www.ThePharmaJournal.com

The Pharma Innovation



ISSN (E): 2277-7695 ISSN (P): 2349-8242 NAAS Rating: 5.23 TPI 2022; 11(12): 2068-2075 © 2022 TPI www.thepharmajournal.com

Received: 02-10-2022 Accepted: 05-11-2022

RM Muchhadiya

Department of Agronomy, College of Agriculture, Junagadh Agricultural University, Junagadh, Gujarat, India

PD Kumawat

Department of Agronomy, College of Agriculture, Junagadh Agricultural University, Junagadh, Gujarat, India

HL Sakarvadia

Department of Soil Sci. & Agril. Chem., College of Agriculture, Junagadh Agricultural University, Junagadh, Gujarat, India

PM Muchhadiya

Department of Biotechnology, College of Agriculture, Junagadh Agricultural University, Junagadh, Gujarat, India

Corresponding Author: RM Muchhadiya Department of Agronomy, College of Agriculture, Junagadh Agricultural University, Junagadh, Gujarat, India

Weed management with the use of nano-encapsulated herbicide formulations: A review

RM Muchhadiya, PD Kumawat, HL Sakarvadia and PM Muchhadiya

Abstract

The overall yield loss in different crops due to weed infestation is estimated at around 36.5% in India. The use of herbicides is the only viable on-farm technology today to control weeds. More than 90% of herbicides are misused, and as a result, they are lost in the environment, do not reach the target site, and do not effectively reduce weeds in crops. Nanoencapsulation of herbicides is a delivery method in which an active ingredient is covered with various materials of different sizes in the nano range and released in a controlled way for achieving season long weed free conditions. Nanoencapsulation can protect active ingredients from premature degradation (*e.g.*, photolysis, hydrolysis, biodegradation, *etc.*) and unnecessary losses by leaching and volatilization. Studies have shown that nanoencapsulation of herbicides can produce more targeted and less toxic formulations for agricultural applications. Due to the enhanced herbicidal activity in comparison with a commercial formulation, the use of nanoencapsulated herbicides is desirable as it reduces the long-term effects of residues of these herbicides in agricultural areas and their toxicity to the environment, maintaining the sustainability of agricultural production. The target specific release of nanoencapsulated herbicide is also helpful in killing the weeds without even interacting with the crop plants, which ultimately results in higher crop yield.

Keywords: Weed management, nanotechnology, nanoencapsulation, nanoherbicides

Introduction

In the current scenario, approximately 4 million tons of pesticides are used annually worldwide for food production, of which 40% are herbicides, 30% are insecticides, and 20% are fungicides (Rojas *et al.*, 2022) ^[42]. A weed is a plant that grows where it is not desired. Weeds are a menace in agriculture. On an average 320 man hours are required to remove weeds from one hectare of land (Chinnamuthu and Viji, 2018) ^[9]. Use of herbicides is the only viable on-farm technology today to control weeds. Since 2/3 of Indian agriculture is rainfed farming where usage of herbicide is very limited, weeds have the potential to jeopardize the total harvest in the delicate agro-ecosystems. Herbicides are chemical substances that are utilized to specifically, partially or totally control or kill plants. An effective herbicide should control weeds with reasonable doses selectively non-toxic to crops, remain in the area where applied, persist throughout the growing season taking care of frequently germinating weeds and leaving no residue at the end of the season permitting subsequent crops in the sequence. It is well recognized that the over dependence on herbicides has caused severe damage to our ecosystem that are manifested into their movement to non-target areas, contamination of soil and water bodies, and development of herbicide-resistant weeds.

Nanotechnology is a technology having the potential ability to study, design, create, synthesis, manipulation of functional materials, devices, and systems to fabricate structures with atomic precision by controlling the size of the matter at the scale 1-100 nanometers (Chinnamuthu and Viji, 2018)^[9]. The main aim of nanotechnology in agriculture is to maximize output (crop yields), minimize the input (fertilizers, pesticides and herbicides) and monitoring environmental factors (sensors) and applying targeted action. Nanotechnology has many applications in all stages of production, processing, storing, packaging and transport of agricultural products. Nanopesticides were named as one of ten chemical innovations that will change the world in a sustainable way by the IUPAC and nanopesticides were ranked first for their potential low impact on the environment and human health (Gomollón-Bel, 2019)^[17]. Since 2016, every year October 9 is a day to celebrate and raise awareness on how nanotechnology has enriched lives. This date, 10/9, pays homage to the nanometer scale, 10⁻⁹ meters.

The Pharma Innovation Journal

Products database available at Statistic Bank (StatNano, 2022) ^[49], there are 10,087 nanotechnology-based products commercialized, only 231 nano-products launched for agriculture application. Most of these products are nanofertilizers (43%), and nanoformulations aimed at improving plant survival against weeds, disease, pest and other stress represent 28% of these products.

Why do we need nanotechnology applications in weed management?

Only a small portion of herbicide is absorbed by the plants. The rest is lost through one or more of the following ways viz., volatilization, adsorption, leaching, photo decomposition, chemical degradation and microbial breakdown. Continuous use of herbicides results in development of resistance in weeds towards that particular herbicide (Jasieniuk et al., 1996) [22]. International survey of herbicide resistant weeds recorded 495 unique cases of herbicide resistant weeds globally, with 255 species (148 dicots and 107 monocots) in 2018. Weeds have evolved resistance to 23 of the 26 known herbicide sites of action and to 163 different herbicides. Herbicide resistant weeds have been reported in 92 crops in 70 countries. The maximum number of herbicide-resistant weed species reported in different crops are in the order of: wheat > maize > rice > soybean > spring barley > canola >cotton (Heap, 2019) [20]. Science of nanotechnology can be used as a tool to fabricate the slow release nanoencapsulated pre-emergence herbicide for achieving season long weed free condition without hampering the environment. Nanoherbicides are being developed to address the problems in perennial weed management and exhausting weed seed bank. Encapsulation by nanomaterials can protect active ingredients from premature degradation and unnecessary losses.

