www.ThePharmaJournal.com

# The Pharma Innovation



ISSN (E): 2277-7695 ISSN (P): 2349-8242 NAAS Rating: 5.23 TPI 2022; 11(12): 5538-5540 © 2022 TPI

www.thepharmajournal.com Received: 22-10-2022 Accepted: 30-11-2022

#### **GP** Jagtap

Associate prof. Department of plant pathology, College of Horticulture, VNMKV, Parbhani, Maharashtra, India

#### **AR** Sawate

Department of plant pathology, College of Agriculture, V.N.M.K.V., Parbhani, Maharashtra, India

#### SJ walke

Department of plant pathology, College of Agriculture, V.N.M.K.V., Parbhani, Maharashtra, India

Corresponding Author: GP Jagtap Associate prof. Department of plant pathology, College of Horticulture, VNMKV, Parbhani, Maharashtra, India

# To evaluate the *in vitro* efficacy of different bioagents against *Pyricularia grisea* (Cooke) Sacc.

# GP Jagtap, AR Sawate and SJ walke

#### Abstract

Biotic and abiotic stresses are major constraints in the production of Pearl millet. Among biotic stresses apart from bacterial and viral diseases, many fungal diseases are of economic importance. Blast was the most important and destructive disease of Pearl millet. Blast caused by *Pyricularia grisea* a serious threat to successful cultivation of Pearl millet. For the management of Blast of bajra, an experiment was conducted to study the efficacy of antagonistic organism against *Pyricularia grisea*. The bio-agents i.e. *Trichoderma viride*, *T harzianum*, *T hamatum*, *T.gliocladium*, *T. konigii*, *Peudomonas fluorescens* and *Bacillus subtilis* etc. were evaluated *in vitro*, found antifungal to *Pyricularia grisea*. However, *P. fluorescens* was found most significant with highest mycelial growth inhibition (64.61%) of the test pathogen. The second and third inhibitoriest antagonists found were *T. asperllum* and *T. harzianum* with mycelial growth inhibition of 62.39 and 57.03%, respectively.

Keywords: Biocontrol, Claviceps fusiformis, in vitro, inhibition

#### Introduction

Pearl Millet [*Pennisetum glaucum* (L.) R. Br)] is one of the assured *Kharif* crop under environment domesticated in the annual rainfall of 150 mm to 1000 mm in India. It has variety of uses for the consumption of human being such as chapatti, breads, snacks, cakes, beverages and predigested weaning food besides poultry feed and fodder for cattle. India and Africa are together occupying 90 per cent area of total pearl millet production in the world (Yadav *et al.*, 2012) <sup>[9]</sup>. It's grain is chiefly served as a food, because of high protein (27 to 32%), higher concentration of essential amino acids, twice the extract (fat) and higher gross energy than maize (Ejeta *et al.*, 1987; Davis *et al.*, 2003) <sup>[3, 2]</sup>. Number of pathogens attack Pearl Millet during its crop growth period, cause low yield and economic loss to the present growers. Downy mildew, blast, ergot, smut and rust are the major diseases of the crop.

Blast incited by *Pyricularia grisea* (Cooke) Sacc. Was first recorded in Uganda in 1933 (Emechebe, 1975)<sup>[4]</sup>. The disease has geographic distribution in India, Singapore on Napier grass and the United States (Buckley and Allen, 1951)<sup>[1]</sup>. Blast incited by *Pyricularia grisea* was first reported in India from Kanpur 1953 (Mehta *et al.*, 1953)<sup>[5]</sup> and remain as minor disease till the end of 20<sup>th</sup> century but from last one decade the disease has occupied a key position among the Pearl Millet diseases particularly in Maharashtra. The disease appears as grayish, water-soaked foliar lesions that enlarge and become necrotic, resulting in extensive chlorosis and pre mature drying of young leaves (Wilson *et al.*, 1989)<sup>[8]</sup>.

Management of disease through resistant cultivars is the most economical and relevant way of controlling pearl millet blast mainly by resource poor and marginal farmers, who cannot afford to control blast disease by the application of chemical. Chemical control of plant pathogens is most effective and may cause hazardous effects on the environment and their residues on grain and fodder may also cause the harmful effect on human being, animals and birds. Hence in search of an alternative to the chemicals there is a need to test the bio - agents and others for the management of the disease. Keeping the above points in mind the present experiments have been carried out.

