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Heredity divergence and genetic components of local potato (*Solanum tuberosum* L.) cultivars as revealed by molecular markers and quantitative traits

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Abstract

Molecular marker association with morphological traits have crucial role for heredity enhancement of local cultivars in different crops. In the present investigation, local potato cultivars were characterized and evaluated using SSRs markers and quantitative traits to recover the heredity divergence and the genetic components were estimated. All seven SSRs were showed 100% polymorphism with 3.71 allele per locus in the study. Based on different marker efficiency parameters, SSR0707 and STI0012 were most efficient primer to distinguish the local potato cultivars. The results of allelic diversity, genetic differentiation and AMOVA indicated that the genetic variability is greater within the population of potato cultivars. All the genotypes were grouped into three cluster as revealed by the Dendrogram utilizing SSRs. Different genetic components and principal component analysis using nine morphological traits suggested that average tuber weight, average tuber length, number of tubers per plant and plant height are potential traits for early generation selection by breeder to develop high tuber yielding potato. Total two phenotypic groups were found based on average silhouette method using nine quantitative traits in the current population. The knowledge acquired from this study, will be helpful for genetic improvement of local potato cultivars by plant breeders.

Keywords: Potato (*Solanum tuberosum* L.), Simple Sequence Repeats (SSR), quantitative traits, genetic components and average silhouette method

Introduction

Potato (*Solanum tuberosum* L.) is the top fourth food crop after maize, rice and wheat (Zeng *et al.*, 2020) [56]. India is the second country after China to produce large quantities of potatoes *i.e* 12% of global production and Punjab, Haryana, Uttar Pradesh, Bihar, West Bengal, Gujarat and Madhya Pradesh are major contributing states (Reddy *et al.*, 2018) [45]. India is one of the priority areas for investment in potato research and innovation (Devaux *et al.* 2014) [13]. According to the Global Hunger Index (GHI), forty seven countries still have 'serious' and 'alarming', and one in 'critically alarming' hunger levels (IFPRI 2019) [29]. So, the availability of nutritious food brings down the hunger value and therefore GHI as well. Potato is a wholesome and versatile food with more favourable overall nutrient-to-price ratio than most of the fruits and vegetables (Drewnowski and Rhem, 2013) [15]. Furthermore, potatoes produce more food per unit of water and cropland in less time than any other major crop and are up to seven times more efficient in using water than grains (FAO 2009 & NPC 2016) [19, 41]. Potato is grown as a staple food in agriculture-based countries like India, and as a high-value crop in urbanized and transforming countries. Thus improvement of potato plays a key role in improving global food security and reducing poverty (Devaux *et al.* 2014) [13]. Little or no efforts have been made to characterize local cultivars of potato in the state West Bengal of India. Modernization bottleneck leads to genetic erosion (Louwaars 2018) [34], generally by ignorance towards landraces and local cultivars. Collection, characterization, and classification of germplasm with morphological and molecular markers are crucial for any crop improvement (Govindaraj 2014) [26]. So identification of genetic diversity through proper investigation of local potato cultivars and subsequent utilization of useful cultivars in crop improvement is imperative (Ferguson 2007) [21]. Genetic relatedness of potato genotypes was most extensively investigated through naked eye polymorphism (Kolech *et al.*, 2016; Mishra *et al.* 2017; Anouma *et al.* 2017; Nasiruddin *et al.* 2017; Berdugo-Cely *et al.* 2017; Prabha *et*

al. 2018) [33, 37, 7, 38, 9, 44] and DNA markers like SSRs (Bali *et al.* 2018; Wang *et al.*, 2019; Cruz *et al.* 2020) [8, 52, 11], EST-SSR (Salimi *et al.* 2016) [46], inter-simple sequence repeats (ISSRs) (Mahgoub *et al.* 2014) [35] AFLP (Wang *et al.*, 2013) [51], SNPs (Kolech. 2016; Berdugo-Cely *et al.* 2017) [33, 9] and RAPD (Hoque *et al.* 2013) [28]. Among them SSRs being the most common as they are co-dominant, randomly distributed throughout the genome, highly polymorphic, high reproducibility, less cost of operation, hyper-variability, amenable to automation, ease of multiplexing, highly conserved across related species and use with low quality DNA (Ghislain *et al.* 2009; Mason 2015; Yang *et al.* 2015; Jian *et al.* 2017; Duan *et al.* 2018) [23, 54, 31, 16]. To explore the extensive knowledge of heredity structure and yield assessment, consolidation of phenotypic and SSR markers have been established their readiness in rice (Ahmad *et al.*, 2015) [5]. Consequently, the present investigation was carried out to obtain the genetic heterogeneity using SSRs marker and quantitative traits in local potato cultivars. In addition, the genetic components of quantitative traits were also estimated to sort out the potential cultivars for future breeding programme for improvement of local potato.

Materials and Method

Plant materials and design of experiments

Fourteen potato cultivars including ten local potato clones and four released varieties were considered in this study. The genotypes were collected from northern part of West Bengal, India by Central Germplasm Conservation Unit, Directorate of Research, Uttar Banga Krishi Viswavidyalaya. The present investigation was conducted at Agricultural Research Farm, Regional Research station, Terai zone, Pundibari, Cooch Behar, West Bengal during rabi season of 2017-18 and 2018-19. Complete randomized block design (CRBD) with three replications was adopted to successfully carry out the experiment.

Morphological data collection

The quantitative value of eight morphological characters *viz.* sprout length (SL), plant height (PH), leaf length (LL), leaf width (LW), number of tubers per plant (NT), tuber length (TL), tuber weight (TW) and tuber yield per plant (TY) were recorded on five randomly selected plants, excluding the plants located at borders to avoid border effect so that highest precision could be achieved. The germination percentage of potato tubers was calculated as described by Clarke and Stevenson, 1943 [10]. All the data were recorded from three replication of the experimental plot and their average value of each replication were used for statistical analysis.

DNA isolation and PCR amplification

DNA was extracted from 80 mg fresh leaf samples of 14 potato genotypes following the method of Mandal *et al.*, 2016 [36]. The extracted DNA samples were stored at -20°C until use. The quality of extracted DNA was analyzed by 0.8%

agarose gel electrophoresis in TBE buffer, followed by ethidium bromide staining and visualized in gel documentation system. The purity of the DNA was estimated by spectrophotometry observing A260nm/A280nm ratio, and the yield was estimated by measuring absorbance at 260 nm.

