, a set of decision rules and an information system (Ferrer *et al.*, 1993) ^[53]. PROGEB forecasting model have PHYTEB as one of their components which forecasts *Phytophthora infestans* and stimulates symptomatic stages of the host (latent period, preinfection, number of infection, amount of dead tissue). PHYTEB consist of two sub models SIM- PHYT-1 and SIMPHYT-2 and forecast the beginning of epidemic 7-10 days ahead (Gutsche, 1993) ^[70]. Wharton *et al.*, (2008) ^[163] developed a web-based expert system to help potato growers in Michigan regarding control measures to mitigate the risk of potato late blight disease development. Fry *et al.*, (2015) ^[57] also discussed the BlightPro, a decision support system developed to aid the management of potato late blight in USA. For Indian condition Indoblighcast were developed for managing the disease in a cost effective and environmental friendly manner (Govindkrishnan *et al.*, 2015) ^[67]. SIM- BLIGHT 1 model had been developed which requires temperature, relative humidity, information on soil moisture, crop prevalence and cultivar susceptibility (Benno kleinhenz *et al.*, 2007) ^[13].

Based on agro advisory system, Pande *et al.*, (2009) ^[111] used mobile phone to forecast late blight disease in which actual late blight queries received from farmers were compared and validated with cumulative composite rise index taken from existing forecasting models. System based on diagnostic approaches was incorporated with forecasting models.

2.7.4 Physiological strategies

Every crop plant has potential to develop physiological strategies to deal with diseases. These physiological strategies are escape, resistance, tolerance and recovery. The fungus *Phytophthora infestans* attack and affect the crop resulting reduction of the leaf area which directly lower down the photosynthesis process and reduce the crop capacity to produce photosynthetic assimilates require for growth and maintenance of haulm and for tuber bulking. The pathogen directly infected the tuber and reduces the yield & market value of tubers. These infested tubers put for storage in cold storage serve as important source of inoculum. An infected crop also serves as source of inoculum which is really unacceptable in area where potato crop is grown in high proportion. Due to these reasons, two physiological strategies *i.e.* escape and resistance work against late blight disease in potato crop. Disease can be escape by accelerating the early canopy growth without advancing tuber bulking. This may be possible through agronomic practices such as extra nitrogen application (which might delay tuber bulking up to some extent). Advancement in tuber bulking without advancing canopy development can also be one of the escape strategies. This can be achieving by manipulating the physiological age of the seed tubers (Struik and Wiersema, 1999) [142]. Struik, (2010) [143] worked on physiological strategies to cope with disease based on escape and resistance. Escape strategy includes advance or change the entire crop cycle so at the time of pathogen attack, the crop will become mature enough to cope with disease. If potato crop is planted in early October in sub-tropical, it may escape blight attack. The advancing of crop cycle can be done by early planting of crop, planting larger seed tubers *etc* (Struik and Wiersema, 1999) [142]. Accelerate the tuber bulking without advancing it through chemical treatment of foliage may be other escape strategy. Other physiological strategies to cope with late blight disease through resistance include strengthening the race non-specific resistance which results delay in initial infestation and slower development of disease. Race non-specific resistance against late blight is strongly associated with foliage maturity type (Visker *et al.*, 2003a, 2003b, 2004, 2005) [158, 157, 159, 156]. The other resistance strategy is influencing the microenvironment around the basal leave which will slow down the development of late blight epidemic and inhibit first infestation. Planting pattern, row to row distance, seed treatment, seed tuber size *etc.* will make such microenvironment which is not conducive to infection by the pathogen.

2.7.5 Biological control

Management of highly destructive disease such as late blight of potato is slightly difficult due to rapid establishment of infection and explosive disease development. So, little information is available regarding biological control as potential alternative against late blight disease. In biological control, living microorganism or abiotic products provide disease protection through production of antibiotics, competition for food and space, induced plant resistance *etc.* Daayf *et al.*, (2003) [37] also studied on biological control of potato late blight by detached leave method, whole planting testing system and *in-vitro*. Shailbala and Pundhir, (2008) [125] observed that *Trichoderma* (formulation) @ 10 g/l and *Pseudomonas* (formulation) @ 10 g/l showed antagonistic behaviour and gave the best results against late blight disease. Various fungi, bacteria and different compost extracts were

tested against *Phytophthora infestans* in potato crop (Ghorbani *et al.*, 2005; Lamsal *et al.*, 2013) [64].

Kaya *et al.* (2006) [87] evaluated me- tabolites produced by *Xenorhabdus* spp against *P. in- festans*. Bio-agent *Steinernema feltiae* was also studied against late blight both *in vivo* and *in vitro*. Yang *et al.*, (2001) [166] tested metabolites from culture broth of *Xenorhabdus nematophilus* isolated from nematode *Steinernema caropcapsae* for control late blight in potato potted plants and revealed that metabolites @ 25 and 50 mg/l found effective in reducing late blight intensity. Application of *Xenorhabdus* spp. gave most consistent results of biological control against late blight disease (Yang *et al.*, 2001) [166].

2.7.5.1 Potential antagonists

Many scientists worked on infection of *Phytophthora infestans* on potato tubers to search for micro-organism as possible antagonists to the test pathogen. Isolates of *Pseudomonas* spp, *Burkholderia* spp., *Streptomyces* spp. and *Trichoderma* spp. were obtained from stems, leaves, tubers and rhizoplane of potato plants. The activity of these micro-organism to A1 and A2 mating type of *P. infestans* were assessed on potato leaves in moist chamber, greenhouse and in field. Reduction in late blight severity occurred with *Burkholderia* spp, *Streptomyces* spp and *Pseudomonas* spp. applied individually or in combination (Lozoya-Saldana *et al.*, 2006) [102]. The impacts of potentially effective bio-control agents against late blight would enhance disease management in organic cropping system and its contribution would be of great relevance. A combination of bio-control agents with products such as neem oil could be effective to manage late blight severity and it could be another option to reduce crop losses caused by the pathogen. Among the seven potato phylloplane fungi, only three fungi *viz.*, *Fusarium* spp, *Trichoderma* spp, *Aspergillus* spp showed antagonistic potential against *P. infestans*, causal agent of late blight of potato (Shailbala and Pundhir, 2007) [123].

2.7.5.2 Systemic acquired resistance

Yan *et al.*, (2002) [165] suggest that induced protection elicited by both bacilli and pseudomonad PGPR strains was SA (salicylic acid) independent but ethylene and jasmonic acid depend- ent, whereas systemic acquired resistance elicited by the pathogen and induced local resistance by BABA (β amino butyric acid) were SA-dependent. BABA and phosphoric acid @ 2 g/l involved in systemic acquired resistance reduced late blight significantly and induced expression of defense gene in Kufri Chandramukhi (CPRI, 2014-15) [34].