What is nanoencapsulated herbicide?

Nanoencapsulation of herbicides is a delivery method in which an active ingredient is covered with various materials of different sizes in nano range and released in a controlled way for achieving season long weed free condition. Substances being coated or encapsulated material are commonly known as core material, filler or internal phase, for example herbicide. Materials used for encapsulation are referred to as coating, external phase, membrane or shell, for instance nano capsules. In general, nano-encapsulated herbicides have two typical release behaviors: (i) sustained (slow) release and (ii) stimuli-responsive release. Rate of release of herbicidal suspension through diffusion is a membrane controlled system. Likewise, there are several systems developed for the controlled release of active ingredients after encapsulation of herbicide according to their properties viz., specific release, moisture release, heat-release, pH release, ultrasound release, magnetic release, etc.

Methods of nanoparticle production

The nanomaterials prepared through two basic methods (according to Royal Society and Royal Academy of Engineering, UK):

- 1. Top-down depending on size reduction from bulk materials
- 2. Bottom- up system where materials are synthesis from atomic level

Also, there are some other methods for producing nanomaterials like attrition and pyrolysis, and biological synthesis of nanoparticles.

Development of new nanoencapsulated herbicide

Encapsulation of active ingredient (a.i.) is done by Kumar and Chinnamuthu (2014)^[29]:

- 1. Indirect method of nanoencapsulation (IDM)
- 2. Direct method of nanoencapsulation (DM)
- 3. Solvent evaporation method (SEM)
- 4. Nano spray method (NSM)

Materials used in herbicide encapsulation

Wilkins (1990) ^[53] first classified the materials used in encapsulation according to their degree of biodegradation:

- 1. Starch and systems based on amylose
- 2. Other polysaccharides (cellulose and derivatives, chitin, chitosan, dextran, alginate)
- 3. Proteins (casein, albumin, gelatin)
- 4. Lipophilic materials (rubbers and waxes)
- 5. Synthetic polymers (polyvinyl alcohol, polylactato, polyglycolato, other polyesthers, polyamines, polyamide-type acids, polyacrylamide)
- 6. Miscellaneous (polyhydroxybutirato, tannins, lignins, resins, biopolymers)

Types of nano-enabled herbicides

There are three types of nano-enabled herbicides

- 1. Organic nano-enabled herbicides: Organic nanomaterials are outstanding materials for assembling nanoherbicides, and they can be based on polymers, lipids, lignocellulosic materials, proteins, complex macromolecules as dendrimers, *etc.* Overall, polymers are widely used in nano-enabled herbicide formulations due to their biodegradability and biocompatibility. Examples of some organic nano-enabled herbicides are given in Table 1.
- 2. Inorganic nano-enabled herbicides: Inorganic nanoenabled herbicides can be based on silica, metal, mesoporous silica nanoparticles, *etc.* Some of these nanoherbicides can release ions, while others can encapsulate organic molecules and release them in a controlled manner. Examples of some inorganic nanoenabled herbicides are provided in Table 2.
- 3. Hybrid nano-enabled herbicides: Hybrid materials have the potential to combine the advantages of two or more materials, such as organic and inorganic, into a single structure. These multifunctional nanomaterials can have a variety of properties, sizes, morphologies, and chemical compositions. Also, hybrid nanoherbicides can promote good targetability, traceability and stimuli-responsiveness properties. Examples of some hybrid nano-enabled herbicides are furnished in Table 3.

Table 1: Examples of some organic nano-enabled herbicides

Nanoherbicides	Plants	References
PCL(poly(ε-caprolactone)_ATZ NPs; PCL_Ametryn NPs; PCL_Simazine NPs	Allium cepa L.	Grillo et al. (2012) ^[18]
Nanoemulsion containing glyphosate	Eleusine indica L.	Jiang et al. (2012) ^[23]
CS(chitosan)/TPP(tripolyphosphate)_PQ(paraquat) NPs	Allium cepa L.; Zea mays L.; Brassica sp. L	Grillo et al. (2014) ^[19]
PCL_ATZ (Atrazine) NPs	Brassica sp.; Zea mays L.;	Pereira et al. (2014) ^[39]
Biochar_2,4-D	Brassica sp.; Zea mays L.	Abigail et al. (2016) ^[1]
Pectin NPs_ metsulfuron methyl	Chenopodium album L.	Kumar et al. (2017) ^[30]
Poly(lactic-co-glycolic acid)_ATZ NPs	Solanum tuberosum L.	Schnoor et al. (2018) ^[43]
Nanoemulsion_palm oil; <i>Parthenium hysterophorus</i> L. crude extract	Diodia ocymifolia	Zainuddin et al. (2019) ^[55]
NCs (nano/cellulose)_savory (<i>Satureja ortensis</i> L.) essential oil	Lycopersicon esculentum; Amaranthus retroflexus L.	Taban et al. (2020) ^[50]
Nano-hydrogel_ glyphosate	Weeds; Oryza sativa L.	Zhang et al. (2020) ^[56]
Carbon nanotubes_Glyphospate	Arabidopsis thaliana	Ke et al. (2021) ^[26]
CS/TPP_PQ NPs	Spinacia oleracea L.	Pontes et al. (2021) ^[41]
PCL_Metribuzin NPs	Ipomoea grandifolia	Takeshita et al. (2022) ^[51]