Seven SSR primer sets distributed on seven chromosome developed by Ghislain *et al.*, 2004; Ghislain *et al.*, 2009 [24]; Xiaoyan *et al.*, 2016 [49] were used for this study and were synthesized by Sigma-Aldrich, India. PCR amplification was carried out in a volume of 25 µl containing 2 µl of 10 ng/µl DNA, 1 µl of each primer (100 ng/ µl), 1 µl of 2.5 mM dNTPs, 2.5 µl of 10X reaction buffer (Thermo Fisher Scientific, India), 0.2 µl of 5 U/µl Taq polymerase (Thermo Fisher Scientific, India) and 16.8 µl HPLC grade water. PCR reactions were performed in Veriti Thermal Cycler (Applied Biosystems, USA), an initial step for 5 min at 94°C with 35 cycles for 1 min at 94°C, 45 s at 52-58°C and 1 min at 72°C, and a final step of 5 min at 72°C. The PCR products were separated on 1.5% agarose gel with ethidium bromide staining and visualized in UV gel documentation system.

Data analysis

Marker efficiency parameters for SSRs *viz.* polymorphic information content (PIC), marker index (MI), resolving power (RP), discriminating power (D_j) and effective number of patterns per assay unit (P_i) were calculated using Microsoft Excel as described by Zargar *et al.*, 2016 and Mandal *et al.*, 2016 [36]. Allelic diversity and analysis of molecular variance were performed with the help of Gen AIEx 6.5 software (Peakall and Smouse, 2012). Genetic differentiation (G_{ST}) and gene flow (N_m) were estimated utilizing Pop Gen v 1.32 software (Yeh, 1997). Cluster analyses were implemented by UPGMA method, and the corresponding dendrogram was constructed using DARwin v 6.0 software. In order to estimate the goodness of fit between similarity matrix and the dendrogram, the coefficient of cophenetic correlation was calculated with help of R v 3.6.0 software with “dendextend” package. Phenotypic data analysis were implemented through R v 3.6.0 software using “Agricolae” package.

Results

Polymorphism and marker efficiency

A total 26 number of allele revealed by seven pairs of SSR markers, which were used for molecular characterization of 14 potato cultivars in the present investigation. The amplified product size ranged from 60 bp (STI0032) to 250 bp (STM2022) with an average number of allele per marker 3.71. In the current study, all the marker showed 100% polymorphism and no rare allele (less than 5% among the cultivars) was observed. To estimate the performance of SSRs markers different efficiency parameters *viz.* polymorphic information content (PIC), resolving power (RP), marker index (MI), discriminating power (D_j) and effective number of patterns per assay unit (P_i) were calculated in this investigation (Table 1).

Table 1: Estimation of different marker efficiency parameters revealed from SSRs profiling

Marker Id	Allele size (bp) range	NTA	NPA	NTB	PIC	RP	MI	D_j	P_i
STG0016	130-180	4	4	18	0.20	1.14	3.67	0.24	1.26
STM2022	190-250	5	5	17	0.39	1.57	6.68	0.51	1.65
SSR0707	130-180	3	3	14	0.56	2.00	7.86	0.60	2.28
STI0012	170-190	3	3	13	0.61	1.57	7.89	0.65	2.55
STI0032	60-140	3	3	15	0.48	1.57	7.27	0.53	1.94

SSR0675	90-180	5	5	22	0.09	1.71	2.02	0.14	1.10
PM0938	120-160	3	3	19	0.12	1.29	2.23	0.15	1.13
Mean		3.71 ± 0.36	3.71 ± 0.36	16.86 ± 1.18	0.35 ± 0.08	1.55 ± 0.11	5.37 ± 1.00	0.40 ± 0.08	1.70 ± 0.22

[NTA = Number of total Allele; NPA = Number of polymorphic Allele; NTB = Number of total bands; PIC = Polymorphic information content; RP = Resolving power; MI = Marker Index; D_j = Discriminating power; P_t = effective number of patterns per assay unit]

The PIC values varied from 0.09 (SSR0675) to 0.61 (STI0012) with an average 0.35 per locus. The estimates of resolving power were found to be the highest for the marker SSR0707 (2.00), followed by SSR0675 (1.71) and was lowest for the marker STG0016 (1.14). The average MI value was 5.37 per SSR and ranged from 2.02 for SSR0675 to 7.89 for STI0012. The highest D_j was found for STI0012 (0.65) and lowest was for SSR0675 (0.14) with an average value of 0.40. The maximum number of P_t were observed for STI0012 and minimum number were for SSR0675 with the average value 1.70 which informed that the individual SSR markers have the ability to acclimated almost 2 accessions within the infinite population size of potato. Association analysis showed that all

the efficiency parameters have positive significant correlation with each other with the except of resolving power.

Allelic diversity, AMOVA and gene differentiation

Allelic diversity was obtained from the two groups of potato accessions using the average number of different allele (N_a), average number of effective allele (N_e), Nei's gene diversity (h) and Shannon's information index (I). Group 1 which included local potato cultivars showed maximum N_a and highest value for I whereas group 2 showed the maximum N_e, greater value for h and uh. N_a, N_e, I, h and uh value for overall total potato collection were 1.48, 1.47, 0.40, 0.27 and 0.33 respectively.

Table 2: Estimated different allelic diversity parameters based on 26 allele of SSRs markers

Population	N	N _a	N _e	I	h	uh
Local Potato	10.000	1.654±0.15	1.423±0.06	0.407±0.05	0.265±0.03	0.295±0.03
Released varieties	4.000	1.308±0.18	1.508±0.09	0.391±0.06	0.274±0.04	0.365±0.06
Total	7.000	1.481±0.12	1.465±0.05	0.399±0.04	0.270±0.03	0.330±0.03

[N = Number of potato clones; N_a = Number of different Alleles; N_e = Number of effective Alleles; I = Shannon's information index; h = Nei's gene diversity; uh = Unbiased Nei's gene diversity]

The results of analysis of molecular variance (AMOVA) showed that 93% variation within the population and 7% variation among the population (groups). The gene

differentiation value (G_{ST}) and gene flow (N_m) between the two potato groups were 0.1241 and 3.5283 respectively.