2.7.5.2.1 Use of endophytic organism

Some endophytic organisms were also tested against late blight of potato. Control of late blight was attempted with arbuscular mycorrhizal fungi (O' herlihy *et al.*, 2003) [110]. It was reported that there was reduction of disease progress rate. For potato late blight, this can be an important strategy to reduce crop losses.

2.7.5.2.2 Bio-fungicide

Several commercial formulations of bio-control agents have been tested for efficacy against late blight. Soytong and Ratanacherdchai, (2005) [141] worked on myco-fungicide against late blight of potato and results indicate that *Chaetomium* myco-fungicide could reduce incidence of late

blight of potato caused by *Phytophthora infestans* and reduce the population in the soil with significant reduction the potato late blight. Formulation of *T. viride* and *Phytophthora viridicatum* significantly reduced *Phytophthora infestans* sporangial germination and has potential to control potato late blight under control condition (Gupta *et al.*, 2004) ^[69]. Trichotoxin A50 is a brown liquid formulation of natural substances that can induce immunity of plants such as potato, cucumber and tomato to disease by interval spraying at 7-10 days to the plant @ 50 cc/20 lit of water (Suwan *et al.*, 2000) ^[146]. Many reports are available about antibiotic-producing microorganisms tested for bio-control capacity (Behal, 2000) ^[12]. However, only few antibiotics have been developed as commercial compounds in agriculture. Some of the reasons include their inconsistent efficacy in the field, their instability and the costs related to their production.

2.7.5.2.3 Rhizobacteria

Drenching with plant growth promoting rhizobacteria isolates increased the total weight of tubers per potato plants, in addition to effectively controlling late blight caused by *P. infestans* (Kim and Jeun, 2006) ^[90]. Beneficial effects reported by plant growth promoting rhizobacteria includes increases in a number of parameters such as germination rate, root, shoot growth, yields etc (Lucy *et al.*, 2004) ^[103]. *Bacillus pumilus* and *Pseudo- monas fluorescens* induced resistance to *P. infestans* and there was reduction in zoospore formation and germination (Yang *et al.*, 2001) ^[166]. In *in vitro* and *in vivo* studies, *Pseudomonas*, *Rahnella* and *Serratia* can less- en late blight symptoms by a combination of antibiosis and induced resistance against *Phytophthora infestans* (Daayf *et al.*, 2003) ^[37]. Therefore, bio-control using plant growth promoting rhizobacteria represents a potentially attractive alternatives disease management approach since they are known to promote growth and reduce disease in crops (Jetiyanon and Kloepferm, 2002) ^[80].

2.7.5.2.4 Essential oils

Mari *et al.*, (2003) ^[104] gave the information about the use of natural compounds derived from plants to control late blight in potato crop. The use of biological compounds extracted from plants may be an alternative to conventionally used fungicides to control phyto-pathogenic fungi, due to their being bio-active chemicals such as flavonoids, phenols, tannins, alkaloids, quinons, saponins and sterols (Burt, 2004) ^[25]. Bio-logically active compounds found in plants appear to be more adaptable, acceptable and safer than synthetic compounds and display a wealthy source of potential pathogens controlling agents (Tripathi *et al.*, 2008) ^[149]. Amini *et al.*, (2012) ^[5] reported antifungal activity of essential oil obtained from three medicinal plants *i.e.* *Zataria multiflora*, *Thymus vulgaris* and *Thymus kotschyanus* against phyto-pathogenic fungi. Even Caraway essential oil delayed the onset of late blight for about 10-14 days under field conditions (Hannukkala *et al.*, 2002) ^[73]. Keskitalo *et al.*, (2005) ^[89] worked and reported that caraway oil delay the onset of disease for 14 days. The control activity of this oil is based on prevention of late blight sporangia production and growth at low oil concentration. Essential oils are oily liquids obtained from plants through fermentation, enfleurage, extraction and steam distillation (Burt, 2004) ^[25]. Essential oils have two prominent features *i.e.* low toxicity for people and environment due to their natural properties and low risk

for resistance development by pathogenic microorganisms (Dafererra *et al.*, 2000) ^[39].

2.7.5.2.5 Plant extracts

Plant extracts are obtained by filtration, distillation and evaporation (Wang *et al.*, 2004) ^[162]. The work on plant extracts with anti-oomycetes activity has been increased over the years and efficacy of plant extracts against pathogen has also been demonstrated. Several preliminary *in vitro* studies has been conducted in China and India (Cao *et al.*, 2003; Deepa *et al.*, 2004) ^[26, 41]. Six extracts from plant material (*Galla chinensis*, *Potentilla erecta*, *Rheum rhabarbarum*, *Salviae officinalis*, *Sophora flavescens*, and *Terminalia chebula*) were tested for controlling effects against the infection of *Phytophthora infestans* on detached potato leaves, seedlings and tuber slices. On detached leaves, *G. chinensis* was the best. On seedlings, *R. rhabarbarum* showed the best inhibiting effect (Shutong Wang *et al.*, 2007) ^[133]. Wang *et al.* (2004) ^[162] reported that *Inula viscosa* reduced 90% late blight severity in potato plants. Extracts of *Terminalia chebula* and *Galla chinensis* were also found effective to control late blight of potato and 30% and 10% inhibition of disease were noticed respectively (Cao *et al.*, 2004) ^[25]. One of the most effective treatments was extract of garlic cloves which at 1 or 2 percent completely inhibit the zoospore formation and colony growth of pathogen the statement supported the references (Cao and Van Bruggen, 2001; Halama and Haluwin, 2004) ^[27, 71]. Ahn *et al.*, (2005) ^[3] reported that methanolic extract of galls (gall caused by an aphid, *Schlechtendalia chinensis* in nutgall sumac tree) reduced approximately 90% late blight severity. Extract derived from *Pseudarthria viscida*, *Cassia tora* and *Catalpa ovata* have also inhibited growth of *Phytophthora infestans* under laboratory condition (Kim *et al.*, 2004; Cho *et al.*, 2006) ^[91, 28].

2.7.5.2.6 Other products

Scheuerell and Mahaffee (2002) ^[118] reported that combination of water based extract fermented tea and plant compost (known as compost tea) as foliar spray in potato not only improve soil fertility but also control pathogen and pests. This compost tea may be considered as one of the best option to control of late blight disease of potato (Rashidul Islam *et al.*, 2013; Al-Dahmani *et al.*, 2005 ; Scheuerell and Mahaffee, 2006) ^[116, 4, 119].

