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Generation mean analysis for submergence tolerant and yield traits in rice

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Abstract

Prior information on genetics and gene action of desirable traits in parents is prerequisite for development of strategies of breeding programs of the crop plants. The components of genetic variation are crucial factors determines destiny of breeding. Assessing genetic variation among genotypes to be utilized as parent in crossing program is paramount importance in harnessing accelerative genetic gain with precision. Results of the study shown the that numbers of gene involvement in breeding value, additive (d) effect and dominant \times dominant (l) gene interaction were the only significant portion of gene controlling plant height, number of effective tillers, grain yield per plant, spikelet fertility and percent survibility under submergence stress. The additive and dominance gene effects were found important in managing submergence stress in rice. The plus sign in the additive gene effect implies that rice variety Hasanta contributes positively to the trait as compared to Swarna Sub1 and *vice versa*.

Keywords: Generation mean analysis, scaling test, epistatic, gene action

Introduction

Rice (*Oryza sativa* L.) is prime source of livelihood in Asian countries, grown extensively across the regions and ecosystems in the world. In Asia, it covers ~50% of the area and ~90.4% of the whole rice production for ~ 2.7 billion peoples (Rout *et al.*, 2020) ^[28]. In addition, this sole crop provides employment for billions of people directly/indirectly (Dat, 2004). India has largest area (44.6 mha) under rice crop and ranked 2^{nd} in production after China (Verma *et al.*, 2016) ^[4]. Rice has played great role in apprehension of 'Green Revolution' in the country (Kumar *et al.*, 2019) ^[17]. However, changing climate and demographic scenario (~9.0 billion people by 2030) creating challenge towards meeting food demand across the world which necessitates further breeding attention (Gurdev, 2006) ^[7]. In addition to that, the water scarcity, decreasing arable land, resurgence of minor insect pests and diseases coupled with climate changes going to be great challenge for sustainable rice production.

Frequent occurrence of abiotic stresses like submergence, drought and salinity are the most detrimental for rice production in rainfed ecosystem (Sekar and Pal 2012) ^[32]. Submergence due to flash flooding (FF) is third important abiotic stress in rainfed ecologies, hampers sustainability of crop plants and causes huge yield loss, annually (Iftekharuddaula *et al.*, 2015) ^[11]. In India, ~36% of the rice area are submergence prone, of that ~90% are distributed in eastern states where rice is major commodity of livelihood (Pradhan *et al.*, 2015) ^[23]. Being semi-aquatic in nature, rice plant is able to modulates in various ways to sustain under submergence (Khush *et al.*, 1997) ^[14]. However, intense and prolonged submergence is fatal for its substantiality (Kumar *et al.*, 2021) ^[16]. Submerged plant canopy faces several challenges like energy shortage, nutrient and oxygen deficiency which affects proper plant growth and survival (Pradhan *et al.*, 2019) ^[24]. Besides, submergence creates very conducive environment for the growth of several biotic stresses like bacterial blight (BB) and sheath blight which makes havoc to the rice sustainability under rainfed shallow-lowland. Changing climate scenario looking very distressing, predicting severe and frequent flooding down the line in coastal regions (Sanchez *et al.*, 2000) ^[30].

Development of climate resilient and multiple disease resistance cultivars found to be substantial to combat rice yield losses (Ronald *et al.*, 1992, Das *et al.*, 2016) ^[27]. Rice plants has inbuilt ability to sustain flooding up to one week (Adkins *et al.*, 1990, Palada *et al.*, 1972) ^[1, 22], however, beyond that only tolerant genotypes can survive (Catling *et al.*, 1992) ^[3].

Tolerant plants make several adaptive changes to sustain under hypoxia/anoxia without major damage (Pradhan et al., 2019^[23], Samanta et al., 2020^[29], Sarkar et al., 2014^[31]). Tolerant plants are able to manipulates synthesis/secretion of several plant growth regulators like ethylene, gibberellic acid (GA), abscisic acid (ABA) which regulates shoot elongation and leaf senescence, thus, enhances plant survival under flood (Rajpurohit et al., 2011). Tolerant plants are able to switchedon alcohol dehydrogenase (ADH) activity which controls judicious energy consumption and enhances plant survivality under submergence (Kumar et al., 2021)^[16].