Table 2: Examples of some inorganic nano-enabled herbicides

Nanoherbicides	Plants	References
Mesoporous silica NPs_Trimethylammonium_2,4-D	Cucumis sativus L.; Triticum aestivum L.	Cao <i>et al.</i> (2018) ^[4]
Layered double hydroxide Imazamox	Brassica nigra L.	Khatem et al. (2019) ^[27]
Clay-imazaquin	Brassica oleracea var. botrytis L.	López-Cabeza et al. (2019) ^[33]
Mg-Al-layered double hydroxide_2,4-D	Arabidopsis thaliana L.	Nadiminti et al. (2019) ^[35]
Diquat dibromide@mesoporous silica nanoparticles-SO3	Datura stramonium L.	Shen et al. (2019) ^[45]
Zinc hydroxide nitrate-Sodium dodecylsulphate-Bispyribac	Oryza sativa L.	Sharif et al. (2020) ^[44]
Nanoformulation-sepiolite_Mesotrione	Zea mays L.; Helianthus annuus L.	Galán-Jiménez et al. (2020) ^[14]
Herbicide@Hydrotalcites	Amaranthus retroflexus	Gao et al. (2021) ^[15]
Tribenuron-methyl zein-based nanoparticles	Convolvulus arvensis L.	Heydari et al. (2021) ^[21]

Table 3: Examples of some hybrid nano-enabled herbicides

Nanoherbicides	Plants	References
AgNPs_CS(chitosan)_PQ(paraquat)	Eichhornia crassipes	Namasivayam et al. (2014) ^[36]
Wheat gluten_organically modified montmorillonite_ethofumesate	Lepidium sativum L.	Chevillard <i>et al.</i> (2014) ^[6]
GSNO (nitrosoglutathione)-containing alginate/chitosan nanoparticles	Zea mays L.; Glycine sp.	Pereira et al. (2015) ^[40]
pH-responsively controlled-release nanopesticide@ Fe ₃ O ₄ NPs_CS	Cynodon dactylon L.	Xiang et al. (2017) ^[54]
Attapulgite-NH4CO3-Gly – Amino silicol oil – poly(vinyl alcohol)	Zoysia matrella L.	Chi et al. (2017) ^[8]
Light-responsively controlled-release herbicide particle	Imperata cylindrica L.	Chen <i>et al.</i> (2018) ^[5]
Near-infrared light-responsively controlled-release herbicide particles	Cynodon dactylon L.	Liu <i>et al.</i> (2019) ^[32]
Magnetic-responsive controlled-release herbicide	Cynodon dactylon L.	Chi et al. (2021) ^[7]
MOF(Metal–organic frameworks) @DiS-O-acetil	Echinochloa crus-galli L.; Amaranthus viridis L.	Mejías <i>et al.</i> (2021) ^[34]

Positive aspects for nanotechnology application in weed management

- Innovative technology that develops precision to meet the intense demand for food.
- Minimizing crop loss, reducing costs, enhancing yield and input use efficiency.
- Nanomaterials synthesized from biopolymers (cellulose and starch) are safe and environmentally friendly.
- The higher solubility of nanoparticles in suspension.
- Higher surface area and improved targeted activity.
- Suppressing toxic residue deposition and altering the efficacy of microorganisms.
- Avoiding the resistances build up in weeds.
- Lower eco harm with safe and relaxed transport.
- Recycling of the magnetic nanocarrier materials (Xiang *et al.*, 2017)^[54].

Negative aspects for nanotechnology application in weed management

- Improper use of this technology can pose a greater threat to living organisms.
- Accumulation of nanomaterials in food products.
- Ability to pierce the healthy human skin.
- Air-borne nanoparticles pose an extreme level of threat to human and animal health.
- Unwanted and undesirable effect on non-target plants and plant-associated organisms.
- Nano-herbicides may block the vascular bundle in plants and may reduce pollination.
- Phytotoxicity, cytotoxicity, and genotoxicity.
- It may cause cell death or DNA mutation.
- Negative impact of nanomaterials on certain soil microbial communities and algae.

Challenges for nanotechnology application in weed management

- The major challenge in adopting this technology on a large scale is overcoming the risk issue. And as we know that 'technology-yes but safety-must', before the adoption of this technology, it is very important to assess the possible risks and consequences of using nanoparticles.
- The agricultural sector is still comparably marginal and has not yet been able to make progress in the market to any greater extent relative to other fields of nanotech application. Nanotechnology remains insignificant and has not yet taken the market because of its poor cargo loading capacity.
- Nanoparticles behave in an unpredictable manner, which may be harmful to life.
- Limited research on nanotechnology-based on its risky elements and toxicity.
- Nanomaterials are still being explored for applications in agrochemicals.
- Production cost, evaluation standards, registration policies and public concern issues must be addressed.