2.7.6 HPR

Due to rapid development of new strains of *Phytophthora infestans*, host plant resistance appears eco-friendly and economic feasible approach for late blight management. Shailbala and Pundhir, 2008; Njuaem *et al.*, 2001) ^[125, 109] also reported variation in resistance against late blight among potato cultivars. Resistant potato cultivars could easily destroyed by newer strains of pathogen since single gene control the resistance. Potato cultivars with high level of resistance can be helpful and allow growing even in cool season without use of fungicides. Durable or polygenic or field resistance is generally controlled by several minor genes which give slow blighting effect. Fry, (1977) ^[60] reported that potato cultivars with durable or polygenic resistance showed significantly less values of area under disease progress curve and low infection rates as compared to susceptible cultivars. Many released and improved resistant potato cultivars lost

their resistance against disease but was able to tolerate the disease and gave satisfactorily yield (GLIB and CIP, 2004a; Jones, 1998; ATTRA, 2004) [66, 29, 83, 11]. Shtienberg *et al.*, (1994) [132] also reported that compatible races of pathogen *Phytophthora infestans* fastly broken down race specific oligogenic resistance (CIP, 1989; FAO, 2008; Popokova, 1972) [29, 48, 114]. Shtienberg *et al.*, (1994) [132] concluded that use of resistant cultivars can be one component to be use in late blight management for tropical condition. Initially resistance gene was identified in *Solanum demissum* which provided base to transfer resistance gene from it and to develop resistant cultivar *i.e.* Kufri jyoti (having R gene 3.4.7) for commercial cultivation in 1968. Low inoculum load and shorter late blight congenial period was important factor behind popularity of this cultivar in several part of country till date. Shekhawat, (2000) [129] reported that *Solanum tuberosum* subsp *andigena* have also been exploited for development of resistant cultivar. late blight resistant cultivars have capacity to delay the disease onset and further disease development so fewer fungicidal sprays on these resistant cultivars can effectively manage the disease supported the references (Agrios, 2005; Binyam *et al.*, 2014; Kirk *et al.*, 2001; Song *et al.*, 2003) [2, 18, 92, 140].

2.7.7 Bio-technological methods

Potato is considered as poor man food and late blight as catastrophic disease. Late blight free potato will have direct impact on people food security and income in developing countries. Worldwide use efficiencies of land, water, nutrients and energy can greatly improve by achieving disease free potato tubers but practically it is not possible. To get disease free potato tubers, bio-technological approach against late blight can be the best option. The knowledge on molecular biology and genetics of interaction between plant and oomycetes mainly focused on discovery of many resistance genes, numerous effector proteins and analysis of their mode of action which provide important information required for development of durable resistance (Haverkort *et al.*, 2009) [74]. A novel approach is mainly based on cisgenic modification which depends on marker free gene pyramiding with their spatial and temporal deployment. This cisgenic modification approach with potato's own gene is societally acceptable and also results in simplification in the legislation on the use of cisgenic modification approach (Haverkort *et al.*, 2009) [74]. So, DuRPh (Durable Resistance against *Phytophthora infestans*) programme were made which focus on durable resistance in potato crop. In DuRPh programme, cloning, transformation and selection of desired resistance gene ids important part of process to develop late blight resistance. The main aim of this programme is to develop cisgenic potato variety with durable resistance feature against late blight disease. Some important features of DuRPh programme includes Genetic modification: This includes detection, isolation, cloning and transformation of gene from wild species into existing varieties through *Agrobacterium tumefaciens*, a bacterial vector. The plantlets regenerated from callus are screened to assess for resistance. Important point is that they should have the same phenotype as the wild variety into which resistance genes introduced Cisgenic approach: In this approach, natural resistance genes from plant itself or from crossable species are used and the choice of cisgenic was mainly based on availability of resistance gene in potato crop. Jochemsen, (2008) [81] reported that this approach was

ethically more acceptable to the public. Through gene pyramiding, several resistance genes are inserted to reduce the rapid break down. In this programme, no markers are used so variety obtained will be marker free. PCR techniques are used to assure that resistance gene is present in genotype show resistant against all known pathotypes of *Phytophthora infestans*. In different varieties at different sites at different times, various combinations of stacked genes are de-ployed resulting spatial and temporal separated de-ployment of resistance gene. The start and end product during this programme is potato varieties which consist of potato genes only. In this programme, no new varieties developed and only point is that in old variety resistance gene of wild potato species were incorporated. The exploitation of the R genes which is not to be homologous to previously discovered ones, is possible by protecting the intellectual property and making them available (not exclusively) to private potato breeding companies.

2.7.8 Compost

The decomposition of organic wastes leads to the formation of the most used soil amendment, the compost. Balanced use of fertilizers increases soil fertility without environmental degradation in a sustainable way (Shrestha *et al.*, 2020) [131].

3. Conclusion

Late blight of potato is the most dreaded disease and will continue to remain as the pathogen is evolving at a fast rate and adapting to new environments and hosts. Due to highly variable nature of late blight pathogen, preventing the establishment of infection is perhaps the most interesting strategy for management of late blight. Detaining pathogen development after infection is difficult. So application of control measures can reduce the survival period as well as the effectiveness of the source of initial inoculum which prevent sporangia germination (germ tube formation) and/or zoospore germination. Late blight forecasting models will help the growers in managing this serious disease. Farmers should contact their local extension office regarding disease forecasting and accordingly they can apply the preventive control measures. Use of resistant varieties along with physiological strategies for disease management can be the best option. Cultural practices for disease management always consider the first line of defense while biological control can become an integral part of management programme aimed at controlling late blight. Biotechnological approaches can help in development of durable resistance varieties. So management strategies include disease forecasting system, host plant resistance, physiological strategies, cultural control, biological control and biotechnological approach, as already discussed certainly can become an integral part in management programme aimed at controlling late blight in both conventional and organic cropping system of potato. So, alternative non-fungicidal eco-friendly control measure against late blight should be implemented under a holistic approach. As more and more information is being generated, there is a need to develop an appropriate disease management strategy based on farmer friendly information technology.