In rice, genomic blocks regulating adaptive modulations under flooding have been identified, validated and successfully deployed in several suitable genetic background (Singh et al., 2016, Rout et al., 2020) [33, 28]. The Sub1 gene controlling two weeks' tolerance is mapped on chromosome-9 alongwith other minor QTLs in the Indian rice landrace FR13A (Xu et al., 1996 and 2006) [37, 36]. In this context, genomics based improvement of plant resistance/tolerance has been proven as most promising strategies towards development of sustainable cultivars. Identification of suitable donors with nature of genetics and gene action with close genetic proximity and duration with recurrent parent is paramount important for precise and fast development of climate resilient cultivars. This study is aimed at assessment of genetics and gene action of traits contributing tolerant as well as yield under submergence for their judicious utilization in breeding programs. Generation mean analysis is used to determine the gene action governing quantitative traits, extent of additive, dominance and epistatic effects for designing appropriate breeding strategies.

Materials and Methods

The experiment was laid down at research farm of ICAR-National Rice Research Institute, Cuttack during both seasons from Kharif 2019 to Khrif-2021. The well fertile land with good irrigation and drainage facilities was selected to conduct the trials. The experimental plot was alluvial loam which is supplemented with per hectare 120 kg N, 60 kg P2O5 and 40 kg K₂O during both seasons. The F1s were generated between Hasanta and Swarna Sub1 (Submergence tolerant) where Hasanta was used as female. During Kharif, 2021 all crosses including F₁, BC₁, BC₂ and F₂ along with both parents, P₁ and P₂ were grown in three replications in RBD fashion. Twenty

days old seedlings of each generations and parents were grown in screening tank in three row with 22 plants per row at 20cm x 15 cm spacing. Subsequently, complete submergence was imposed for 14 days with 1.5-meter water depth. After fourteen days of submergence, water was removed from the tanks and regeneration was recorded after one week (IRRI, 2014)^[12]. The data were recorded on five plants from each of the entries for the characters namely: days to 50% flowering (DFF), days to maturity (DM), plant height (PH), flag leaf length (FLL), flag leaf width (FLW), number of effective tillers per plant (NETPP), panicle length (PL), panicle weight (PW), number of grains per panicle (NGPP), number of chaffs per panicle (NCPP), Spikelets fertility % (SF%), test weight (TW), grain yield per plant (GYPP) and % survival (% S). Data on agro-morphological and % survibility traits were recorded after each one week of submergence (0, 7, 14 and 21 days) on randomly selected ten plants t in each generation.

Statistical Analysis

Analysis of variance was performed by using procedure of randomized block design analysis given by Panse and Sukhatme (1967), for all six generations. Scaling test was followed for estimation of genetic component of variation for studied traits (Mather, 1949 and Hayman and Mather, 1955) ^[19, 8]. Six parameter model (Hayman, 1958 and Jinks and Jones, 1958) ^[10, 13] was analyzed for all possible genetic components of variation. The 'weighted least square method' was utilized for estimation of the genetic parameters, m, d and h (Nelder, 1960 and Hayman, 1960)^[21, 9]. Above estimates were utilized for calculation of expected generation means which compared the observed generation mean values by using χ^2 test. The significant χ^2 value indicates the nonadequacy of model and presence of non-allelic interactions.

Result and Discussion Analysis of variance

The analysis of variance revealed that all six generations (F_1, F_2) BC₁, BC₂, F₂, P₁, P₂) under submergence stress differed significantly for all the traits (Table 1). It evidenced from analysis of variance that lines and treatments showed significant variation for all traits under all submergence regimes. Similarly, significant difference among lines for yield traits under submergence was reported by Rao et al 2017 [26].

Source of variation	DF	DFF	DM	PH(c m)	FLL (cm)	FLW (cm)	NETP P	PL (cm)	PW (g)	NGP P	NCPP	SF%	TW	GYPP (g)	KL (mm)	KW(m m)	L/B Ratio	HRR %	SV%
Replication	2	0.66	26.85*	10.09* *	0.18	2.25*	362.05 *	131.34 **	0.10	3.21* *	414.4*	18.88*	363.74 **	49.91* *	0.89	15.63*	0.05*	*	22.50 *
Genotype (a)	10	**	357.81 **	**	*	0.027*	6.49**	20.35* *	11.61* *	607.4 **	341.75 **	24.90* *	10.34* *	131.84 **	0.549*	0.070*	0.671* *	29.20 **	490.7 3
Treatment (b)	3	0.94*	7.44**	17.98* *	2.15*	4.47**	2207.4 0**	1328.9 1**	136.00 **	1.31*	351.67 **	236.10 **	921.39 **	39.95* *	9.36*	11.45*	7.53**	2.38*	15.66 **
a x b	30	15.44* *	97.17* *	171.24 **	0.57	12.89**	105.30 **	78.50* *	100.12 **	8.13* *	42.23* *	11.08* *	101.89 **	22.93* *	8.22*	0.75	1.58*	9.36* *	28.34 **
Error	88	4.24	5.06	26.10	1.69	0.005	1.26	0.582	0.56	62.33	39.727	6.07	3.86	0.48	0.008	0.007	0.023	2.47	9.75