Table 3: Analysis of molecular variance among the two groups of potato revealed by SSRs markers

Source	df	SS	MS	Est. Var.	%	F _{ST}	P(rand >= data)
Among Pops	1	5.679	5.679	0.283	7%		
Within Pops	12	48.750	4.063	4.063	93%		
Total	13	54.429		4.345	100%	0.06	0.183

Cluster analysis based on SSRs

All the fourteen potato cultivars were grouped into three cluster at the similarity coefficient of 0.65 as the result of dendrogram analysis based on UPGMA method (Figure 1). Cluster I had nine number of genotypes followed by cluster

III which included three number of genotypes and cluster II containing only two genotypes. The range of genetic similarity was observed between the cultivars from 46% (Kufri Arun and LPCH-01; LPCK-01 and K. Poukhraj) to 100% (LPCD-01 and LPCD-02).

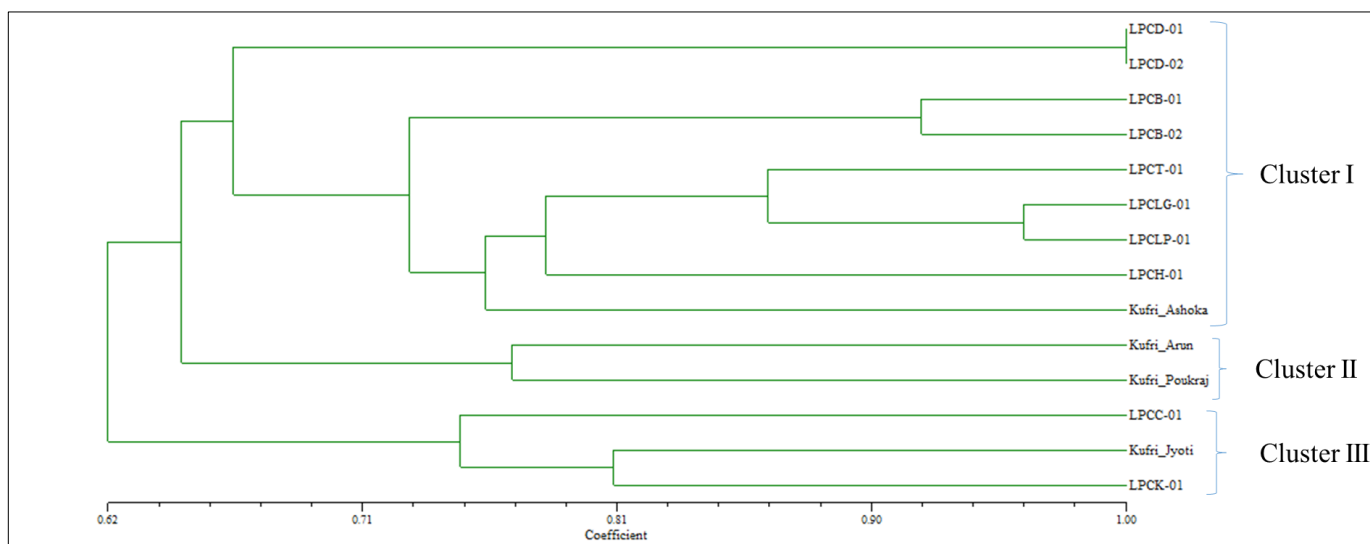


Fig 1: A dendrogram accomplished using 26 allele of seven SSRs

Phenotypic variability

Nine morpho-phenetic quantitative characters *viz.* germination percentage (GP), sprout length (SL), plant height (PH), leaf length (LL), leaf width (LW), number of tubers per plant (NT), average tuber length (TL), average tuber weight (TW) and yield per plant (TY) were evaluated in two successive 'rabi' season on 14 potato cultivars. The result of ANOVA explained that all the cultivars had significant variation ($p < 0.01$) based on the morpho-phenetic characters. The cultivars LPCD-01 showed the shortest sprout length (1.00 cm), lowest plant height (15.38 cm), minimum leaf length (10.26 cm),

smallest tuber length (1.78 cm) and lower value for tuber weight (1.59 g). Minimum germination percentage (92%), lowest leaf width (5.81 cm) and maximum average number of tubers per plant (40.82) were found in the genotypes LPCD-02. The cultivar LPCLP-01 had the prolonged sprout length (1.82 cm), least average number of tubers per plant (2.88) and lowest tuber yield per plant (394.50 g). Longest plant height (42.40 cm), highest average tuber weight (6.05 g) and maximum tuber yield (3080.83 g) were obtained by the cultivars Kufri Arun.

Table 4: ANOVA represented mean square value of nine quantitative traits

Characters	Sources of variance					Mean
	Mean sum of squares					
	Rep	Genotype	Year	Genotype × Year	Residual	
Degrees of freedom (DF)	2	13	1	13	54	
Germination percentage (GP)	0.16	34.61**	5.25	0.28	5.29	98.06
Sprout length (SL)	0.00	0.32**	0.00	0.01	0.05	1.43
Plant height (PH)	6.30	312.44**	0.96	0.48	4.56	23.73
Leaf length (LL)	6.98	87.01**	0.06	0.15	1.18	16.04
Leaf width (LW)	6.51	57.66**	0.12	0.07	0.54	11.01
Number of tubers per plant (NT)	4.09	977.08**	2.40	0.72	5.40	10.10
Average tube length (TL)	0.42	23.53**	0.05	0.01	0.16	5.61
Average tuber weight (TW)	0.19	9.42**	0.50	0.02	0.09	4.22
Tuber yield per plant (TY)	27816.00	155134.00**	7180.00	1425.00	4007.00	245.20

Genetic components and association among the nine quantitative traits

In the present study, GCV and PCV ranged from 2.07% (GP) to 125.88% (NT) and from 3.07% (GP) to 127.19% (NT) respectively. Among the nine traits, PH, LL, LW, NT, TL, TW and TY were showed the higher percentage (> 20%) of

GCV and PCV whereas, displayed moderate GCV and PCV value. The broad sense heritability varied from 45% (GP) to 98% (LW, NT, and TL) for nine morpho-phenetic traits. NT (256.63%) showed the uppermost value for genetic advance followed by TY (125.29%) whereas, GP have lowest value (2.87%).

Table 5: Predicted genetic components for different quantitative traits in potato

Characters	GCV (%)	PCV (%)	ECV (%)	H ² _{bs}	GAM (%)
Germination percentage (GP)	2.07	3.07	2.27	0.45	2.87
Sprout length (SL)	14.17	19.95	14.05	0.50	20.73
Plant height (PH)	30.04	31.13	8.14	0.93	59.74
Leaf length (LL)	23.49	24.23	5.95	0.94	46.91
Leaf width (LW)	28.04	28.38	4.37	0.98	57.07
Number of tubers per plant (NT)	128.88	127.19	18.25	0.98	256.63
Average tube length (TL)	35.15	35.57	5.47	0.98	71.54
Average tuber weight (TW)	29.51	29.92	4.95	0.97	59.96
Tuber yield per plant (TY)	64.32	68.01	22.12	0.89	125.29

The result of genotypic and phenotypic correlation explained that, PH, LL, LW, TL and TW had the highly significant ($p < 0.01$) positive correlation with TY. NT showed the significant ($p < 0.05$) negative correlation with TY at genotypic level but phenotypically the relationship was non-significant.