4. References

1. Adams SS, Stevenson WR. Water management, disease development and potato production. *Am. Potato J.* 1990;67(1):3-11.

2. Agrios GN. Plant Pathology. 5th edition. Academic Press, London, New York, 2005, 922p.
3. Ahn Y, Lee H, Oh H, Kim H, Lee Y. Antifungal activity and mode of action of gallarhois derived phenolic against phyto-pathogenic fungi. Pesticide Bio-chem. Physiol. 2005;81:105-112.
4. Al-Dahmani JH, Abbasi PA, Sahin F, Hoitink HJ, Miller SA. Reduction of bacterial leaf spot severity on radish, lettuce and tomato plants grown in compost amended potting mixes. Can. J Plant Path. 2005;27:186-193.
5. Amini M, Safaie N, Salmani MJ, Shams-Bakhsh M. Antifungal activity of three medicinal oils against some phytopathogenic fungi. Trakia J Sci. 2012;10:1-8.
6. Andrare-Piedra JL, Forbes GA, Shtienberg D, Grunwald NJ, Taipe MV, Hijmans RJ. Qualification of a plant disease simulation model: Performance of the late blight model across a broad range of environments. Phytopathol. 2005;95:1412-1422.
7. Arora RK, Kamble SS, Gangawane LV. Resistance to metalaxyl in *P. infestans* in Nilgiri hills of southern India. Phytophthora Newsl. 1992;18:8-9.
8. Arora RK, Sharma S, Singh BP. Late blight disease of potato and its management. Potato J. 2014;41(1):16-40.
9. Arora RK. Late blight an increasing threat to seed potato production in North-Western plains of India. Acta Horticulturae. 2008;834:201-202.
10. Arora S, Steuernagel B, Gaurav K, Chandramohan S, Long Y, Matny. Resistance gene cloning from a wild crop relative by sequence capture and association genetics. Nature Biotechnology. 2019;37:139-143.
11. ATTRA. Organic alternatives for late blight control in potatoes. National Sustainable Agriculture Information Service; c2004.
12. Behal V. Bio-active products from *Streptomyces*. Adv. Appl. Microbiol. 2000;47:113-157.
13. Benno Kleinhenz, Kristina F, Joachim K, Dietmar R. SIMBLIGHT1-A new model to predict first occurrence of potato late blight. EPPO Bull. 2007;37:339-343.
14. Berg A. Tomato late blight and its relation to late blight of potato. West Virginia Agric. Exp. Stn. Tech. Bull. 1926;205:1-31.
15. Bhattacharyya SK, Singh BP. Durable resistance against the potato late blight in India. Indian Phytopath. 1986;39:615-619.
16. Bhattacharyya SK, Raj S, Singh DS, Khanna R, Sharma SR. Forecasting late blight of potato in Indian hills. In: B. B. Nagaich *et al.* (eds.) Potato in developing countries. Indian Potato Association. CPRI. Shimla; c1983. p. 20-23.
17. Bhattacharyya SK, Singh BP, Sharma VC, Bambawal OM, Arora RK, Singh PH. Mode of survival and source of primary inoculum of late blight of potato. Intern. J Tropic. Pl. Dis. 1990;8:78-88.
18. Binyam T, Hussen T, Tsegaw T. Efficacy of reduced dose of fungicide sprays in the management of late blight (*Phytophthora infestans*) disease on selected potato (*Solanum tuberosum* L.) varieties Haramaya, East-ern Ethiopia. J of Biology, Agriculture and Healthcare, 2014;4(20):46-52.
19. Bourke PMA. Potato blight and weather a fresh approach. Technical Note No-12. Irish Meteorological Service. 1953.
20. Bourke PMA. Use of weather information in the prediction of plant disease epiphytotics. Ann. Rev. Phyto pathol. 1970;8:345-370.
21. Bouws H, Finckh MR. Growing potatoes in narrow strips perpendicular to wind and separated by non-potatoes. Plant Path. 2008;52(5):916-927.
22. Bruhn JA, Fry WE. Analysis of potato late blight epidemiology by simulation modeling. Phytopathol. 1981;71:612-616.
23. Bruhn JA, Fry WE. A statistical model of fungicide deposition on potato foliage. Phytopathol. 1982a;73:1301-1305.
24. Bruhn JA, Fry WE. A mathematical model of the spatial and temporal dynamics of chlorothalonil residues on potato foliage. Phytopathol. 1982b;72:1306-1312.
25. Burt S. Essential oils: Their antibacterial properties and potential applications in foods- A review. Intern. J Food Microbiol. 2004;94:223-253.
26. Cao KQ, Wang ST, Forrer HR, Fried PM. Inhibitory effects of chinese medicinal plants against *Phytophthora infestans* on potatoes. Regional workshop on potato late blight for east and south-east Asia and the Pacific, International Potato Center and Ministry of Agriculture and Irrigation, Myanmar, Yezin Agricultural University. c2004. p. 45-53.
27. Cao KQ, Wang ST, Kessler P, Fried PM, Forrer HR. Potato late blight: A new approach for an outdoor screening of substitutes for copper fungicides. Krautfaulebekämpfung im Bio-Kartoffelanbau ohne Kupfer? Agrarforschung. 2003;10:182.
28. Cao KQ, Van Bruggen AHC. Inhibitory efficacy of several plant extracts and plant products on *Phytophthora infestans*. J Agric. Univ. Hebei. 2001;24:91-99.
29. Cho JY, Kim HY, Choi GJ, Jang KS, Lim HK, Lim CH. Dehydroalpha-lapachone isolated from *Catalpa ovata* stems: Activity against plant pathogenic fungi. Pest Management Sci. 2006;62:414-41.
30. CIP. Fungal diseases of potato. Report of planning conference on fungal diseases of the potato. CIP, Lima, Peru, 21th - 25th September; 1989. p 216.
31. CPRI. Annual Scientific Report. Central Potato Research Institute, Shimla; 1981. p. 61-64.
32. CPRI. Annual Scientific Report. Central Potato Research Institute, Shimla; 1987. p. 96-109.
33. CPRI. Annual Scientific Report. Central Potato Research Institute, Shimla; 1999. p 70-77.
34. CPRI. Annual Progress Report. Central Potato Research Institute, Shimla; c2013; p. 158.
35. CPRI. Annual Progress Report. Central Potato Research Institute, Shimla; c2014-15, p. 198.
36. Cristinzio G, Testa A, Pugliano P. Races of *Phytophthora infestans* in Italy. Informatore Fitopatol. 1998;48(9):49-51.
37. Daayf F, Platt HW. Changes in metalaxyl resistance among glucose phosphate isomerase genotypes of *Phytophthora infestans* in Canada during 1997-1998. Am. J Potato Res. 2000;77:311-318.
38. Daayf F, Adam L, Fernando WGD. Comparative screening of bacteria for biological control of potato late blight (strain US-8), using *in vitro*, detached leaves, and whole-plant testing systems. Can. J Plant Pathol. 2003;25:276-284.
39. Daayf F, Platt HW, Mahuku G, Peters RD. Relationships between RAPDs, Gpiallozyme patterns, mating types and