Table 1: Analysis of variance for Agro-morphological traits in 11 rice genotypes under different submergence (0, 7, 14, 21 days) regimes

Estimates from scaling tests

The gene action of the traits undertaken in this study were investigated based on simple additive × dominance model to understand the pattern of the gene action of the target trait (Sub1) as well as basic traits of Hasanta to be recovered in end product (Table-2). The scaling test showed all scale, A, B,

C and D were significant for almost all the traits except panicle weight (B, C and D), Number of chaffs per plant (A, B and D), spikelets fertility percent (A and B), test weight and grain yield per plant (A, C and D), Kernel length (B, C and D), Kernel width (A, B and D), L/B ratio (A, C and D), HRR% (A, B and C). The all studied traits indicates the

presence of non-alleliec interaction. All the basic traits like duration, plant height, fertility, grain type and value added trait, submergence tolerance were significant in more than three scales or in combination indicates the presence of epistatic between the genes responding these traits.

Table 2: Scaling test and generation mean analysis for yield, quality and submergence traits in the parents P1, P2 and combinations F1, F2, B1
and B2 of Hasanta/Swarna Sub1

The start		Scal	ing test		Generation mean analysis								
Traits/ Parameters	А	В	С	D	m (Hayman)	d (Hayman)	h (Hayman)	i (Add × Add)	j (Add × Dom)	l (Dom × Dom)	X ² test		
DFF	12.48**	1.22**	3.71**	2.32**	5.45**	22.65**	-15.55**	-1.82	18.65**	17.25**	142.13**		
DM	16.45**	10.09**	-25.50**	-22.52**	49.21**	-16.01**	12.34**	57.74**	-2.9	2.17**	1604.87**		
PH(cm)	13.6**	8.08**	-4.30**	22.36**	52.78**	2.92**	2.95**	3.18**	4.91**	23.42**	872.33**		
FLL (cm)	37.89**	27.34**	24.32**	10.6**	31.07**	52.59**	1.78**	2.81**	-5.03**	1.69**	1034.27**		
FLW (cm)	13.55**	-12.7**	15.08**	8.12**	1.44**	54.72**	37.89**	24.65**	13.88**	24.01**	1436.02**		
NETPP	8.25**	6.88**	7.29**	-22.54**	11.44**	27.67**	-0.42	0.16**	-11.64**	4.88**	659.25**		
PL (cm)	28.25**	48.17**	12.00**	8.66**	52.61**	4.13**	3.79**	-1.72	2.5**	85.33**	1120.45**		
PW (g)	-4.11	23.65**	15.6**	0.48**	4.08**	12.08**	12.47**	5.18**	-0.18	1.55**	183.9**		
NGPP	4.56**	23.4**	4.18**	10.75**	1.55**	1.88**	12.9**	5.19**	0.98**	1.09**	427.25**		
NCPP	6.1**	-11.5**	-2.34	33.68**	49.93 **	3.9**	3.75*	7.6**	-7.39**	17.71**	1955.85**		
SF%	4.58**	0.6**	-0.48	-0.85	-14.87**	18.28**	-25.1**	23.42**	-1.98	-2.44	642.14**		
TW	0.82**	-0.22	1.25**	1.57**	1.22**	2.85**	-3.65	-1.65	-0.31	1.86**	120.82**		
GYPP (g)	0.36**	0.08	0.10**	0.09**	0.25**	1.62**	0.68**	-0.28	-0.14	0.87**	634.21**		
KL (mm)	0.018	2.48**	2.03**	12.6**	12.56**	9.63**	4.55**	0.65**	22.9**	3.84**	244.085**		
KW(mm)	6.24**	54.36**	-2.80	7.55**	61.25**	-4.87	3.21**	-1.62	2.038**	35.43**	953.57**		
L/B Ratio	1.15**	-0.11	0.42**	0.17**	2.58**	1.47**	2.35**	1.98**	0.87**	1.68**	56.47**		
HRR%	8.45**	33.08**	7.25**	-2.44	22.78**	-2.55	40.9**	-2.44	-8.56**	1.76**	139.6**		
%SV	10.08**	10.95**	1.86**	3.95**	8.06**	7.22**	47.56**	4.36**	14.7**	0.92**	1208.52**		