Use of resistant cultivars is the best alternative to overcome yield losses caused by *Pyricularia grisea*. It has the ability to overcome resistance within two to three years after the release of resistant variety and made breeding for resistance a constant the pathogen since major studies were carried out on rice blast pathosystems (Sere *et al.*, 2007) <sup>[6]</sup>. However such studies are very limited with the pearl millet blast pathosystem (Sharma *et al.*, 2013) <sup>[7]</sup>. Severe infection of the disease reduced the yield particularly fodder value hence there is a need to work on the line of management of the disease.

#### **Material and Method**

In dual culture technique, twenty ml of sterilized and cooled potato dextrose agar was poured into sterile Petri plates. Fungal antagonists were evaluated by inoculating the pathogen at one side of Petri plate and the antagonist inoculated at exactly opposite side of the same plate by leaving 3 cm gap. For this actively growing cultures were used. In case of bacterial antagonist evaluation, two mycelial discs of pathogen was inoculated and bacterial antagonist was streaked in the center of the plate. Each treatment was replicated three times. After required period of incubation i.e., in the control plate growth reached 90 mm diameter, the radial growth of pathogen was measured.

Per cent inhibition over control was calculated by applying the formula given.

 $R = \{(C - T) / C\} \times 100$ 

#### Where

R= Per cent inhibition,

C = Radial growth of pathogen colony in control and

T = Radial growth of pathogen colony in treatment

#### **Details of experiment**

Design: CRD Replications: Three Treatments: Eight

# **Treatment details**

In vitro evaluation of bioagents against Pyricularia grisea

Treatment	Bioagents
$T_1$	Pseudomonas fluorescens
$T_2$	Trichoderma harzianum
T3	Trichoderma viride
$T_4$	Trichoderma gliocladium
T5	Trichoderma hamatum
$T_6$	Trichoderma koningi
<b>T</b> <sub>7</sub>	Bacillus subtilis
T8	Control

Treatment details

Observations on radial mycelial growth of the test fungus and bio control were recorded at 24 hrs interval and were continued till growth of the test pathogen in untreated control plate was fully covered. Per cent inhibition of the test pathogen was calculated by applying formula given.

 $R = \{(C - T) / C\} \times 100$ 

# Where,

R= Per cent inhibition

C= growth of the test pathogen in untreated Control plates

T= growth of the test pathogen a in treated plates

#### Results

In this study, pure culture of bio-agents *Pseudomonas* fluorescens Trichoderma viride, Trichoderma harzianum, Trichoderma (Gliocladium) virens, Trichoderma hamatum, Trichoderma koningi and Bacillus subtilis obtained from Department of Plant Pathology, VNMKV, Parbhani,

(Maharashtra). In the present study, the effects of six known

https://www.thepharmajournal.com

antagonists were tested against P. grisea by dual culture.

<b>T T T T T T T T T T</b>	CC'	C1 · ·	• .	D · 1 ·	•
Table 1: In vitro	efficacy c	of bloagents	against	Pvricularia	grisea
	erriede) (	or orougenes	agamot		0.1000

Sr. No	Treatments	Colony Dia*(mm)	% Inhibition
T <sub>1</sub>	Pseudomonas fluorescens	32.00	64.61 (53.49)
T <sub>2</sub>	Trichoderma harzianum	38.67	57.03 (49.04)
T <sub>3</sub>	Trichoderma viride	34.00	62.39 (52.17)
T4	Trichoderma gloicladium	71.33	20.74 (27.09)
T <sub>5</sub>	Trichoderma hamatum	42.67	52.59 (46.48)
T <sub>6</sub>	Trichoderma koningi	39.00	56.66 (48.83)
<b>T</b> <sub>7</sub>	Bacillus subtilis	47.33	47.40 (43.51)
T8	Control	90.00	0.00 (0.00)
	$S.E(m) \pm$	0.80	0.89
C.D (P=0.01)		2.42	2.70

\*Mean of three replication, Dia: Diameter

# Figures in parenthesis are Aresin transformed value

The data presented in (Table No 1, Fig. 1 and plate 1) revealed that all the antagonists were found significantly superior in checking the growth of the pathogen over control. Out of seven antagonists tested, significantly least growth of the pathogen was recorded in *Pseudomonas fluorescens* (32.00 mm). However the efficiency of *Trichoderma viride* (34.00 mm) was statistically at par with that of *Pseudomonas fluorescens*. Next best in order of efficiency were *Trichoderma harzianum* (38.67 mm), *Trichoderma koningi* (39.00 mm), *Trichoderma hamatum* (42.67 mm) and *Trichoderma (Gliocladium) virens* (71.33 mm).