Principal Component analysis

First three PCs with Eigen value greater than one acquired 92.41% of total variation as revealed from nine morphological traits. TW, TL, NT, LW and LL in the PC1, GP, TY, PH, SL and NT in the PC2, and PH, LW, LL and SL in the PC3 had individually more than 10% contribution towards the variability. According to the PCA-biplot, all the fourteen

cultivars were grouped into two distinct cluster. Cluster I had the two genotypes and rest of the genotypes were placed into the cluster II.

Genetic structure and dendrogram analysis

The average silhouette width illustrated a recognizable peak at 2 ($K = 2$) which predicted that, the 14 potato cultivars were classified into the two phenotypic groups (Figure 2) with nine morpho-phenetic traits. A dendrogram was constructed based on mean phenotypic performance with 14 potato cultivars using Euclidian distance method (Figure 3).

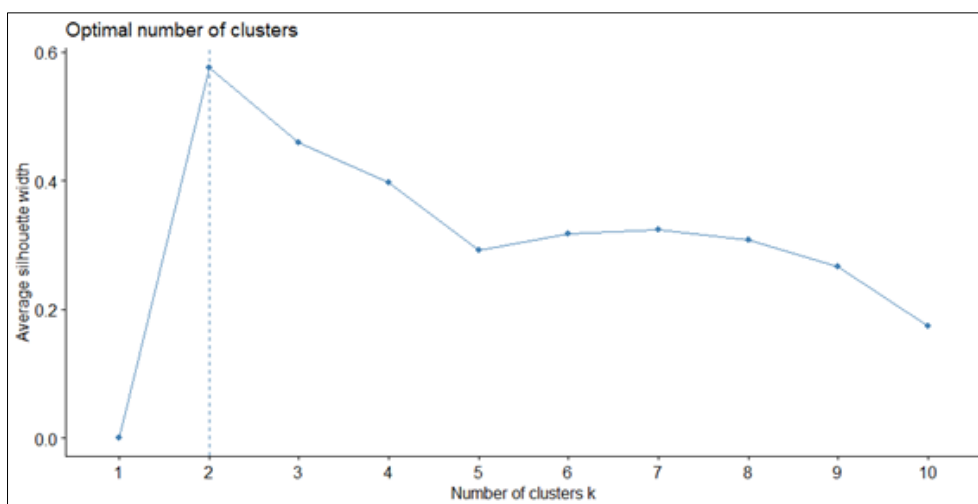


Fig 2: Estimation of optimum number of cluster using average silhouette method

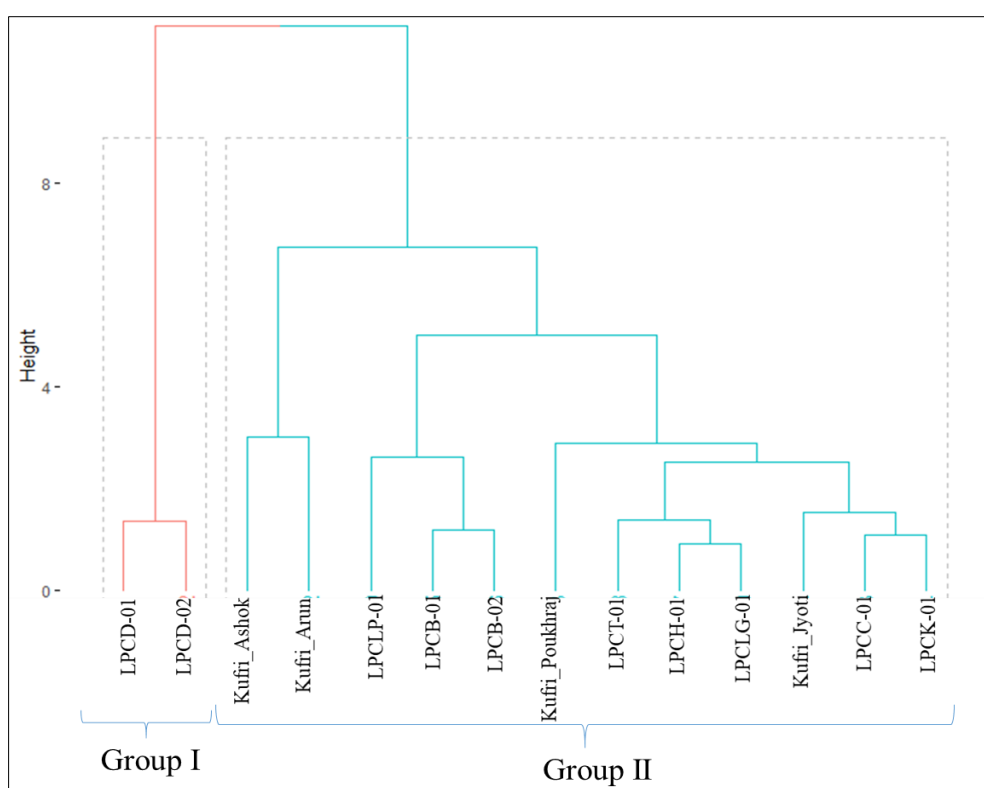


Fig 3: Dendrogram constructed from quantitative traits using Euclidian distance

Discussion

Estimation of genetic variability with molecular marker and morpho-phenetic traits together gives the in depth knowledge about the genetic structure and heredity information of local germplasm (Ahmad *et al.*, 2015 and da Silva *et al.*, 2017) ^[5, 12]. In the present study, seven pairs of SSRs markers which were distributed in seven number of chromosome and nine quantitative traits were used for the evaluation of 14 local potato cultivars. The amplified allele size produced by the SSR marker in 14 potato cultivars was minimum for STI0032 (60 bp) and maximum for STM2022 (250 bp). The results agreed with previous findings where the product size ranged from 106 bp (STI0032) to 244 bp (STM2022) reported by Xiaoyan *et al.*, 2016 ^[49]. Average allele per SSR found in this investigation was 3.71 which was relatively less from the earlier study by Xiaoyan *et al.*, 2016

