



ISSN (E): 2277-7695
ISSN (P): 2349-8242
NAAS Rating: 5.23
TPI 2022; 11(12): 4105-4108
© 2022 TPI
www.thepharmajournal.com
Received: 05-10-2022
Accepted: 08-11-2022

D Prathyusha
Ph.D. Scholar, Department of
Plant Pathology, ANGRAU
Agricultural College, Bapatla
Andhra Pradesh, India

AK Patibanda
Professor, Department of Plant
Pathology, ANGRAU, SMGR
Agricultural College, Udayagiri,
Nellore, Andhra Pradesh, India

V Prasanna Kumari
Professor and Head, Department
of Plant Pathology, ANGRAU,
Agricultural College, Bapatla,
Andhra Pradesh, India

P Madhusudhan
Senior Scientist, Department of
Plant Pathology, ANGRAU
ARS, Nellore, Andhra Pradesh,
India

K Jayalalitha
Principal Scientist, RRU (Crop
Physiology), ANGRAU RARS,
LAM, Guntur, Andhra Pradesh,
India

V Srinivasa Rao
Professor and University Head
Department of Statistics and
Computer Applications,
ANGRAU, Agricultural College
Bapatla, Andhra Pradesh, India

Corresponding Author:
D Prathyusha
Ph.D. Scholar, Department of
Plant Pathology, ANGRAU
Agricultural College, Bapatla,
Andhra Pradesh, India

Evaluation of different single and combination fungicides against *in vitro* growth of *Pyricularia oryzae* causing rice blast disease

D Prathyusha, AK Patibanda, V Prasanna Kumari, P Madhusudhan, K Jayalalitha and V Srinivasa Rao

Abstract

Rice blast disease caused by *Pyricularia oryzae* is a major threat to the farmers as the disease severity may lead up to 80% crop loss in severe conditions. A laboratory experiment was conducted at Department of Plant Pathology, Acharya N.G. Ranga Agricultural University, Agricultural College, for testing the efficacy of eight different single and combination fungicides adopting poisoned food method against the pathogen *Pyricularia oryzae* causing rice blast disease. The experiment was carried out in a completely randomized design with eight treatments including control following 3 replications of each. Three concentrations (R – recommended dose; R/2 and 2R) each of different fungicides viz., Beam (Tricyclazole 75% WP @ 0.06%), Amistar (Azoxystrobin 25% SC @ 0.10%), Fujione (Isoprothiolane 5% EC @ 0.15%), Kasugamycin @ 0.25%, Folicur (Tebuconazole 25% EC @ 0.15%), Nativo 75% WG (Trifloxystrobin 50% + Tebuconazole 25% WG @ 0.08%), Custodia (Azoxystrobin + Tebuconazole @ 0.15%) and Folia (Propiconazole + Tricyclazole @ 0.10%) were evaluated against the radial growth of the pathogen. Results showed that except two fungicides viz., Kasugamycin and Azoxystrobin rest all the fungicides fully inhibited (100% inhibition) the mycelial growth of the pathogen at all the three different concentrations compared to control.

Keywords: Rice, *Pyricularia oryzae*, Fungicides, Poisoned Food Technique

Introduction

Rice (*Oryza sativa* L.) is the second most important cereal cultivated globally and has been nourishing more than half of the world's population (Singh and Singh, 2019). India is the second largest producer (110.15 MT) of rice in the world while Andhra Pradesh is one among the leading states where huge production (7.49 MT) is achieved. Of all the biotic and abiotic factors that impact the yield, leaf blast disease caused by *Pyricularia oryzae* is of significant importance which can scale down the yield with 80% losses depending on the crop stage it's going to affect (Neupane & Kiran Bhusal, 2021) [9]. It is polycyclic, hemibiotrophic foliar pathogen, prevalent globally in humid areas where the rice is being cultivated (Neelakanth *et al.*, 2017) [7]. Seed borne nature and its capacity to infect all crop stages can impose severe devastation in crop. Cultural practices like use of resistant varieties, proper fertilizer application and weed management besides biological and chemical control measures offer the reliable disease control against leaf blast. The pathogen is highly variable and numerous pathotypes were reported so far. As the resistant cultivars are frequently being susceptible to the developing pathotypes, fungicide spraying is inevitable for efficient control of leaf blast where the resistance in crop is lacking (Groth, 2006; Morton and Staub, 2008; Pooja and Katoch, 2014) [2, 6, 10]. The present study was deployed to evaluate the efficacy of single and novel combinations of fungicide against rice blast.