It is evident from these studies that among all the antagonists evaluated by dual culture technique, *Pseudomonas fluorescens* (64.61%) showed maximum growth inhibition as an antagonistic and *Trichoderma viride* (62.39%) was found statistically at par with the *Pseudomonas fluorescens* followed by *Trichoderma harzianum* (57.03%), *Trichoderma koningi* (56.66%), *Trichoderma hamatum* (52.59%) and *Trichoderma (Gliocladium) virens*, (20.74%) inhibit growth of *P. grisea*. (Table No1, Fig 2 1and plateV1).Similar work carried out by Gnanamanickam *et al.*, (1990) when he experimently found that *Pseudomonas fluorescens* strain was most effective against *P. oryzae* Cav. as it produced fluorescent antiblast antibiotics in culture which inhibited the conidial germination of *P. oryzae* Cav. *in vitro*.



Plate I: In vitro efficacy of various bio-agents against P. grisea

https://www.thepharmajournal.com

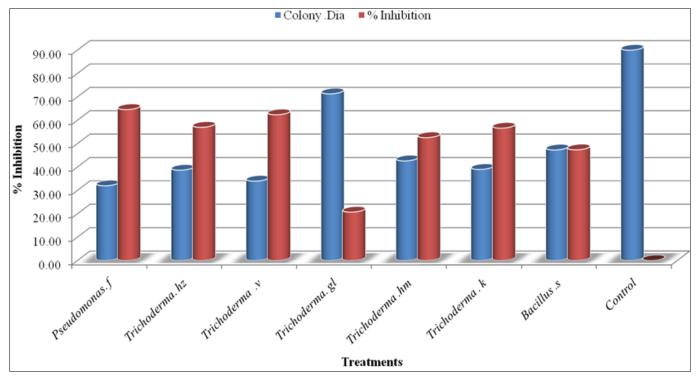


Fig 1: Effect of Bioagents on Mycelial Growth Inhibition of P.grisea.

# Conclusion

From the present study, it may be concluded that, in biological control, *P. flurosence* was found most significant.

## Acknowledgment

We are thankful to the Head of the Department of Plant Pathology, College of Agriculture, Parbhani for providing the laboratory facilities.

# Reference

- 1. Buckley TA, Allen BF. Notes on current investigations. April to June, 1951. Malaysian Agric. J. 1951;34:133-141.
- 2. Davis AJ, Dale NM, Ferreira FJ. Peael millet as an alternative feed ingredient in broiler diets. J of Applied Poultry Res. 2003;12:137 144.
- 3. Ejeta G, Hansen MM, Mertz ET. *In vitro* digestibility andamino acid composition of pearl millet (*Pennisetum typhoides*) and other cerearls. Proceedings of the National Academy of Sci; c1987.
- 4. Emechebe AM. Some aspects of crop disease in Uganda UAFRO, Seren (unpublished); c1975.
- 5. Mehta PR, Singh B, Mathur SC. A new leaf spot disease of bajra (*Pennisetum typhoides* Staph and Hubbard) caused by a species of *Pyricularia*. Indian Phytopathology. 1953;5:140-143.
- Sere Y, Onasanya A, Afolabi A, Mignouna HD, Akator K. Genetic diversity of the blast fungs *Magnaporthe* grisea (Hebert) Barr, in Burkina Faso. African J Biotechnology. 2007;6(22):25682577.
- Sharma OP. Text Book of Fungi. Publ. by Tata McGraw Hill Publishing Company Ltd., New Delhi; c2003. p. 268-269.
- 8. Wilson JP, Wells HD, Burton GW. Inheritance of resistance to *Pyricularia grisea* in pearl millet accessions from Burkino Faso and inbred Tift 85DB. Journal of Heredity. 1989;80:499-501.

9. Yadav Ravi, Pandya RK, Singh DP. To study the extend of blast severity of Pearl millet field in Morena, Bhind and Gwalior Districts of Madhya Prasesh. Society for Sci. Dev.in Agric. And tech. Progre. Res. 2012;7(2):313-314.