^[49] (8.2 per SSRs) and Ghebreslassie *et al.*, 2016 (8.0 per SSRs). According to DeWoody *et al.*, 1995, A SSRs marker will be informative if their PIC value is more than 0.50. Based on this report, SSR0707 (0.56) and STI0012 (0.61) were the most significant SSRs markers for evaluation local potato cultivars in this investigation. In addition, STI0012 showed highest value for marker index, discriminating power and effective number of patterns per assay unit but moderate value for resolving power. The relationship among the different marker efficiency parameters suggested that all the parameters more or less equally important except resolving power. Hence, considering PIC, MI, D_j, and P_i most efficient SSRs were STI0012 followed by SSR0707 for evaluation of local potato cultivars under this study. For example, Xiaoyan *et al.*, 2016 ^[49] also found that STI0032, STI0012, SSR0707 and STG0016 were most informative SSR marker for

molecular dissection of 192 potato cultivars.

The fourteen potato clones were classified into two groups' viz. group I that included 10 local potato cultivars from northern part of West Bengal and group II with 4 released varieties from the CPRI, Shimla, India to estimate the allelic diversity and to acquire the satisfactory knowledge of gen diversity. Among the 26 number of allele revealed by seven SSRs markers, 22 were found in group I and 18 were found in group II with 14 number of common allele. Group I have higher number of different allele and lower number of effective allele as compared to group II. The Shannon information index (I) was greater for the group I which indicated higher allelic richness and homogeneous scattering of the allele within the local potato collections. Group II population showed the higher level of gene diversity in comparison with group I which explained that the released varieties have the broad genetic base. The genetic variability within population was significantly higher than the variation between the group I and group II potato collections as demonstrated by AMOVA. The genetic differentiation value also suggested that more numbers of identical potato clones are present in the current population. The gene flow between the local potato cultivars and released varieties was higher means evolutionary the genotypes of these two groups have close ancestry and narrow genetic diversity between the populations. Allelic diversity, genetic differentiation and AMOVA were also performed for exploring the genetic variability by Juyó *et al.*, 2015 [32] with Phureja potato collection using SSRs markers.

The dendrogram constructed by the genotyping data revealed from seven SSRs markers showed high range of genetic relatedness and formed three cluster. Cluster I consisted with eight number of local potato cultivars viz. LPCD-01, LPCD-02, LPCB-01, LPCB-02, LPCT-01, LPCLG-01, LPCLP-01 and LPCH-01 and one released variety (Kufri Ashok). Remaining three potato released varieties were placed in to the cluster II and cluster III whereas rest of the two local potato cultivar were situated in the cluster III. Among the potato clones of cluster I, LPCD-01 and LPCD-02 exhibited 100% similarity, LPCLG-01 and LPCH-01 showed 96.15% similarity and LPCB-01 and LPCB-02 demonstrated 92.31% genetic identity. The estimated cophenetic coefficient value of dendrogram was 0.80 which indicated that the excellent ability of genotypic data revealed from the SSRs profiling for genetic variability analysis (Ahmad *et al.*, 2015) [5].

Fourteen potato cultivars were also evaluated based on their nine quantitative characters recorded in successive years. Analysis of variance explained that all the traits have significant ($p < 0.01$) variation among the cultivar at genotypic level. The interaction between genotype and year was non-significant may be due to the consistent phenotypic performance of genotypes for all the traits in two subsequent years. An expanding range (2.88 to 40.82) of number of tuber per plant was found in local potato cultivars with an average

value 11.94 whereas the average number tuber per plant was 5.50 in release varieties. The average tuber length and tuber weight indicated that local cultivars (TL = 5.05 cm; TW = 3.87 g) have small size tuber in comparison with release varieties (TL = 7.02 cm; TW = 5.12 g). The yield performance of the release varieties (2059 g) was significantly ($p < 0.05$) higher than the local potato cultivars (892.92 g). In spite of low yield, farmers are more prefer the local potato cultivars than the released varieties due to some qualitative traits like colour (anthocyanin content), taste preference (total solid and starch content), disease resistance and long term storage capability. For instance, plant height, leaf area, number of tubers per plant, average tuber length and tuber yield showed significant variation among the potato accessions as previously reported by Nasiruddin *et al.*, 2017 and Fekadu *et al.*, 2013 [38, 20].

Among the 14 potato cultivars, number of tuber per plants showed the highest GCV and PCV with moderate ECV, suggestive that selection could be effective with these traits in future breeding programme. Yield have second highest GCV and PCV but showed the higher ECV value means the greater environment as influence on this may be misleading for selection by breeders. High H^2_{bs} with high GAM (%) were observed for the plant height, leaf length, leaf width, number of tubers per plant, tuber length, tuber weight and tuber yield which were inhibited by the additive gene action and will be beneficial for early generation selection by potato breeders (Gopal., 199; Adjah *et al.*, 2020) [25, 1].

Among the nine quantitative traits, plant height, leaf length, leaf width, tuber length and tuber weight showed significant positive relationships with tuber yield per plant and the higher degree of genotypic correlation coefficient value than the analogous phenotypic correlation suggested that selection made through these traits will be rewarding for development of high yielding genotypes. Identical findings in potato also made known by Nasiruddin *et al.*, 2017 [39] and Fekadu *et al.*, 2013 [20].

PCA have the important role to select the morphological traits for breeding programme in different crops including potato (Heberger *et al.* 2003; Placide *et al.* 2015; Yuan *et al.*, 2016) [27, 43, 55]. The results of PCA described, average tuber weight, average tuber length, number of tubers per plant, leaf length, leaf width, tuber yield per plant and plant height had the major contribution towards genetic variability in potato genotypes. These findings indicated that, genetic enhancement possible in local potato cultivars if selection was made through these phenotypic characters. This finding also is in agreement with genetic component analysis. PCA provided the necessary knowledge for development of high yielding potato genotype through selection of suitable morphological traits like tuber size (Yuan *et al.*, 2016 and Struik and Wiersema 1999) [55, 50]. Three cluster were found among the 14 potato genotypes with common phenotypic performance revealed by PCA-biplot (Figure 4).