- resistance to metalaxyl of *Phytophthora infestans* in Canada in 1997. *Am. J Potato Res.* 2001;78:129-139.
40. Dafererra DJ, Ziogas BN, Polissiou MG. GC-MS analysis of essential oils from some greek aromatic plants and their fungitoxicity on *Penicillium digitatum*. *J. Agric. Food Chem.* 2000;48:2576-2581.
 41. De Bary A. Research into the nature, the potato fungus *Phytophthora infestans*. *J.R. Agric. Sco. Eng.* 1876;12:239-269.
 42. Deepa MA, Bai VN, Basker S. Antifungal properties of *Pseudarthria viscida*. *Fitoterapia.* 2004;75:581-584.
 43. Deshraj Garg VK, Singh S. Effect of date of planting and haulms killing on total yield and seed size tubers in potato cultivar K. Jyoti. *J Indian Potato Assoc.* 1997;24(1-2):71-73.
 44. Dick MW. Stramenopilous fungus. Kluwar, Hingham MA, Diniz LP, Maffia LA., Dhingra OD, Casali VWD, Santos RHS and Mizubuti ESG. 2006. Avaliacao de produtos alternativos para controle da requeima do tomateiro. *Fitopatologia Brasileira.* 2001;31:171-179.
 45. Du Y, Chen X, Guo Y, Zhang X, Zhang H, Li F. *Phytophthora infestans* RXLR effector PITG20303 targets a potato MKK1 protein to suppress plant immunity. *New Phytologist.* 2021;229:501-515.
 46. Dutt BL. Late blight of potato in India. In: *Distribution and blight period.* *Indian Potato J.* 1964;6:34-41.
 47. Dutt BL. Late blight of potato in India. In: *Distribution and incidence of physiologic races.* *Indian Potato J.* 1965;7:23-28.
 48. Eduardo SGM, Valdir LJ, Gregory AF. Management of late blight with Alternative products. *Pest Tech.* 2007;1(2):106-116.
 49. FAO. 2008. *Potato World: Africa-International year of the potato;* c2008. <http://www.potato2008.org/en/world/africa.html>. Accessed at: 12/5/2014.
 50. Fernandez-Pavia SP, Grunwald NJ, Diaz-Valasis M, Cadena-Hinojosa M, Fry WE. Soil-borne oospores of *Phytophthora infestans* in Central Mexico survive winter fallow and infect potato plants in the field. *Plant Dis.* 2004;88:29-33.
 51. Ferrandino FJ. Spatial and temporal variation of a defoliating plant disease and reduction in yield. *Agricultural and Forest Meteorology.* 1989;47:273-289.
 52. Forbes GA, Landeo JA. late blight. In: (Gopal, J.P.K.S.M. ed.) *Handbook of potato production, improvement and post harvest management,* Haworth Press Inc., Binghamton, New York; c2006. p. 279-320.
 53. Forbes GA, Fry WE, Andrade Piedra JL, Shtienberg D. Integrated management of disease caused by fungi, phytoplasma and bacteria. (A. Ciancio and K.G. Mukherji eds). Springer; c2008. p. 161-177.
 54. Forrer HR, Gujer HO, Fried PM. Phyto-PRE-A comprehensive information and decision support system for late blight in potatoes. In: *Workshop on computer based decision support system (DSS) in crop protection.* Italy; c1993.
 55. Forster H, Coffey MD, Elwood H and Sagin ML. Sequence analysis of the small sub-unit ribosomal RNAs of the zoospore fungi and implication for fungal evolution. *Mycologia.* 1990;82:306-312.
 56. Forsund E. Late blight forecasting in Norway 1957-1980. *EPPPO Bull.* 1983;13:255-258.
 57. Fry WE, Apple AE, Bruhn JA.. Evaluation of potato late blight forecast modified to incorporate host resistance and fungicide weathering. *Phytopathol.* 1983;73:1054-1059.
 58. Fry WE, Small IM, Danies G. Epidemiology and decision support system. In: *3rd International symposium on Phytophthora: Taxonomy, genomics, pathogenicity, resistance and disease management.* 9th- 12th September, 2015. Bengaluru, India; c2015. p. 31.
 59. Fry WE, Smart CD, Monti L, Leone A, Struik PC, Hide GA, Storey RMJ. The return of *Phytophthora infestans*, a potato pathogen that just won't quit. In: (Struik and Hide eds). *Proceedings of the 14th Triennial conference of the European association for potato research, Sorrento, Italy* 2nd-7th May, Extra edition. 1999;42:279-82.
 60. Fry WE, Thurston HD, Stevenson WR. late blight. In : Stevenson WR, Loria R, Franc GD, Weingartner DP. (eds.) *Compendium of potato diseases* (2nd edn), The American Phytopathological Society, St Paul, USA; c2001. p. 22-23.
 61. Fry WE. Integrated control of late blight effects of polygenic resistance and techniques of timing fungicide applications. *Phytopathol.* 1977;67:415-420.
 62. Fry WE. *Vegetable crops: late blight potatoes and tomatoes.* Cornell University Cooperative Extension. New York State. USA; c1998. Available on: Page: 726.20 Date 7-1998). (http://vegetablemdonline.ppath.cornell.edu/factsheets/Potato_LateBlt.htm).
 63. Garg VK Deshraj, Singh S. Influence of date of planting and haulm killing on the yield of seed sized tubers in Shimla hills. *J Indian Potato Assoc.* 1999;26(1-2):1-6.
 64. Garrett KA, Dendy SP. Cultural practices in potato late blight management. In: *Complementing resistance to late blight (Phytophthora infestans) in the Andes* (Fernandez-Northcoted, N. (ed.). *Proceedings of GILB Latin American workshop I, 13th-16th February, 2001, Cochabamba, Bolivia;* c2001. p. 107-113.
 65. Ghorbani R, Wilcockson S, Leifert C. Alternative treatments for late blight control in organic potato: Antagonistic micro-organism and compost extracts for activity against *Phytophthora infestans*. *Potato Res.* 2005;48:181-189.
 66. Giddings NJ, Berg A. A comparison of the late blight of tomato and potato: A preliminary report. *Phytopathol.* 1919;9:209-10.
 67. GILB, CIP. Ethiopia: Potato production areas and average yields; c2004. Available on: Accessed at 29/3/2014 <http://gilb.cip.cgiar.org/countryprofiles/africa/ethiopia/>.
 68. Govindakrishnan PM, Singh BP, Ahmad I, Rawat S, Sharma S. Indoblighcast- A simple generic DSS for late blight. In: *3rd international symposium on Phytophthora: Taxonomy, genomics, pathogenicity, resistance and disease management.* 9th-12th September, 2015. Bengaluru, India; c2015. p 32.
 69. Guo J, Lee TV, Qu DY, Yao YQ, Gong XF, Liang DL. *Phytophthora infestans* isolates from northern China show high virulence diversity but low genotypic diversity. *Plant Biol.* 2009;11(1):57-67.
 70. Gupta H, Singh BP, Jitendra M. Bio-control of late blight of potato. *Potato J.* 2004;31:39-42.
 71. Gutsche V. PROGEB- A model aided forecasting service