Material and Methods

A total of eight fungicides (Table 1) at three different concentrations (recommended (R), R/2 and 2R) were evaluated *in vitro* against *P. oryzae* by poisoned food technique (Nene and Thapliyal, 1993). Fungicide stock solution of 100000 ppm was prepared initially by dissolving 1 g of fungicide in 10 ml of sterile distilled water. Later, working standards of desired concentration of the poisoned media was made by using the formula $C_1V_1 = C_2V_2$

Where,

C_1 = concentration of the stock solution (ppm)

V_1 = volume of the stock solution to be added (ml)

C_2 = desired concentration (ppm)

V_2 = volume of PDA in which fungicide is to be amended (ml)

The poisoned medium (20ml) of the required concentration was poured in sterile petri plates under laminar air flow in triplicate and allowed to solidify. Culture discs of 5 mm diameter cut from the periphery of actively growing mycelium (seven days old) were inoculated in the centre of each petriplate under aseptic conditions and incubated at 28 ± 1 °C in a BOD incubator. Unamended PDA plate inoculated

with *P. oryzae* served as check. Radial growth of the test fungus was recorded 14 days after inoculation. Percent inhibition of growth over check was calculated using the formula given by Vincent (1927).

$$I = \frac{C - T}{C} \times 100$$

Where,

I = percent inhibition.

C = growth of *P. Oryzae* in unamended medium.

T = growth of *P. Oryzae* in amended medium.

Table 1: Details of Treatments for *in vitro* evaluation of fungicides

Treatments	Fungicides	Trade Name	Recommended Conc (R)	R/2	2R
T ₁	Tricyclazole	Beam	0.06%(600 ppm)	0.03%(300 ppm)	0.12%(1200 ppm)
T ₂	Azoxystrobin	Amistar	0.10%(1000 ppm)	0.05%(500 ppm)	0.20%(2000 ppm)
T ₃	Kasugamycin	Kasu B	0.25(2500 ppm)	0.125(1250 ppm)	0.5(5000 ppm)
T ₄	Isoprothiolane	Fuji-one	0.15%(1500 ppm)	0.075%(750 ppm)	0.30%(3000 ppm)
T ₅	Tebuconazole	Folicur	0.15%(1500 ppm)	0.075%(750 ppm)	0.30%(3000 ppm)
T ₆	Trifloxystrobin + Tebuconazole	Nativo	0.08(800 ppm)	0.04(400 ppm)	0.16(1600 ppm)
T ₇	Azoxystrobin 11% + tebuconazole 18.3 % SC (0.15%)	Custodia	0.15%(1500 ppm)	0.075%(750 ppm)	0.30%(3000 ppm)
T ₈	Propiconazole + Tricyclazole	Filia	0.10%(1000 ppm)	0.05%(500 ppm)	0.20%(2000 ppm)
T ₉	Control				

Result and Discussions

Upon evaluation of fungicides mentioned in Table 2, a significant reduction in mean radial growth of the pathogen was noticed with the increasing concentration. All the treatments except for Kasugamycin and Azoxystrobin, tested at three concentrations were found effective over the control with cent % inhibition of the pathogen.

Decrease in mycelial growth with the increased concentration was noticed in both Kasugamycin and Azoxystrobin fungicide treatments. At half of the recommended concentration (R/2), least percent inhibition of 67.44% was noticed in Azoxystrobin where the maximum radial growth of 2.93 cm was observed, followed by Kasugamycin with 84.11% inhibition (1.43 cm).

Table 2: *In vitro* efficacy of fungicides against *Pyricularia oryzae*

S. No.	Treatments	Dose R (%)	Colony diameter (cm)* and Percent inhibition over control					
			R/2		R		2R	
			Radial growth (cm)	Percent inhibition over control	Radial growth (cm)	Percent inhibition over control	Radial growth (cm)	Percent inhibition over control
1	Tricyclazole 75% WP	0.06	0.00(1.00)	100.00	0.00(1.00)	100.00	0.00(1.00)	100.00
2	Azoxystrobin 25% SC	0.10	2.93(1.98)	67.44	2.60(1.89)	71.11	2.50(1.87)	72.22
3	Isoprothiolane 5% EC	0.15	0.00(1.00)	100.00	0.00(1.00)	100.00	0.00(1.00)	100.00
4	Kasugamycin 3% SL	0.25	1.43(1.56)	84.11	1.30(1.51)	85.55	0.80(1.34)	91.11
5	Tebuconazole 25% EC	0.15	0.00(1.00)	100.00	0.00(1.00)	100.00	0.00(1.00)	100.00
6	Trifloxystrobin 50% + Tebuconazole 25% WG	0.08	0.00(1.00)	100.00	0.00(1.00)	100.00	0.00(1.00)	100.00
7	Azoxystrobin 11% + Tebuconazole 18.3% SC	0.15	0.00(1.00)	100.00	0.00(1.00)	100.00	0.00(1.00)	100.00
8	Propiconazole 10.7% + Tricyclazole 34.2% SE	0.10	0.00(1.00)	100.00	0.00(1.00)	100.00	0.00(1.00)	100.00
	Mean	-	0.48		0.47		0.46	
9	Control		9.00 (3.16)					
	SEM ±		0.01		0.01		0.01	
	C.D. ($p \leq 0.05$)		0.02		0.03		0.02	
	CV%		1.40		1.27		1.13	