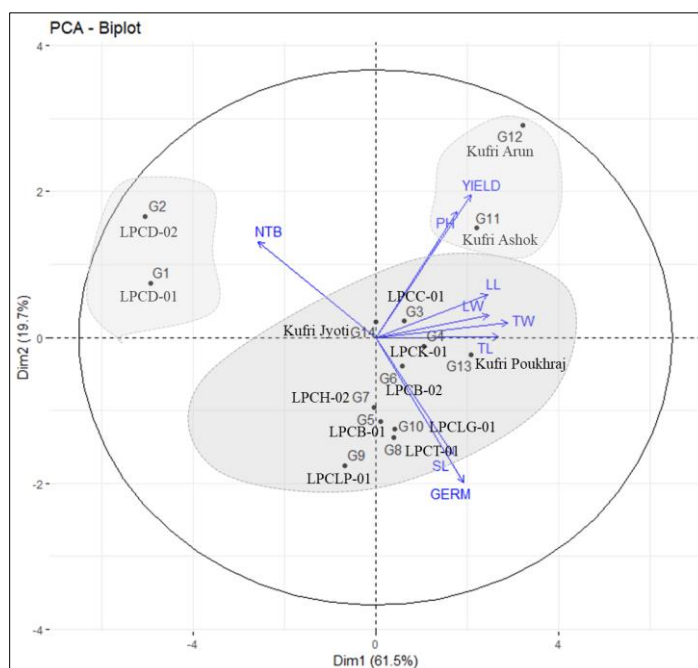


Fig 4: PCA-biplot revealed from the nine quantitative traits of 14 potato cultivars

Cluster I had two cultivars *viz.* LPCD-01 and LPCD-02 which are 100% genetically similar to each other based on SSRs profiling and produced higher number of tubers per plant. Cluster II consisted also two release varieties namely Kufri Ashok and Kufri Arun with high tuber yield potentiality. Cluster III contained ten genotypes with moderate phenotypic performance. This finding illustrated that hybridization between the genotypes of cluster I and cluster II may produce high yielding genotypes with greater number of tuber per plant. Cluster analysis based on PCA biplot generated through morphological traits were reported in pea (Espósito *et al.*, 2007) [18] and sweet potato (Afuape, 2011 and Afuape *et al.*, 2015) [2, 4].

The average silhouette width method presumed and validated

the number of rational cluster generated from morphological traits (Peter, 1987) [42]. This method divided the fourteen potato genotypes into two phenotypic groups (PG) *viz.* PG I included only two genotypes with small tuber size and greater number tubers per plant and PG II had twelve genotypes which have normal tuber size and higher tuber yield (Figure 5). This method was also used to find out the phenotypic population group in blackberry described by Jared *et al.*, 2019 [30]. Dendrogram obtained from the morphological data based on the Euclidian distance generated two cluster same as the average silhouette width method. The cophenetic coefficient value revealed from dendrogram was 0.99 (> 0.80) indicated that best fit of phenotypic data for analysis of diversity in potato.

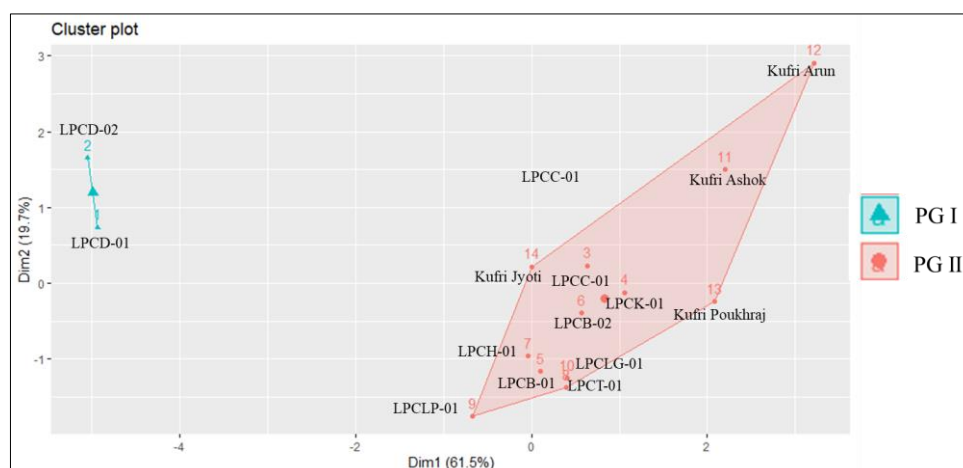


Fig 5: Cluster plot generated from nine quantitative traits using average silhouette method

Conclusion

Overall findings suggested that adequate genetic variability exist in the local potato cultivars taken into consideration in this study based on genotyping and phenotyping. Both SSRs and morphological markers together scrupulously established their competency to scrutinize the heredity information in

potato cultivars. SSRs markers also secured that genetic diversity was higher in released varieties whereas greater allelic richness and Shannon information index were higher in local potato collections. Genetic components analysis and PCA based on the quantitative traits illustrated that synchronous choice of tuber size (average tuber weight and

average tuber length), number of tubers per plant, leaf size (leaf length and leaf width) and plant height might be impressive in enhancement of tuber yield in local potato. Finally cluster study concluded that heredity intensification could be possible though hybridization programme between small sized potato cultivars (LPCD-01 and LPCD-02) and released varieties (Kufri Ashok and Kufri Arun).