Effect of nanoencapsulated herbicide formulations Application rate of herbicide

Nanoencapsulation potentiated the activity of the herbicide in relation to commercial herbicide formulation, allowing maintenance of its activity even when applied at a reduced dosage. So, the nanoencapsulated herbicide presented a low environmental risk, with increased weed control. It may be due to improved site of action by nanoencapsulation, which maintained the product's efficacy even with a reduction in the applied dose. Namasivayam et al. (2014)^[36] observed that all the tested concentration of silver nanoparticles-chitosan encapsulated paraquat induced necrotic lesions on Eichhornia crassipes with the diameter of 1.0, 1.2 and 1.5 cm at 05, 10 and 25 µg/ml. Free paraquat recorded 0.8 and 1.0 cm of necrotic lesions at 10 and 25 µg/ml concentration. Sousa et al. (2018) ^[48] found that the use of 10-fold diluted atrazineloaded PCL nanocapsules (200 g a.i./ha) resulted in the same inhibitory effect in root and shoot growth of weeds as the commercial formulation at the standard atrazine dose (2,000 g a.i./ha). Sousa et al. (2020) [47] revealed that at standard dosage (2,000 g a.i./ha) and at half-dosage (1,000 g a.i./ha), nanoatrazine was equally or more efficient in affecting most of the evaluated parameters than conventional atrazine at full dosage. It might be associated with the modified release of atrazine by the nanocapsules, with a better adhesion of the nanoformulation to the leaves or with the uptake of the nanocapsules by the leaf stomates, thereby preventing atrazine loss into the environment and improving the delivery of the herbicide to the target organism. Longer term presence of the nanoencapsulated herbicide in plant metabolism, surpassing the tolerance mechanisms of the weed. Similar findings reported by Sousa et al. (2022)^[46] and Takeshita et al. (2022) [51]

Effect of herbicide on weed growth

Slow release and improved delivery of nanoencapsulated herbicides to the target organism leads to lower weed growth. Sousa *et al.* (2018)^[48] found that the growth of *Amaranthus viridis* plants was equally reduced by nanoatrazine and commercial formulation. In the case of *Bidens pilosa*,

atrazine-loaded nanocapsules decreased more effectively the root and shoot growth than the commercial formulation, leading to a loss of plant biomass. Vimalrajiv et al. (2019)^[52] recorded lower weed density and weed dry weight with application of pendimethalin + hand weeding at 35 DAS, which was at par with the application of H₂O₂ fb pendimethalin + Ag Nps. This might be due to the strong oxidizing capacity of H₂O₂ that results in oxidative stress that causes cellular damage and death of seeds. The applied nanoparticles might have entered into the weed system and degraded the phenol and starch present in the tubers. Further combined application of silver nanoparticles with pendimethalin might have reduced the emergence in addition to killing of emerged weeds resulting in lower weed population and dry weight of weeds. Bommayasamy and Chinnamuthu (2021)^[3] recorded the lowest weed density and weed dry matter production with the application of oxadiargyl loaded with zeolite at 20 DAT. It might be due to zeolite and biochar entrapped herbicides have increased sorption and decreased the dissipation of herbicide in soil which helps to release herbicide slowly through the entire season which destroys food reserves of weed seeds and causes lesser regeneration of weeds. Takeshita et al. (2022) [51] observed that plants treated with nanoMTZ and MTZ for the highest dose (480 g a.i./ha), and nanoMTZ for the lowest dose (48 g a.i./ha) induced similar reductions in shoot and root dry masses of Ipomoea grandifolia compared to the control samples. These results indicated that the nanoencapsulation potentiated the activity of the herbicide in relation to commercial formulation.

Effect of herbicide on plant physiological parameters

Both commercial atrazine and PCL nanocapsules containing atrazine did not lead to any side effects on maximum quantum vield of PSII, net photosynthesis or leaf lipid peroxidation of the maize plants, as compared to the control (Oliveira et al., 2015b) [38]. Negative effects of atrazine were transient, probably due to the ability of maize plants to detoxify the herbicide. Sousa et al. (2020) [47] revealed that at standard dosage (2,000 g a.i./ha), nanoatrazine induced higher and faster reductions of maximum PSII activity of Digitaria insularis at two expanded leaves than conventional atrazine. Sousa et al. (2022) [46] recorded that plants treated with nanoatrazine (at 200 g a.i./ha) induced significantly higher inhibition of the maximum quantum efficiency of photosystem II than conventional atrazine at the same concentration, 24 and 96 hours after application. The enhanced inhibition of PSII activity by nanoatrazine might be due to greater uptake of the nanoherbicide by stomata and hydathodes of leaves. Takeshita et al. (2022)^[51] reported the maximum inhibition of PSII activity and decrease in pigment levels of Ipomoea grandifolia under nanoencapsulated metribuzin and commercial metribuzin with the highest dose (480 g a.i./ha) was observed. It might be due to improved site of action by nanoencapsulation. Although the findings provided clear evidence of the ability of PCL nanocapsules to increase the activity of atrazine towards a target species, the mechanisms involved remain unclear.

Effect of herbicide on crop yield

Crop growing in an environment with minimum disturbance due to weeds reflected on crop yield by enhancing the growth and seed yield. This might be due to reduced competition

The Pharma Innovation Journal

between crop and weed for different resources. Kumar and Chinnamuthu (2017) ^[28] indicated that 6 percent starch encapsulated pendimethalin by solvent evaporation method resulted in higher growth and yield of blackgram. Vimalrajiv *et al.* (2019) ^[52] concluded that among the herbicide treatments, the pre-emergence application of pendimethalin + HW on 35 DAS recorded the highest seed yield and haulm yield. The next best treatment was application of $H_2O_2 fb$ pendimethalin + Ag Nps. Bommayasamy and Chinnamuthu (2021) ^[3] recorded higher grain yield of rice with the application of butachlor at 1.25 kg/ha fb hand weeding on 40 DAT followed by oxadiargyl loaded with zeolite. This might be due to better control of weeds over a longer period of time, thus providing a favourable environment for better growth and development of crop, leading to enhanced crop yield.