- for pest management in cereals and potatoes. *EPPO Bull.* 1993;23:577-581.
72. Halama P, Haluwin V. Antifungal activity of lichen extracts and lichenic acids. *Biocontrol.* 2004;49:95-107.
 73. Haldar K, Kamoun S, Hiller NL, Bhattacharje S, Van Ooij V. Common infection strategies of pathogenic eukaryotes. *Nature Review Microbiol.* 2006;4:922-931.
 74. Hannukkala A, Keskitalo M, Laamanen J, Rastas M. Control of potato late blight with caraway and dill extracts. In: (Schepers HTAM and C.E. *West-erdijk*, eds). Proceedings of the 6th workshop of a European network for development of an integrated control strategy of potato late blight. Edinburgh, Scotland, 26th- 30th September 2001. PPO-Special Report. 2002;8:279-280.
 75. Haverkort AJ, Struik PC, Visser RGF, Jacobsen E. Applied biotechnology to combat late blight in potato caused by *Phytophthora infestans*. *Potato Res.* 2009;52:249-264.
 76. He Q, Naqvi S, McLellan H, Boevink PC, Champouret N, Hein I. Plant pathogen effector utilizes host susceptibility factor NRL1 to degrade the immune regulator SWAP70. Proceedings of the National Academy of Sciences of the United States of America, 2018;115:E7834-E7843.
 77. Henshall WR, Shtienberg D, Beresford RM. A new potato late blight disease prediction model and its comparison with two previous models. *New Zealand Plant Protec.* 2006;59:150-154.
 78. Hohl HR, Iselin K. Strains of *Phytophthora infestans* from Switzerland with A2 mating type behaviour. *Trans. Br. Mycol. Soc.* 1984;83:529-30.
 79. Hospers-Brands AJTM, Ghorbani R, Bremer E, Bain R, Litterick A, Haldar F. Effect of pre-sprouting, planting dates, plant population and configuration of late blight and yield of organic potato crop grown with different cultivars. *Potato Res.* 2008;51:131-150.
 80. Imas P, Bansal SK.. Potassium and integrated nutrient management in potato. In: Global conference on potato. 6th-11th November, 1999, New Delhi, India. 1999
 81. Jetiyanon K, Kloepperm JW. Mixtures of plant growth promoting rhizobacteria for induction of systemic resistance against multiple plant diseases. *Biol. Control.* 2002;24:285-291.
 82. Jochemsen H. An ethical assessment of cisgenesis in breeding late blight resistant potato. *Potato Res.* 2008;51:59-73.
 83. Johnson DA, Alldredge JR, Vakoch DL. Potato late blight forecasting models for the semi-arid environment of South-Central Washington. *Phytopathol.* 1996;86:480-484.
 84. Jones GD. The epidemiology of plant diseases. 3rd edition. Kluwer Academic Publishes. London; c1998. p. 371-388.
 85. Kamoun S, Nusbaum C. Genomic sequence and analysis of the Irish potato famine pathogen *Phytophthora infestans*. *Nature.* 2009;461:393-398.
 86. Kamoun S, Smart CD. Late blight of potato and tomato in genomics era. *Plant Dis.* 2005;89:692-699.
 87. Kassa B, Sommartha T. Effect of intercropping of potato late blight *Phytophthora infestans* (Mont.) de Bary development and potato tuber yield in Ethiopia. *Kasetsar J (Nat. Sci.).* 2006;40:914-924.
 88. Kaya HK, Aguillera MM, Alumai A, Choo HY, Dela Torre M, Fodor A. Status of entomopathogenic nematodes and their symbiotic bacteria from selected countries or regions of the world. *Bio. Control.* 2006;38:134-155.
 89. Keane T. Potato blight warning practice in Ireland. In: *Phytophthora 150.* (Dowley, L.W *et al* eds), Boole Press Ltd. Dublin. 1995. p. 191-200.
 90. Keskitalo M, Fabritius AL, Hakala K, Hannukkala A, Ketoja E, Lehtinen A. Control of potato late blight by caraway oil in organic farming. In: Nordiska Jordbruksforskarens Frening NJF-Seminar 369 on organic farming for a new millennium status and future challenges. 2005, p. 77-79.
 91. Kim HJ, Jeun YC. Resistance induction and enhanced tuber production by pre-inoculation with bacterial strains in potato plants against *Phytophthora infestans*. *Mycobiol.* 2006;34:67-72.
 92. Kim Y, Lee C, Kim H, Lee H. Anthraquinones isolated from *Cassia tora* (leguminosae) seed show an antifungal properties against Phytopathogenic fungi. *J Agricul. and Food Chem.* 2004;52:6096-6100.
 93. Kirk KK, Felcher KJ, Douches DS, Coombs J, Stein JM, Baker KM. Effect of host resistance and reduced rates and frequencies of fungicide application to control potato late blight. *Plant Dis.* 2001;85(10):1113-1118.
 94. Kirk W, Wharton P, Hammerschmidt R, Abuel Samen F, Douches D. Late blight. Michigan state university extension bulletin E-2945. East Lansing; c2013. MI. Available on: <http://www.potatodiseases.org/lateblight.html>
 95. Kirk W. Potato late blight alert for the mid west. Field crop advisory team alert. *Current News Articles.* 2009.
 96. Kirk WW, Samen F, Abu EL, Thumbalam P, Wharton P, Douehes D. Impact of different US genotypes of *Phytophthora infestans* on potato seed tuber rot and plant emergence in a range of cultivars and advanced breeding lines. *Potato Res.* 2009;52:121-140.
 97. Krause RA, Massie LB, Hyre RA. BLITECAST: A computerized forecast of potato late blight. *Plant Dis. Repr.* 1975;59:95-98.
 98. Kumaon S. Non-host resistance to *Phytophthora*: Novel prospects for a classical problem. *Curr. Opin. Plant Biol.* 2001;4:295-300.
 99. Lamichhane JR, Osdaghi E, Behlau F, Köhl J, Jones JB, Aubertot JN. Thirteen decades of antimicrobial copper compounds applied in agriculture. A review. *Agronomy for Sustainable Development.* 2018;38:28.
 100. Lamsal K, Kim SW, Kim YS, Lee YS. Bio-control of late blight and plant growth promotion in tomato using rhizobacterial isolates. *J Microbiol. Biotech.* 2013;23:885-892.
 101. Lee SK, Kelley BS, Damasceno CMB, St. John B, Kim B, Kim BD. A functional screen to characterize the secretomes of eukaryotic pathogens and their hosts in planta. *Molecular Plant Microbe Interactions,* 2006;19:1368-1377.
 102. Lehtinen A, Hannukkala A. Oospores of *Phytophthora infestans* in soil provide an important new source of primary inoculum in Finland. *Agricultural and Food Science in Finland.* 2004;13:399-410.
 103. Lozoya-Saldana H, Coyote-Palma MH, Ferrera-Cerrato R, Lara-Hernaandez ME. Microbial antagonism against *Phytophthora infestans* (Mont) de Bary. *Agrociencia.*