In addition, goodness of fit of this model was confirmed by chi-square analysis. The abundancy of simple additivedominance model suggests the absent of non-allelic interaction effect (epistasis) and means value of different generations depends only on additive \times dominance gene interaction or effect of the gene. The chi-squire analysis showed that all the 19 studied traits of cross derivatives of Hasanta and Swarna Sub1 had significant value, indicates existence of epistatic effect among these traits. The results of this study revealed that the data does not fit into simple additive \times dominance model; role of epistatic is prevalent which is not fit into three parameter model, hence further subjected for six-parameter model analysis.

Six-parameter analysis

Digenic non-allelic interaction model consisting six parameters, m, d, h, i, j and l showed adequacy of the epistatic model to explain the gene action in the traits days to fifty percent flowering, days to maturity, plant height, flag leaf length, flag leaf width, number of effective tillers, panicle length, panicle weight, spikelets fertility, test weight, grain yield per plant, quality traits and percent survibility. The gene effect value was clearly illustrated the high variation among the observed agro-morphological traits (Table-2). The means and additive components for all studied traits were found highly significant.

The six-parameter analysis of Hasanta/Swarna Sub1 revealed the dominance (h) and dominance \times dominance (l) gene interaction or effects opposite signs for the traits *viz.*, days to fifty percent flowering, number of effective tillers per plant and test weight were fit into the duplicate type of epistatic. Whereas, values of most of the traits showed same sign for h and l scale were fit into complementary non-alleliec model.

The type of the gene interactions is depending on weight and sign of the estimates of 'h' and 'l' scale or effects in the case

many genes pair interacting. The sign of the estimates of 'd' and 'h' scale indicates about the parent which involves high number of genes for a trait. The additive effects and dominant \times dominant (l) interaction are found to be significant part of gene controlling grain yield and spikelets fertility in rice. The results of this study revealed that additive and dominance effects were substantial in expression of submergence tolerant in rice. The plus sign of additive gene effect infers that Hasanta contributes positively to the trait. The positive sign for the scale 'd' was in most of the traits except days to maturity, kernel length and HRR%, whereas, it was recorded negative 'h' in the traits days to fifty percent flowering, number of effective tillers, spikelets fertility and grain yield per plant (Table-1).

Characterizing genetics of the yield and contributing traits is prerequisite for planning successful breeding strategies in crop plants. The fundamental concept for analyzing genetics and gene action in crop plants is based on 'generation mean analysis' (Hayman, 1958 and Jinks and Jones 1958) ^[10, 13] which can detect epistasis/non-alleliec effect of genes. It is reported that additive and dominance and epistatic effects contributes substantially towards mean of the population (Viana, 2000) ^[35]. Though, epistatic effects creates biasness in the estimation of additive and dominance components which depends mainly on the relative values of epistatic. Analyzing epistasis is very critical, hence analyses priortingly before advance estimation for gene effect.

The traits studied were significant for more than two scale representing the presence of non-alleliec interactions among the genes. The abundancy of simple additive-dominance-model advocates the absent of epistasis effects. The means value of different generations is depending on additive \times dominance gene interaction or effect. The chi-squire test showed all 19 traits of the Hasanta/Swarna Sub1 derivatives (all six generations) were significant which indicates presence

of epistatic among these traits. This finding has full corroboration with the findings of Mahalingam and Nadarajan, 2010; Gnanamalar and Vivekanandan, 2013; and Kiani, 2013 ^[18, 6, 15].