Effect of herbicide on nutrient uptake

Slow release of nanoencapsulated herbicides significantly reduced weed dry matter production which led to synthesizing more food materials through effective photosynthesis and higher nutrient uptake by crop. Bommayasamy and Chinnamuthu (2021)^[3] recorded the higher N, P and K uptake by rice with the application of butachlor at 1.25 kg/ha fb hand weeding on 40 DAT, which was comparable with oxadiargyl loaded with zeolite and oxadiargyl loaded with biochar. It might be due to zeolite and biochar entrapped herbicides have increased sorption and decreased the dissipation of herbicide in soil which helps to release herbicide slowly through the entire season which destroys food reserves of weed seeds and causes lesser regeneration of weeds. Further application of zeolite which acted as a soil conditioner; increased fertilizer use efficiency by controlled release of nutrient in match with the requirement of plants during different physiological growth stages.

Effect of herbicide on weed control

Slow release of entrapped herbicide reduced the herbicide movement within the soil column and kept the sizable portion of the active ingredient in the upper soil layer over a long period. This may lead to improved weed control efficiency and reduce the frequencies of herbicide application. Oliveira et al. (2015a) [37] reported that atrazine-containing PCL nanocapsules provided very effective post-emergence herbicidal activity and the use of nanoencapsulated atrazine enabled the application of lower dosages of the herbicide, without any loss of efficiency. Kumar and Chinnamuthu (2017) ^[28] indicated that 6 percent starch encapsulated pendimethalin by solvent evaporation method, released the nanoencapsulated pendimethalin slowly with sufficient quantity to control the weed seed germination. Divanat and Saeidian (2019)^[12] recorded the lowest percentage of purslane emergence under the PCL nanoencapsulated metribuzin treatment, which was significantly lower than the commercial metribuzin treatment. Vimalrajiv et al. (2019) [52] recorded higher weed control efficiency using H_2O_2 fb pendimethalin + Ag Nps. They also noted lower weed index with the application of pendimethalin + hand weeding at 35 DAS, which was followed by treatment H_2O_2 fb pendimethalin + Ag Nps. This improved weed control might be due to the strong oxidizing capacity of H₂O₂ that results in oxidative stress that causes cellular damage and death of seeds. Further combined application of silver nanoparticles with pendimethalin might have reduced the emergence in addition to killing of emerged

weeds resulting in lower weed population and dry weight of weeds. Kurniadie *et al.* (2022)^[31] found that a mixture of paraquat (280 g a.i./ha) and chitosan significantly increased the herbicidal efficacy against *Ageratum conyzoides*, *Paspalum conjugatum* and *Borreria alata* under the rainfall conditions. The chemical structure of chitosan might have contributed to the penetration of paraquat into plant tissues.

Effect of nanoencapsulation on stability of herbicide

Encapsulation controls the active ingredients' release rate and protects susceptible materials from undesirable environmental conditions. Daneshvari *et al.* (2021) ^[11] observed that unlike the EC formulation, the amount of active ingredient in the encapsulated formulation was not significantly decreased following 8 and 16 h of exposure to natural sunlight. They also found that with increasing UV exposure, the active ingredient in the EC formulation decreased linearly and reached 43% after 8 h. In comparison, only 0.9% of the initial herbicide level in the encapsulation was lost during the same time. Results indicated that an effective herbicide such as trifluralin can be protected from volatilization and photodegradation by developing an encapsulated formulation.

Effect of nanoencapsulation on availability of herbicide

Nanoencapsulation of herbicide decreases its absorption by the soil colloids and increases its availability for the target plant. Diyanat *et al.* $(2019)^{[13]}$ observed that polycaprolactone nanocapsules containing pretilachlor were distributed in the layers of 0–5 and 5–10 cm and not found at the depth of 10–15 cm. Most of the encapsulated pretilachlor remained in the top layers (0–10 cm), which is desirable in weed control.

Effect of herbicide on soil microbial population

Microorganisms are influenced by several factors including the application of herbicides. Among the different soil microbes, more sensitive microbes to herbicides are bacteria (Ghinea et al., 1998)^[16]. Sulfentrazone applied to sugarcane crop at lower doses of 720 and 840 g a.i./ha did not affect the microflora but in case of higher doses of 1320 and 2400 g a.i./ha initial reduction of microflora was observed and recovered 30 days after application (Kalaiyarasi, 2012)^[24]. This might be due to carbon released from degraded herbicide which leads to an increase of the soil microflora population (Bera and Ghosh, 2013)^[2]. Kannamreddy et al. (2020)^[25] reported that all the tested commercial and nanoencapsulated herbicides showed reduction in bacterial, fungal and actinomycetes population at 25 DAS compared to initial population. But at 50 DAS there was a great increase in microbial population compared to 25 DAS in all herbicide applied treatments. There was no significant difference among all the treatments at 50 DAS in the microbial population. They also found that pendimethalin @ 1 kg a.i./ha and encapsulated sulfentrazone @ 0.3 kg a.i./ha recorded higher nodule count and nodule dry weight of blackgram at 30 DAS. But at 60 DAS there was no significant difference among the treatments except with unweeded control. It might be due to herbicidal degradation by microbes, there was gradual increase in soil microbiome and nodulation ability at 60 DAS.

Phytotoxic effect of herbicide

Modified release and improved target delivery of the nanoencapsulated herbicides reduce side effects/ phytotoxic effects on non-target plants and plant-associated organisms.