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References

1. Adjah KL, Abe A, Adetimirin VO, Asante MD. Genetic variability, heritability and correlations for milling and grain appearance qualities in some accessions of rice (*Oryza sativa* L.). *Physiol Mol Biol Plants*. 2020;26:1309-1317. doi.org/10.1007/s12298-020-00826-x
2. Afuape SO, Okocha PI, Njoku D. Multivariate assessment of the agromorphological variability and yield components among sweet potato (*Ipomoea batatas* (L.) Lam) landraces. *Afr. J. Plant Sci*. 2011;5:123-132
3. Afuape S. Multivariate assessment of the agromorphological variability and yield components among sweetpotato (*Ipomoea batatas* (L.) Lam) landraces. *African Journal of Plant Science*. 2011;5:123-132.
4. Afuape S, Omodamiro RA, Njoku JC, Ogbonna CL, Uzuegbu DC. Targeted Breeding for Sweetpotato-Based Enterprises: Variability, Genotype-by-Environment Interaction, Heritability and Correlation Studies of Important Sweetpotato Root Processing Quality Traits. *International Journal of Plant Breeding and Genetics*. 2015;9:206-217. 10.3923/ijpb.2015.206.217.
5. Ahmad F, Hanafi MM, Hakim MA, Rafii MY, Aroliu IW, Akmar Abdullah SN. Genetic Divergence and Heritability of 42 Coloured Upland Rice Genotypes (*Oryza sativa*) as Revealed by Microsatellites Marker and Agro-Morphological Traits. *PLoS One*. 2015;10(9):e0138246. <https://doi.org/10.1371/journal.pone.0138246>
6. Ahmadzadeh M, Felenji H. Evaluating diversity among potato cultivars using agro-morphological and yield components in fall cultivation of Jiroft area. *Am. Eurasian J. Agric. Environ. Sci*. 2011;11:655-662
7. Anoumaa M, Kanmegne G, Kouam EB, Amzati GS, Yao NK, Fonkou T, et al. Characterization of Potato (*Solanum tuberosum* L.) Genotypes from the Western Highlands Region of Cameroon Using Morphological and Agronomic Traits. *Journal of Plant Sciences*. 2016;4(6):185-194. doi: 10.11648/j.jps.20160406.17
8. Bali S, Patel G, Novy R, Vining K, Brown C, Holm D, et al. Evaluation of genetic diversity among Russet potato clones and varieties from breeding programs across the United States. *PLoS One*. 2018;13(8):e0201415. <https://doi.org/10.1371/journal.pone.0201415>.
9. Berdugo-Cely J, Valbuena RI, Sañchez-Betancourt E, Barrero LS, Yockteng R. Genetic diversity and association mapping in the Colombian Central Collection of *Solanum tuberosum* L. Andigenum group using SNPs markers. *PLoS One*. 2017;12(3):1-27. Doi: 10.1371/journal.pone.0173039
10. Clarke AE, Stevenson FJ. Factors influencing the germination of seeds of the potato. *American Potato Journal*. 1943;20:247-258. <https://doi.org/10.1007/BF02881698>
11. Cruz GD, Miranda TY, Blas RH, Neyra E, Orjeda G. Simple Sequence Repeat-Based Genetic Diversity and Analysis of Molecular Variance among on-Farm Native Potato Landraces from the Influence Zone of Camisea Gas Project, Northern Ayacucho, Peru. *American Journal of Potato Research*, 2020. <https://doi.org/10.1007/s12230-020-09763-7>
12. da Silva MJ, Pastina MM, de Souza VF, Schaffert RE, Carneiro PCS, Noda RW, et al. Phenotypic and molecular characterization of sweet sorghum accessions for bioenergy production. *PLoS One*. 2017;12(8):e0183504. <https://doi.org/10.1371/journal.pone.0183504>
13. Devaux A, Kromann P, Ortiz O. Potatoes for Sustainable Global Food Security Potato Research. 2014;57:185-199. doi: 10.1007/s11540-014-9265-1.
14. DeWoody JA, Honeycutt RL, Skow LC. Microsatellite markers in white-tailed deer. *J Hered*. 1995;86:317-319. pmid:7658002
15. Drewnowski A, Rehm CD. Vegetable Cost Metrics Show That Potatoes and Beans Provide Most Nutrients Per Penny. *PLoS One*. 2013;8(5):e63277. doi:10.1371/journal.pone.0063277
16. Duan Y, Liu J, Xu J, Bian C, Duan S, Pang W, et al. DNA Fingerprinting and Genetic Diversity Analysis with Simple Sequence Repeat Markers of 217 Potato Cultivars (*Solanum tuberosum* L.) in China. *American Journal of Potato Research*. 2018;96:21-32. doi: 10.1007/s12230-018-9685-6
17. Dumnil J, DiMichele M. Plant Species delimitation: A comparison of morphological molecular markers. *Plant Biosyst*, 2009, 1-15.
18. Espósito MA, Milanese LA, Martin EA, Cravero VP, Anido FSL, COUNTRY EL. Principal Component Analysis Based on Morphological Characters in Pea (*Pisum sativum* L.) *international journal of plant breeding*. 2007;1(2):135-137
19. FAO. International year of the potato 2008: new light on a hidden treasure, 2009. <http://www.fao.org/potato-2008/en/events/book.html>
20. Fekadu A, Petros Y, Zelleke H. Genetic variability and association between agronomic characters in some potato (*Solanum tuberosum* L.) genotypes in SNNPRS, Ethiopia. *Inter J Biod and Conser*. 2013;5(8):523-528.
21. Ferguson AR. The need for characterisation and evaluation of germplasm: kiwifruit as an example *Euphytica*. 2007;154:371-382. doi: 10.1007/s10681-006-9188-2 123
22. Gana AS, Shaba SZ, Tsado EK. Principal component analysis of morphological traits in thirty-nine accessions of rice (*Oryza sativa* L.) grown in a rainfed lowland ecology of Nigeria. *J. Plant Breed. Crop Sci*. 2013;5:120-