Epistatic model with six-parameters revealed that the it was adequate to elucidate the gene action in most of the traits studied. The assessed gene effect evidently explains the high variation in all traits. The mean and additive variances for days to fifty percent flowering, days to maturity, plant height, number of ear bearing tillers, panicle length, number of grain per panicle, spikelets fertility, test weight, grain yield per plant, % survibility, head rice recovery, kernel length, kernel breadth, and kernel L/B ratio were highly significant (Murugan and Ganesan 2006) [20]. These results are in conventionality with the previous findings of Kiani et al. (2013)^[15] and Divya et al. (2014)^[5] for plant height, number of tillers, panicle length, days flowering, etc. It is found in this study that genic effects were cross-specific and fits to the complementary recessive interaction for most of the traits (Thirugnanakumar et al., 2007, Divya et al. 2014)^[34, 5].

The result of our study has full corroboration with the findings of Divya *et al.* (2014) ^[5] who has reported complementary interaction for most of the quantitative traits.

Conclusion

The results of this study has revealed the presence of high number of genic factors are involved in breeding value. Additive (d) effect and dominant \times dominant (l) epistatic were the significant part of gene controlling grain yield per plant, spikelet fertility and survibility under submergence. The additive and dominance genic effects were found substantial in controlling submergence tolerance in rice. The plus sign in the additive gene effect implies that Hasanta contributes positively than the Swarna.

Reference

- Adkins SW, Shiraishi T, McComb JA. Submergence tolerance of rice, a new glasshouse method for the experimental submergence of plants Plant Physiology. 1990; 80:642-646
- 2. Azizi F, Rezai AM, Saeidi G. Generation mean analysis to estimate genetic parameters for different traits in two crosses of corn inbred lines at three planting densities, J Agric Sci Technol. 2006; 8:153–169.
- 3. Catling D. Rice in deep water International Rice Research Institute, Manila, Philippines, 1992.
- Dash AK, Rao RN, Rao GJN, Verma RL, Katara JL, Mukherjee AK *et al.* Phenotypic and Marker-Assisted Genetic Enhancement of Parental Lines of Rajalaxmi, an Elite Rice Hybrid Front Plant Sci. 2016; 7:1005. doi: 103389/fpls201601005.
- Divya B, Biswas A, Robin S, Rabindran R, Joel AJ. Gene interactions and genetics of blast resistance and yield attributes in rice (Oryza sativa L), J Genet. 2014; 93(2):415–424.
- Gnanamalar RP, Vivekanandan P. Genetic architecture of grain quality characters in rice (Oryza sativa L), Eur J Experi Biol. 2013; 3(2):275–279.
- 7. Gurdev SK. Rice Breeding: Accomplishments and Challenges for future food security, Ritsumeikan International Affairs. 2006; 4:25-36.
- 8. Hayman BI, Mather K. The description of genetic interaction in continuous variation, Biometrics. 1955;

16:369–381.

- 9. Hayman BI. Maximum likelihood estimation of genetic components of variation, Biometrics. 1960; 16:369–381.
- Hayman BI. The separation of epistatic from additive and dominance variation in generation means, Heredity. 1958; 12:371–390.
- Iftekharuddaula KM, Ahmed HU, Ghosal S, Moni ZR, Amin A, Ali MS. Development of a New Submergence Tolerant Rice Variety for Bangladesh Using Marker-Assisted Backcrossing Rice Science. 2015; 22:15-26.
- 12. IRRI. International Rice Research Institute: Standard evaluation system for rice (SES) 5 Los Baños: International Rice Research Institute, 2014.
- 13. Jinks JL, Jones RM. Estimation of components of heterosis, Genetics. 1958; 43:223–234
- 14. Khush GS. Origin, dispersal, cultivation and variation of rice Plant Molecular Biology. 1997; 35:25–34.
- 15. Kiani SH, Kazemitabar SK, Jelodar NAB, Ranjbar GA. Genetic evaluation of quantitative traits of rice (*Oryza* sativa L) using generation mean analysis, Int J Agri Crop Sci. 2013; 5(19):2329–2336
- 16. Kumar A, Nayak AK, Hanjagi PS, Kumari K, Vijaykumar S, Mohanthy S *et al.* Submergence stress in rice: Adaptive mechanism, coping strategies and future research needs Environmental and Experimental Botany. 2021; 186:104448.
- 17. Kumar M, Singh RP, Singh ON, Singh P, Arsode P, Jena D *et al.* Generation mean analysis for bacterial blight resistance and yield traits in rice Journal of Pharmacognosy and Phytochemistry. 2019; 8(4):2120-2124.
- Mahalingam L, Nadarajan N. Genetic analysis of grain quality characteristics of two line rice hybrids, Elect J Plant Breed. 2010; 1(4):983–988.
- 19. Mather K. Biometrical genetics, Dover Publication, Inc, New York, 1949.
- 20. Murugan S, Ganesan J. Generation mean analysis in rice Oryza sativa L crosses utilizing WA cytosteriles, Plant Arch. 2006; 6(1):165–167.
- Nelder JA. The estimation of variance components in certain types of experiment on quantitative genetics, Biometrical Genetics (Eds Kempthorne, O), Pergaman Press, London, 1960, 139–158p.
- 22. Palada MC, Vergara BS. Environmental effects on the resistance of rice seedlings to complete submergence Crop Science. 1972; 12:209-212.
- 23. Pradhan SK *et al.* Pyramiding of three bacterial blight resistance genes for broad-spectrum resistance in deepwater rice variety, Jalmagna Rice, 2015. https://doiorg/101186/s12284-015-0051-8.
- 24. Pradhan SK, Pandit E, Pawar S, Baksh SY, Mukherjee AK, Mohanty SP. Development of flash-flood tolerant and durable bacterial blight resistant versions of mega rice variety 'Swarna' through marker-assisted backcross breeding Scientific Reports. 2019; 9:12810. https://doiorg/101038/s41598-019-49176-z.
- 25. Rajpurohit D, Kumar R, Kumar M, Paul P, Awasthi A, Osman BP *et al.* Pyramiding of two bacterial blight resistance and a semi dwarfing gene in Type 3 Basmati using marker-assisted selection Euphytica. 2011; 178:111–126.
- 26. Rao M, Grithlahre SP, Bisen P, Loitongbam B, Dar MH, Zaidi NW *et al.* Generation mean analysis for grain yield