Namasivayam *et al.* (2014) ^[36] observed that silver nanoparticles-chitosan encapsulated paraquat induced necrotic lesions on *Eichhornia crassipes*. Whereas, necrotic lesions were not recorded on paddy and wheat leaves. So, there was no phytotoxicity of nanoencapsulated paraquat against paddy and wheat leaves. Oliveira *et al.* (2015b) ^[38] suggested that the use of PCL nanocapsules containing atrazine did not lead to persistent side effects in maize plants, and that the technique could offer a safe tool for weed control without affecting crop growth. Diyanat and Saeidian (2019) ^[12] also indicated that PCL nanoencapsulated metribuzin had no side effect on soybean.

Toxicity of herbicide

Studies have shown that nanoencapsulation of herbicides can produce more targeted and less toxic formulations for agricultural applications. Clemente et al. (2013)^[10] reported that poly(*\varepsilon*-caprolactone) (PCL) nanocapsules containing ametryn and atrazine resulted in lower toxicity to the human and alga (Pseudokirchneriella subcapitata) and higher toxicity to the microcrustacean (Daphnia similis) as compared to the herbicides alone. These findings indicated that encapsulation of the herbicides acted to reduce the damage caused to the cells, which can be explained by the slower release and consequently smaller quantities of the herbicides available to promote cellular toxicity. Grillo et al. (2014)^[19] noted that chitosan and tripolyphosphate encapsulated nanoherbicides are environmentally safer alternatives for weed control. Takeshita et al. (2022) [51] also verified that nanoencapsulated metribuzin in polymeric nanoparticles presented a low environmental risk, with increased weed control.

Conclusion

The use of herbicides is fundamental to maintaining crop yields and expanding the extent of the production area. Herbicides have eased the problem of weed control, but there are still problems, such as herbicide persistence in soil, that have been lowering the quality of soil. Apart from that, the trends among the weeds of developing resistance to the herbicides have been a serious issue. Nanotechnology, with its unique way of releasing herbicides, can give promising results. Studies have shown that nanoencapsulation of herbicides can produce more targeted and less toxic formulations for agricultural applications. Due to the enhanced herbicidal activity in comparison with a commercial formulation, the use of nanoencapsulated herbicide would permit the application of lower dosages of the herbicide. The use of lower doses of herbicides is desirable as it reduces the long-term effects of residues of these herbicides in agricultural areas and their toxicity to the environment. Encapsulated herbicides can aid in the easy delivery of herbicides to weed plants, reducing residual accumulation in soil. The target specific release is also helpful in killing the weeds without even interacting with the crop plants and ultimately results in a higher crop yield. Nanotechnology is thus a boon that can be further developed with regard to the target site inhibition of the biochemical reactions of weed.

References

1. Abigail MEA, Samuel SM, Ramalingam C. Application of rice husk nanosorbents containing 2,4dichlorophenoxyacetic acid herbicide to control weeds and reduce leaching from soil. Elsevier BV. 2016;63:318-326.

- 2. Bera S, Ghosh RK. Soil microflora and weed management as influenced by atrazine 50% WP in sugarcane. Univers. J Agric. Res. 2013;1(2):41-47.
- Bommayasamy N, Chinnamuthu CR. Effect of encapsulated herbicides on weed control, productivity and nutrient uptake of rice (*Oryza sativa*). J. Environ. Biol. 2021;42:319-325.
- Cao L, Zhou Z, Niu S, Cao C, Li X, Shan Y *et al.* Positive-charge functionalized mesoporous silica nanoparticles as nanocarriers for controlled 2,4dichlorophenoxy acetic acid sodium salt release. J. Agric. Food Chem. 2018;66:6594-6603.
- 5. Chen C, Zhang G, Dai Z, Xiang Y, Liu B, Bian P *et al.* Fabrication of light-responsively controlled-release herbicide using a nanocomposite. Chem. Eng. J. 2018;349:101-110.
- Chevillard A, Angellier-Coussy H, Guillard V, Bertrand C, Gontard N, Gastaldi E. Biodegradable herbicide delivery systems with slow diffusion in soil and UV protection properties. Pest Manag. Sci. 2014;70:1697-1705.
- 7. Chi Y, Chen C, Zhang G, Ye Z, Su X, Ren X *et al.* Fabrication of magneticresponsive controlled-release herbicide by a palygorskite-based nanocomposite. Colloids Surf. B: Biointerfaces. 2021;208:112-115.
- Chi Y, Zhang G, Xiang Y, Cai D, Wu Z. Fabrication of a temperature-controlled-release herbicide using a nanocomposite. ACS Sustain. Chem. Eng. 2017;5:4969-4975.
- Chinnamuthu CR, Viji N. Trends and developments of nanotechnology application in weed management in India. In: Fifty years of weed science research in India. Indian Society of Weeds Science, Jabalpur; c2018. 51-69.
- Clemente Z, Grillo R, Jonsson M, Santos NZP, Feitosa LO, Lima R *et al.* Ecotoxicological evaluation of poly(epsilon-caprolactone) nanocapsules containing triazine herbicides. J. Nanosci. Nanotechnol. 2013;13:1-7.
- 11. Daneshvari G, Yousefi AR, Mohammadi M, Banibairami S, Shariati P, Rahdar A *et al.* Controlled-release formulations of trifluralin herbicide by interfacial polymerization as a tool for environmental hazards. Biointerface Res. Appl. Chem. 2021;11(6):13866-77.
- 12. Diyanat M, Saeidian H. The metribuzin herbicide in polycaprolactone nanocapsules shows less plant chromosome aberration than non-encapsulated metribuzin. Environ. Chem. Lett. 2019. https://doi.org/10.1007/s10311-019-00912-x.
- 13. Diyanat M, Saeidian H, Baziar S, Mirjafary Z. Preparation and characterization of polycaprolactone nanocapsules containing pretilachlor as a herbicide nanocarrier. Environ. Sci. Pollut. Res. 2019;26:21579-88.
- Galán-Jiménez M, Maria del Carmen E, Morillo BF, Mallet C, Undabeytia T. A sepiolite-based formulation for slow release of the herbicide mesotrione. Appl. Clay Sci. 2020;189:105503.
- 15. Gao Y, Zhou Z, Chen X, Tian Y, Li Y, Wang H *et al.* Controlled release of herbicides by 2,4-D-, MCPA-, and bromoxynil-intercalated hydrotalcite nanosheets. Green Chem. 2021, 23.
- 16. Ghinea L, Lancu M, Turcu M, Stefanic G. The impact of