23. Ghislain M, Núñez J, del Rosario Herrera M, Pignataro J, Guzman F, Bonierbale M, *et al.* Robust and highly informative microsatellite-based genetic identity kit for potato. *Molecular Breeding*. 2009;23:377-388. doi: 10.1007/s11032-008-9240-0
24. Ghislain M, Núñez J, Herrera MDR, Pignataro J, Guzman F, Bonierbale M, *et al.* Robust and highly informative microsatellite-based genetic identity kit for potato. *Molecular Breeding*. 2009;23(3):377-388.
25. Gopal J. Genetic parameters and character association for clonal selection in potato breeding programmes. *Agronomie*. 1999;19:531-539
26. Govindaraj M, Vetriventhan M, Srinivasan M. Importance of Genetic Diversity Assessment in Crop Plants and Its Recent Advances: An Overview of Its Analytical Perspectives. *Genetics Research International*. 2014;2015:14.
27. Heberger K, Csomós E, Simon-Sarkadi L. Principal component and linear discriminant analysis of free amino acids and biogenic amines in Hungarian wines. *J Agric. Food Chem*. 2003;51:8055-8060
28. Hoque ME, Huq H, Moon NJ. Molecular Diversity Analysis in Potato (*Solanum tuberosum* L.) through RAPD Markers. *SAARC J. Agri*. 2013;11(2):95-102.
29. IFPRI. Global food security index, the challenge of hunger: building resilience to achieve food and nutrition security. The International Food Policy Research Institute (IFPRI), Concern Worldwide, and Welthungerhilfe, 2019.
30. Jared AO, Robert MG, Maurice EO, Paul CK, James OO, Miheso M. Morphological characterization of blackberry (*Rubus subgenus Rubus watson*) genetic resources in Kenya. *Afr. J. Plant Sci*. 2019;13(11):297-308, DOI: 10.5897/AJPS2018.1703
31. Jian W, Lu H, Wang RY, He MM, Liu QC. Genetic diversity and population structure of 288 potato (*Solanum tuberosum* L.) germplasms revealed by SSR and AFLP markers. *Journal of integrative agriculture*. 2017;16:2434-2443. doi: 10.1016/S2095-3119(16)61619-2
32. Juyó D, Sarmiento F, Álvarez M, Brochero H, Gebhardt C, Mosquera T. 'Genetic diversity and population structure in diploid potatoes of *Solanum tuberosum* group phureja', *Crop Science*. The Crop Science Society of America, Inc. 2015;55(2):760-769. doi: 10.2135/cropsci2014.07.0524.
33. Kolech SA, Halseth D, Perry K, Wolfe D, Douches DS, Coombs J, *et al.* Genetic Diversity and Relationship of Ethiopian Potato Varieties to Germplasm from North America, Europe and the International Potato Center. *J. Potato Res*, 2016. doi: 10.1007/s12230-016-9543-3
34. Louwaars NP. Plant breeding and diversity: A troubled relationship? *Euphytica*. 2018;214:114. <https://doi.org/10.1007/s10681-018-2192-5>
35. Mahgoub HAM, Eisa GSA, Youssef MAH. Molecular, biochemical and anatomical analysis of some potato (*Solanum tuberosum* L.) cultivars growing in Egypt. *Journal of Genetic Engineering and Biotechnology*. 2014;13:39-49.
36. Mandal R, Nag S, Tarafdar J, Mitra S. A comparison of efficiency parameters of SSR markers and genetic diversity analysis in *Amorphophallus paeoniifolius* (Dennst.) Nicolson. *Braz. arch. biol. technol*. 2016;59:e16160439. <http://dx.doi.org/10.1590/1678-4324-2016160439>.
37. Mishra S, Singh J, Sharma PK. Studies on Parameters of Genetic Variability for Yield and its Attributing Traits in Potato (*Solanum Tuberosum* L.). *Bio Sci. Biotech Res Asia*, 2017,14(1).
38. Nasiruddin M, Haydar FMA, Islam AKMR. Genetic diversity in potato (*Solanum tuberosum* L.) genotypes grown in Bangladesh. *Int. Res. J. Biological Sci*. 2017;6(11):1-8.
39. Nasiruddin MD, Ali, Fawzi, Islam AKM. Genetic diversity in potato (*Solanum tuberosum* L.) genotypes grown in Bangladesh. 2017;6:1-8.
40. National Potato Council. Potato facts. <http://www.nationalpotatocouncil.org/potato-facts/>, 2016.
41. National statistical year book, 2016 <https://www.nationalpotatocouncil.org/files/7014/6919/7938/NPCyearbook2016-FINAL.pdf>
42. Peter JR. Silhouettes: A graphical aid to the interpretation and validation of cluster analysis, *Journal of Computational and Applied Mathematics*. 1987;20:53-65.
43. Placide R, Shimelis H, Laing M, Gahakwa D. Application of principal component analysis to yield and yield related traits to identify sweet potato breeding parents. *Trop. Agric*. 2015;92:1-15
44. Prabha N, Nanda HC, Sharma SK. Genetic Divergence Analysis in Potato (*Solanum tuberosum* L.). *Int. J Curr. Microbiol. App. Sci*. 2018;7(2):3152-3157.
45. Reddy BJ, Mandal R, Chakroborty M, Hijam L, Dutta P. A Review on Potato (*Solanum tuberosum* L.) and its Genetic Diversity. *International Journal of Genetics*. 2018;10(2):360-364.
46. Salimi H, Bahar M, Mirolohi A, Talebi M. Assessment of the Genetic Diversity Among Potato Cultivars from Different Geographical Areas Using the Genomic and EST Microsatellites. *Iran J Biotechnol*. 2016;14(4):270-277.
47. Sharma SK, Bolser D, Boer JD, Sønderkær M, Amoros W, Carboni MF. Construction of reference chromosome-scale pseudo molecules for potato: integrating the potato genome with genetic and physical maps. *Journal of Chemical Physics*. 2013;3(11):2255-2256.
48. Singh BP, Rajesh KR. History of Potato and its Emerging Problems in India. ICAR-Central Potato Research Institute, Shimla-171001, HP, 2014.
49. Song Xiaoyan, Zhang Chunzhi, Li Ying, Feng Shuangshuang, Yang Qing, Huang Sanwen. SSR Analysis of Genetic Diversity among 192 Diploid Potato Cultivars, *Horticultural Plant Journal*, 2016. <http://dx.doi.org/doi:10.1016/j.hpj.2016.08.006>.
50. Struik PC, Wiersema SG. Seed potato technology. Wageningen Pers, Wageningen, The Netherlands, 1999.
51. Wang F, Li F, Wang J. Genetic Diversity of Chinese and CIP Potato (*Solanum tuberosum* L.) Germplasm Assessed by Amplified Fragment Length Polymorphism (AFLP) Markers. *Potato Research*. 2013;56:167-178.
52. Wang Y, Rashid MAR, Li X, Yao C, Lu L, Bai J, *et al.* Collection and Evaluation of Genetic Diversity and Population Structure of Potato Landraces and Varieties in China. *Front. Plant Sci*. 2019;10(139):1-11. doi: 10.3389/fpls.2019.00139
53. Wiesmann D, Biesalski HK, Grebmer KV, Bernstein J.

Methodological Review and Revision of the Global Hunger Index. ZEF Working Paper Series No. 139. Bonn: University of Bonn, Center for Development Research (ZEF), 2015.

54. Yang XS, Su WJ, Wang LJ, Lei J, Chai SS, Liu QC. Molecular diversity and genetic structure of 380 sweetpotato accessions as revealed by SSR markers. *Journal of Integrative Agriculture*. 2015;14:633-641. doi: 10.1016/s2095-3119(14)60794-2
55. Yuan J, Murphy A, Koeyer DD, Lague M, Bizimungu B. Effectiveness of the field selection parameters on potato yield in Atlantic Canada. *Canadian Journal of Plant Science*. 2016;96(4):701-710, 10.1139/cjps-2015-0267
56. Zheng X, Jiang H, Bi Y, Wang B, Wang T, Li Y *et al.* Comparison of wound healing abilities of four major cultivars of potato tubers in China. *Postharvest Biology and Technology*. 2020;164:111167. doi.org/10.1016/j.postharvbio.2020.111167