and its component traits in submergence rice. SABRAO J Breed Genet. 2017; 49(4):327-335.

- 27. Ronald PC, Albano B, Tabien R, Abenes MLP, Wu KS, McCouch SR. Genetic and physical analysis of the rice bacterial blight disease resistance locus Xa21 Mol Gen Genet. 1992; 236:113–120.
- Rout D, Jena D, Singh V, Kumar M, Arsode P, Singh P et al. Hybrid Rice Research: Current Status and Prospects Recent Advances in Rice Research, 2020, DOI: 105772/intechopen93668.
- Samanta P, Ganie SA, Chakraborty A, Dey N. Study on regulation of carbohydrate usage in a heterogeneous rice population under submergence J Plant Biochem Biotechnol, 2020, https://doiorg/101007/s13562-020-00577-6
- Sanchez AC, Brar DS, Huang N, Li Z, Khush GS. Sequence tagged site marker-assisted selection for three bacterial blight resistance genes in rice Crop Sci. 2000; 40:792–797.
- Sarkar RK, Das KK, Panda D, Reddy JN, Patnaik SSC, Patra BC *et al.* Submergence tolerance in rice: Biophysical Constraints, Physiological basis and Identification of Donors Central Rice Research Institute, Cuttck, India, 2014, 36p.
- 32. Sekar I, Pal S. Rice and wheat crop productivity in the Indo-Gangetic plains of india: changing pattern of growth and future strategies Ind J Agric Eco. 2012; 67(2):238–252.
- Singh P, Sinha AK. A Positive Feedback Loop Governed by SUB1A1 Interaction with MITOGEN-ACTIVATED PROTEIN KINASE3 Imparts Submergence Tolerance in Rice Plant Cell. 2016; 28:1127-43 doi: 101105/tpc1501001 Epub, Apr 14 PMID: 27081183, PMCID: PMC4904673
- Thirugnanakumar S, Narassiman R, Senthil N, Eswaran R, Kumar CP. Gene effects for grain yield and its component characters in rice (Oryza sativa L), Crop Imp. 2007; 34:19–23.
- 35. Viana JMS. Generation mean analysis to polygenic systems with epistasis and fixed genes, Pesq Agropec Bras, Brasília. 2000; 35:1159-1167.
- 36. Xu K, Xu X, Fukao T, Canlas P, Maghirang-Rodrigue R, Heuer S *et al.* Sub1A is an ethylene-response-factor-like gene that confers submergence tolerance to rice Nature. 2006; 442:705-708.
- Xu K, Mackill DJ. A major locus for submergence tolerance mapped on rice chromosome 9 Mol Breed. 1996; 2:219–224