sulfonyl-urea and non-selective herbicides on biological activity of sandy soils. Rom. Agric. Res. 1998;9(1):55-57.

- 17. Gomollón-Bel F. Ten chemical innovations that will change our world: IUPAC identifies emerging technologies in chemistry with potential to make our planet more sustainable. Chem. Int. 2019;41:12-17.
- Grillo R, dos Santos NZP, Maruyama CR, Rosa AH, de Lima R, Fraceto LF. Poly(ε-caprolactone) nanocapsules as carrier systems for herbicides: Physicochemical characterization and genotoxicity evaluation. J. Hazard. Mater. 2012;231-232:1-9.
- 19. Grillo R, Pereira AES, Nishisaka CS, de Lima R, Oehlke K, Greiner R *et al.* Chitosan/tripolyphosphate nanoparticles loaded with paraquat herbicide: An environmentally safer alternative for weed control. J. Hazard. Mater. 2014;278:163-171.
- 20. Heap I. International survey of herbicide resistant weeds. www.weedscience.org. 21 October, 2022.
- Heydari M, Yousefi AR, Rahdar A, Nikfarjam N, Jamshidi K, Bilal M *et al.* Microemulsions of tribenuronmethyl using pluronic F127: Physico-chemical characterization and efficiency on wheat weed. J. Mol. Liq. 2021;326:115263.
- 22. Jasieniuk M, Brûl'e-Babel AL, Morrison IN. The evolution and genetics of herbicide resistance in weeds. Weed Sci. 1996;44(1):176-193.
- 23. Jiang LC, Basri M, Omar D, Rahman MBA, Salleh AB, Rahman RNZ *et al.* Green nano-emulsion intervention for water-soluble glyphosate isopropylamine (IPA) formulations in controlling *Eleusine indica*. Pestic. Biochem. Physiol. 2012;102:19-29.
- 24. Kalaiyarasi D. Evaluation of sulfentrazone for weed control in sugarcane and its residual effect on succeeding crops. Ph.D. Thesis, Tamil Nadu Agricultural University, Coimbatore, 2012.
- 25. Kannamreddy V, Chinnamuthu CR, Marimuthu S, Bharathi C. Effect of nanoencapsulated pre-emergence sulfentrazone herbicide on soil microbiome and nodulation of irrigated Blackgram (*Vigna mungo L.*). Int. J. Curr. Microbiol. App. Sci. 2020;9(7):1348-54.
- 26. Ke M, Ye Y, Zhang Z, Gillings M, Qu Q, Xu N *et al.* Synergistic effects of glyphosate and multiwall carbon nanotubes on *Arabidopsis thaliana* physiology and metabolism. Sci. Total Environ. 2021;769:145156.
- 27. Khatem R, Celis R, Hermosín MC. Cationic and anionic clay nanoformulations of imazamox for minimizing environmental risk. Appl. Clay Sci. 2019;168:106-115.
- Kumar P, Chinnamuthu CR. Assembly of nanoencapsulated pendimethalin herbicide using solvent evaporation method for season long weed control under irrigated ecosystem. Int. J. Pure App. Biosci. 2017;5(1):349-357.
- 29. Kumar P, Chinnamuthu CR. Synthesis and fabrication of nano-encapsulated herbicide using direct and indirect core/core-shell and solvent evaporation method for slow release in irrigated blackgram. Ph.D. research. Tamil Nadu Agricultural University, Coimbatore, 2014.
- Kumar S, Bhanjana G, Sharma A, Dilbaghi N, Sidhu MC, Kim KH. Development of nanoformulation approaches for the control of weeds. Sci. Total Environ. 2017;586:1272-78.
- 31. Kurniadie D, Umiyati U, Widianto R, Kato-Noguchi H.

Effect of chitosan molecules on paraquat herbicidal efficacy under simulated rainfall conditions. Agronomy 2022;12:1666.