- 2006;40:491-499.
104. Lucy M, Reed E, Glick BR. Application for free living plant growth promoting rhizobacteria. *Antonie Van Leeuwenhoek*. 2004;86:1-25.
 105. Mari M, Bertolini P, Pratella G. Non-conventional methods for the control of post-harvest pear diseases. *J. Applied Microbiol.* 2003;94:761-766.
 106. Marschner H. Mineral nutrition on higher plants. 2nd Edition, Academic Press, London; c1995.
 107. Martin AD, Gary AS, Neil CG, Arthur HL, Duane P. Leaf blight diseases of potato. North Dakota State University Agriculture and University Extension; c1994.
 108. Minogue KP, Fry WE. Models for the spreads of disease: Some experimental results. *Phytopathol.* 1983;73:1173-1176.
 109. Naumann M, Koch M, Thiel H, Gransee A, Pawelzik E. The Importance of Nutrient Management for Potato Production Part II: Plant Nutrition and Tuber Quality. *Potato Research*. 2020;63:121-137.
 110. Njualedem DK, Demo P, Mendoza HA, Koi JT, Nana SF. Reaction of some potato genotypes to late blight in Cameroon. *African Crop Sci. J.* 2001;1(1):209-213.
 111. O'herlihy EA, Duffy EM, Cassells AC. The effects of arbuscular mycorrhizal fungi and chitosan sprays on yield and late blight resistance in potato crops from micro-plants. *Folia Geobotanica*. 2003;38:201-207.
 112. Pande A, Jagyasi BG, Choudhuri R. Late blight forecast using mobile phone based agro advisory system. (Choudhary *et al* eds). Springer-Verlag Berlin Heidelberg; c2009, p. 609-614.
 113. Paysour RE, Fry WE. Inter plot interference: A model for planning field experiments with aerially disseminated pathogen. *Phytopathol.* 1983;73:1014-1020.
 114. Phadtare SG, Dutt BL, Dhingra MK, Raj S. A new race of *Phytophthora infestans* from Shimla hills. *Indian Phytopath.* 1973;26:589-590.
 115. Popokova KV. Late blight of potato in Moscow. Review of crop production research in Ethiopia. In: (Tsedeke Abate ed.). Proceedings of the 1st Ethiopian crop protection symposium. Institute of Agricultural Research. Addis Ababa. Ethiopia; c1972.
 116. Poudel A, Pandey M, Shah K, Acharya B, Shrestha J. Evaluation of fungicides for management of late blight (*Phytophthora infestans*) of potato. *Agrica*. 2020;9:10-17.
 117. Rashidul Islam M, Mandal C, Hossain I, Meah BM. Organic management: An alternative to control late blight of potato and tomato caused by *Phytophthora infestans*. *Int. J Theoretical and Applied Sci.* 2013;5:32-42.
 118. Rivera-Pena A. *Phytophthora infestans* European association for potato research, Pathology section conference, Durbin, Ireland; c1995. p. 116-121.
 119. Scheuerell S, Mahaffee W. Compost tea: Principles and prospects for plant disease control. *Compost Science and Utilization*. 2002;10:313-338.
 120. Scheuerell SJ, Mahaffee WF. Variability associated with suppression of gray mould (*Botrytis cinerea*) on geranium by foliar applications of non-aerated and aerated compost teas. *Plant Dis.* 2006;90:1201-1208.
 121. Schick R. Uber das verhalten von *Solanum demissum*, *Solanum tuberosum* und ihren Basterden gegenuber verschiedenen Herkunftjen von *Phytophthora infestans*. *Zuechter*. 1932;4:233-237.
 122. Seong K, Seo E, Witek K, Li M, Staskawicz B. Evolution of NLR resistance genes with noncanonical N-terminal domains in wild tomato species. *New Phytologist*. 2020;227:1530-1543.
 123. Shailbala, Pundhir VS. Effect of date of planting on late blight severity and yield of potato. *Ann. Pl. Protec. Sci.* 2006;14:404-406.
 124. Shailbala, Pundhir VS. Effect of date of planting and fungicidal sprays on potato phylloplane fungi. *Ann. Pl. Protec. Sci.* 2007;15(2):434-437.
 125. Shailbala, Pundhir VS. Effect of potato mustard intercropping on late blight severity, yield and economics of potato. *Plant Dis. Res.* 2007;22(1):67-69.
 126. Shailbala, Pundhir VS. Integration of host resistance and fungicides for management of late blight of potato. *Potato J* 2008.35(1-2): 97-99.
 127. Shailbala, Pundhir VS. Efficacy of fungicides and bio-agents against late blight severity, infection rate and tuber yield of potato. *J Pl. Dis. Sci.* 2008;3(1):4-8.
 128. Shanmugam V. Epidemiology and management of late blight of potato. M.Sc. Thesis, G.B.P.U.A.T., Pantnagar. 2001, p. 83.
 129. Sharma KK. Influence of meteorological factors on potato late blight development in North-Western plains of India. *J Indian Potato Assoc.* 2000;27:1-3.
 130. Shekhawat GS. Management of potato diseases through host resistance. *J Mycol. Pl. Pathol.* 2000;30(2):143-150.
 131. Shinnars CT, Bains P, McLaren D, Thomson J. Commercial potato production disease management. Guide to commercial potato production prairies. Western Potato Council. 2003. Available on: <http://www.gov.mb.ca/agriculture/crops/potatoes/bda04s07>.
 132. Shrestha, J, Shah KK, Timsina K. Effects of different fertilizers on growth and productivity of rice (*Oryza sativa* L.): A review. *International Journal of Global Science Research*. 2020;7(1):1291-1301.
 133. Shtienberg D, Raposo R, Bergerson SN, Legard DE, Dyer AT, Fry WE. Inoculation of cultivar resistance reduced spray strategy to suppress early and late blight on potato. *Plant Dis.* 1994;78:23-26.
 134. Shutong Wong, Tongle Hu, Fengqiao Z, Forrer HR, Keqiang Cao. Screening for plant extracts to control potato late blight. *Frontiers Agric. China*, 2007;1:43-46.
 135. Singh BP, Gupta H, Roy S, Shekhawat GS. Ploidy status and its role in aggressiveness of *Phytophthora infestans*. Abstracts (518-006). Indian Phytopathological Society-Golden Jubilee International Conference, 10th-15th November, 1997, New Delhi; c1997.
 136. Singh BP, Islam Ahmed, Sharma VC, Shekhawat GS. JHULSACAST: A computerized forecast of potato late blight in Western Uttar Pradesh. *J Indian Potato Assoc.* 2000;27:25-34.
 137. Singh BP, Roy S, Bhattacharyya SK. Occurrence of A2 mating type of *Phytophthora infestans* in India. *Potato Res.* 1994;37:227-231.
 138. Singh PH, Singh BP, Bhat NM. Mating types, metalaxyl resistance and racial complexity in *Phytophthora infestans* population-present status. *Potato J.* 2005;32:177-178.
 139. Singh BP. Status of late blight in sub tropics. In: Potato global research and development (Khurana SMP, Shekhawat, GS, Singh BP and Pandey SK. eds.). Indian Potato Association. CPRI, Shimla, H.P. India. 2000, p. 525-533.