❖Means of three replications

*Values in parenthesis are square root transformed values

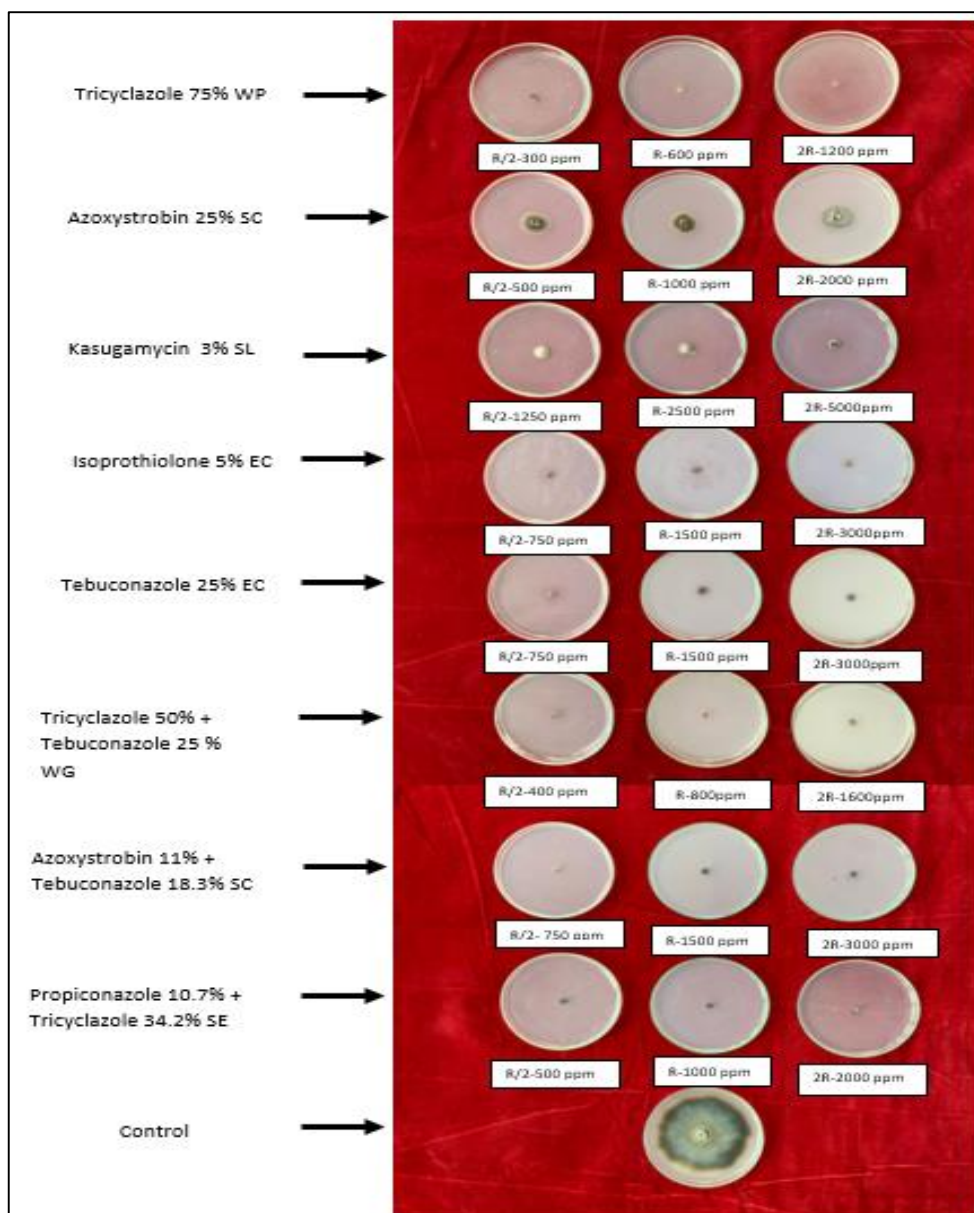


Fig 1: Efficacy of chemicals on radial growth of *P. oryzae* in vitro

Though, at the recommended concentration (R) complete inhibition of the pathogen was not noticed in treatments of Kasugamycin and Azoxystrobin. The maximum inhibition of 85.55% was observed in Kasugamycin (1.30 cm), while Azoxystrobin found to be the least effective with 71.11% inhibition over the control. However at twice the dose of recommendation (2R), Azoxystrobin and Kasugamycin was continued to be the least performing among all with 72.22% and 91.11% inhibition respectively, compared to control (Table 2; Fig 1).