- 32. Liu B, Chen C, Wang R, Dong S, Li J, Zhang G *et al.* Nearinfrared light-responsively controlled-release herbicide using biochar as a photothermal agent. ACS Sustain. Chem. Eng. 2019;7:14924-32.
- López-Cabeza R, Poiger T, Cornejo J, Celis RA. A claybased formulation of the herbicide imazaquin containing exclusively the biologically active enantiomer. Pest Manag. Sci. 2019;75(7):1894-1901.
- 34. Mejías FJR, Trasobares S, Varela RM, Molinillo JMG, Calvino JJ, Macías FA. One-step encapsulation of orthodisulfides in functionalized zinc MOF. Enabling metalorganic frameworks in agriculture. ACS Appl. Mater. Interfaces 2021;13:7997-8005.
- 35. Nadiminti PP, Sharma H, Kada SR, Pfeffer FM, O'Dell LA, Cahill DM. Use of Mg-Al nanoclay as an efficient vehicle for the delivery of the herbicide 2,4-dichlorophenoxyacetic acid. ACS Sustain. Chem. Eng. 2019;7:10962-70.
- Namasivayam SKR, Aruna A, Gokila. Evaluation of silver nanoparticles-chitosan encapsulated synthetic herbicide paraquat (AgNp-CS-PQ) preparation for the controlled release and improved herbicidal activity against *Eichhornia crassipes*. Res. J. Biotechnol. 2014;9(9):19-27.
- 37. Oliveira HC, Stolf-Moreira R, Martinez CBR, Grillo R, de Jesus MB, Fraceto LF. Nanoencapsulation enhances the post-emergence herbicidal activity of atrazine against mustard plants. PLoS ONE 2015a;10(7):e0132971.
- 38. Oliveira HC, Stolf-Moreira R, Martinez CBR, Sousa GFM, Grillo R, de Jesus MB *et al.* Evaluation of the side effects of poly(epsilon-caprolactone) nanocapsules containing atrazine toward maize plants. Front. Chem. 2015b;3:61.
- Pereira AES, Grillo R, Mello NFS, Rosa AH, Fraceto LF. Application of poly(epsilon-caprolactone) nanoparticles containing atrazine herbicide as an alternative technique to control weeds and reduce damage to the environment. J. Hazard. Mater. 2014;268:207-215.
- 40. Pereira AES, Narciso AM, Seabra AB, Fraceto LF. Evaluation of the effects of nitric oxide-releasing nanoparticles on plants. J. Phys.: Conf. Ser. 2015;617:012025.
- 41. Pontes MS, Antunes DR, Oliveira IP, Forini MML, Santos JS, Arruda GJ *et al.* Chitosan/tripolyphosphate nanoformulation carrying paraquat: Insights on its enhanced herbicidal activity. Environ. Sci.: Nano 2021;8:1336-51.
- 42. Rojas S, Rodríguez-Diéguez A, Horcajada P. Metal– organic frameworks in agriculture. ACS Appl. Mater. Interfaces 2022;14:16983-17007.
- 43. Schnoor B, Elhendawy A, Joseph S, Putman M, Chacón-Cerdas R, Flores-Mora D *et al.* Engineering atrazine loaded poly (lactic-co-glycolic acid) nanoparticles to ameliorate environmental challenges. J. Agric. Food Chem. 2018;66:7889-98.
- 44. Sharif SNM, Hashim N, Isa IM, Bakar SA, Saidin MI, Ahmad MS *et al.* Controlled release formulation of zinc hydroxide nitrate intercalated with sodium dodecylsulphate and bispyribac anions: a novel herbicide nanocomposite for paddy cultivation. Arab. J. Chem.

The Pharma Innovation Journal

2020;13:4513-27.

- 45. Shen Z, Zhou X, Sun X, Xu H, Chen H, Zhou H. Preparation of 2,4-dichlorophenoxyacetic acid loaded on cysteamine-modified polydopamine and its release behaviors. J. Appl. Polym. Sci. 2019;136:47469.
- 46. Sousa BT, Pereira AES, Fraceto LF, Oliveira HC, Dalazen G. Post-emergence herbicidal activity of nanoatrazine against *Alternanthera tenella* plants compared to other weed species. Heliyon 2022;8:e09902.
- Sousa BT, Pereira AES, Fraceto LF, Oliveira HC, Dalazen G. Effectiveness of nanoatrazine in postemergent control of the tolerant weed *Digitaria insularis*. J. Plant Prot. Res. 2020;60(2):185-192.
- 48. Sousa GFM, Gomes DG, Campos EVR, Oliveira JL, Fraceto LF, Moreira RS *et al.* Post-emergence herbicidal activity of nanoatrazine against susceptible weeds. Front. Environ. Sci. 2018;6:12.
- 49. StatNano. Nanotechnology Products Database. https://product.statnano.com/industry/ agriculture. 10 September, 2022.
- 50. Taban A, Saharkhiz MJ, Khorram M. Formulation and assessment of nanoencapsulated bioherbicides based on biopolymers and essential oil. Ind. Crop. Prod. 2020;149:112348.
- 51. Takeshita V, Carvalho LB, Galhardi JA, Munhoz-Garcia GV, Pimpinato RF, Oliveira HC *et al.* Development of a preemergent nanoherbicide: From efficiency evaluation to the assessment of environmental fate and risks to soil microorganisms. ACS Nano 2022;2:307-323.
- 52. Vimalrajiv B, Chinnamuthu CR, Subramanian E, Senthil K. Management of weed seed bank using nanoparticles in combination with pendimethalin and hydrogen peroxide in irrigated blackgram (*Vigna mungo* L.). Madras Agric. J. 2019;106(1-3):36-31.
- 53. Wilkins RM (ed.). Controlled delivery of crop-protection agents. Taylor and Francis Ltd., London, 1990, 325.
- 54. Xiang Y, Zhang G, Chi Y, Cai D, Wu Z. Fabrication of a controllable nanopesticide system with magnetic collectability. Chem. Eng. J. 2017;328:320-330.
- 55. Zainuddin NJ, Ashari SE, Salim N, Asib N, Omar D, Lian GEC. Optimization and characterization of palm oilbased nanoemulsion loaded with parthenium hysterophorus crude extract for natural herbicide formulation. J. Oleo Sci. 2019;68(8):1-11.
- Zhang L, Chen C, Zhang G, Liu B, Wu Z, Cai D. Electrical-driven release and migration of herbicide using a gel-based nanocomposite. J Agric. Food Chem. 2020;68:1536-45.