140. Smith LP. Potato blight forecasting by 90% humid ity criteria. *Plant Path.* 1956;5:83-87.
141. Song J, Bradeen JM, Naess SK, Raasch JA, Wielgus SM, Haberlach GT. Gene RB cloned from *Solanum bulbocastanum* confers broad spectrum resistance to potato late blight. *Proceeding of the National Academy of Sciences of U.S.A.* 2003;100(16):9128-9133.
142. Soyong K, Ratanacherdchai R. Application of mycofungicide to control late blight of potato. *J. Agricultural Technology.* 2005;1:19-32.
143. Struik PC, Wiersema SG. Seed potato technology. Wageningen Pers, Wageningen; c1999.
144. Struik PC. Can physiology help us to combat late blight in Potato? *Potato Res.* 2010;53:277-287.
145. Sujkowski LS, Goodwin SB, Dyer AT, Fry WE. Increased genotypic diversity via migration and possible occurrence of sexual reproduction of *Phytophthora infestans* in Poland. *Phytopathol.* 1994;84:201-207.
146. Sundaresha S, Kumar S, Singh BP, Jeevalatha A, Rawat S, Mahota AK. Comparative genome analysis of Irish famine pathogen with Indian *Phytophthora infestans* isolate. In: 3rd International symposium on Phytophthora: Taxonomy, genomics, pathogenicity, resistance and disease management. 9th-12th September, 2015. Bengaluru, India; c2015. p. 24.
147. Suwan S, Isobe M, Kanokmedhakul S, Lourit N, Kanokmedhakul K, Soyong K. Elucidation of high micro-heterogeneity of an acidic-neutral trichotoxin mixture from *Trichoderma harzianum* by electrospray ionization quadrupole time-of-flight mass spectrometry. *J. Mass Spectrometry*, 2000;35:1438-1451.
148. Tian M, Win J, Song J, Van Der Hoorn R, Van Der Knaap E, Kamoun S. A *Phytophthora infestans* cystatin-like protein targets a novel tomato papain like apoplastic protease. *Plant Physio.* 2007;143:364-377.
149. Tiwari I, Shah KK, Tripathi S, Modi B, Subedi S, Shrestha J. Late blight of potato and its management through the application of different fungicides and organic amendments: a review. *Journal of Agriculture and Natural Resources.* 2021;4(1):301-320.
150. Tripathi M, Dubey NK, Shukla AK. Use of some essential oils as post-harvest botanical fungicides in the management of grey mould of grapes caused by *Botrytis cinerea*. *World J Microbiol. Biotechnol.* 2008;24:39-46.
151. Tsedaley B. Late blight of potato (*Phytophthora infestans*) biology, economic importance and its management approaches. *J Biology Agriculture and Healthcare.* 2014;25(4):215-225.
152. Turner RS. After the famine: Plant pathology, *Phytophthora infestans* and the late blight of potatoes, 1845-1960. *Historical Studies in the Physical and Biological Sciences.* 2005;35(2):341-370.
153. Ullrich J, Schrodter H. Das Problem der vorher- sage des aufretens der Kartoffelkrautfaule (*Phytophthora infestans*) und die möglichkeit seiner losung durch eine negative prognose. *Nachrichtenblatt Dt. Pflanzenschutzdienst (Braunschweig).* 1966;18:33-40.
154. Van de Weyer AL, Monteiro F, Furzer OJ, Nishimura MT, Cevik V, Witek K. A Species-Wide Inventory of NLR Genes and Alleles in *Arabidopsis thaliana*. *Cell.* 2019;178:1260-1272 e1214.
155. Van Everdingen E. Hetverband tusschen de weergesteldhied en de aarolppelziekte, *Phytophthora infestans* (the relation between weather conditions and potato blight, *Phytophthora infestans*) *Tijdschr. Plantenziekten.* 1926;32:129-140.
156. Van Oijen M. Selection and use of a mathematical model to evaluate components of resistance to *Phytophthora infestans* in potato. *Netherland J Plant Path.* 1992;98:192-202.
157. Visker MHPW, Heilersig HJB, Kodde LP, Van de Weg WE, Voorrips RE, Struik PC. Genetic linkage of QTLs for late blight resistance and foliage maturity type in six related potato progenies. *Euphytica.* 2005;143:189-199.
158. Visker MHPW, Keizer LCP, Budding DJ, Van Loon LC, Colon LT, Struik PC. Leaf position prevails over plant age and leaf age in reflecting resistance to late blight in potato. *Phytopathol.* 2003b;93:666-674.
159. Visker MHPW, Keizer LCP, Van Eck HJ, Jacobsen E, Colon LT, Struik PC. Can the QTL for late blight resistance on potato chromosome 5 be attributed to foliage maturity type? *Theor. Appl. Gen.* 2003a;106:317-325.
160. Visker MHPW, Van Raaij HMG, Keizer LCP, Struik PC, Colon LT. Correlation between late blight resistance and foliage maturity type in potato. *Euphytica.* 2004;137:311-323.
161. Waggoner PE. Weather and rise and fall of fungi. (Lowry, W.P eds). Oregon State, Corvallis. USA; c1968. p. 45-66.
162. Wallin JR. Summary of recent progress in predicting late blight epidemics in United States and Canada. *Am. Potato J.* 1962;39:306-312.
163. Wang W, Daniel B, Cohen Y. Control of plant diseases by extracts of *Inula viscosa*. *Phytopathol.* 2004;94:1042-1047.
164. Wharton PS, Kirk WW, Baker KM, Duynslager L. A web-based interactive system for risk management of potato late blight in Michigan. *Computers and Electronics in Agriculture.* 2008;61:136-148.
165. Widmark, Anna-Karin. The late blight pathogen, *Phytophthora infestans*: Interaction with the potato plant and inoculum sources. Thesis. Swedish University of Agricultural Sciences, Uppsala; c2010. p. 67.
166. Yan Z, Reddy MS, Ryu CM, Mcinroy JA, Wilson M, Kloepper JW. Induced systemic protection against tomato late blight elicited by plant growth- promoting rhizobacteria. *Phytopathol.* 2002;92:1329-1333.
167. Yang X, Zhang Z, Yang H, Jian H. Inhibition of metabolites from *Xenorhabdus nematophilus* against *Phytophthora infestans*. *J Hebei Agric. Univ.* 2001;24:65-68.