The present findings are in agreement with the results of Kavanashree *et al.*, 2019; Kulmitra *et al.*, 2017 [5]; Neelakanth *et al.*, 2017 [7]; Kafle *et al.*, 2021; Singh *et al.*, 2019 [11]; Vidhyashankar *et al.*, 2020 [12]. Ergosterol, being the essential for fungal cell wall structure can cause deleterious effect on cell wall integrity if its biosynthesis was inhibited (Dupont *et al.*, 2012) [1]. Majority of the fungicides which showed complete inhibition of mycelia growth were of triazole group members. They proved to specifically inhibit sterol biosynthesis which is essential for maintaining cell wall integrity.

Conclusion

In the present study, the efficacy of recent fungicides is explored for their potential against the rice blast pathogen. Novel fungicide combinations which can control the pathogen efficiently with least dosages can offer multiple sites of action, exhibiting least chances of resistance development. Fungicide formulations of such *viz.* Trifloxystrobin + Tebuconazole (Nativo), Azoxystrobin + Tebuconazole (Custodia) and Propiconazole + Tricyclazole (Filia) can be exploited for the rice blast control even at half of the recommended doses.

Conflict of Interest: None declared

References

- Dupont S, Lemetais G, Ferreira T, Coyot P, Gervais. Ergosterol biosynthesis: A fungal pathway for life on land. *Evolution*. 2012 Sep;66(9):2961-2968.
- Groth DE. Azoxystrobin rate and timing effects on rice head blast incidence and rice grain and milling yields. *Plant Disease*. 2006 Aug;90(8):1055-1058.

3. Kafle K, Abichal P, Shrinkhala M, Nirmal A. Efficacy of different fungicides against the *in-vitro* growth of *Pyricularia oryzae* causing Rice blast disease. International Journal of Environment, Agriculture and Biotechnology. 2021;6:3.
4. Kavanashree K, Ramanathan A, Darshan K. *In vitro* evaluation of single and combined formulations of the fungicides against rice blast caused by *Magnaporthe oryzae* Cav. International Journal of Chemical Studies. 2019;7(5):1072-1076.
5. Kulmitra AK, Sanath Kumar VB, Thejesha AG, Ghosh A, Sahu P. *In vitro* evaluation of fungicides against *Pyricularia oryzae* (Cav.) causing rice blast disease. International Journal of Chemical Studies. 2017;5(4):506-509.
6. Morton V, Staub T. A Short History of Fungicides. Online, APS net Features; c2008.
7. Neelakanth, Sidde Gowda DK, Chethana BS, Parasappa HH. *In vitro* and *In vivo* Evaluation of Fungicides against *Pyricularia oryzae* Causing Blast of Rice. International Journal of Pure and Applied Bioscience. 2017;5:3.
8. Nene YL, Thapliyal PN. Poison food technique. *Fungicides in Plant Disease Control* (2nd Edition). Oxford and IBH publication, New Delhi, India; c1993. p. 413-415.
9. Neupane N, Kiran Bhusal. A Review of Blast Disease of Rice in Nepal. Journal of Plant Pathology and Microbiology. 2021;12:1.
10. Pooja K, Katoch A. Past, present and future of rice blast management. Plant Science. Today. 2014;1:165-173.
11. Singh HS, Kaushik SS, Chauhan MS, Negi, RS. Efficacy of different fungicides against rice blast caused by *Pyricularia oryzae* (Cav.) under field condition in satna district of Madhya Pradesh. International Journal of Current Microbiology and Applied Sciences. 2019;8(06):63-69.
12. Vidyashankar D, Kamanna BC, Nagaraju, P. Bio-Efficacy of Fungicides against *Magnaporthe oryzae* Causing Blast of Rice. International Journal of Current Microbiology and Applied Science. 2020;9(3):3042-3047.
13. Vincent JM. Methods for the study of their fungi static properties. Journal of the Society of the Chemical Industry London. 1927;16:746-755.