



ISSN (E): 2277-7695
ISSN (P): 2349-8242
NAAS Rating: 5.23
TPI 2022; SP-11(7): 3038-3039
© 2022 TPI
www.thepharmajournal.com
Received: 05-04-2022
Accepted: 09-06-2022

Basant

MVSc. Animal Biotechnology,
Department of Veterinary
Microbiology and Biotechnology,
College of Veterinary and Animal
Sciences, Rajasthan University
of Veterinary and Animal
Sciences, Bikaner, Rajasthan,
India

Parma Ram Gorachiya

Ph.D., Livestock Products
Technology, College of
Veterinary and Animal Science,
Rajasthan University of
Veterinary and Animal Sciences,
Bikaner, Rajasthan, India

Narendra Singh

Ph.D., Scholar, Department of
Veterinary Surgery and
Radiology, College of Veterinary
and Animal Science, Rajasthan
University of Veterinary and
Animal Sciences, Bikaner,
Rajasthan, India

Corresponding Author

Basant

MVSc. Animal Biotechnology,
Department of Veterinary
Microbiology and Biotechnology,
College of Veterinary and Animal
Sciences, Rajasthan University
of Veterinary and Animal
Sciences, Bikaner, Rajasthan,
India

Phylotyping of *Escherichia coli* Isolates from calf diarrhea

Basant, Parma Ram Gorachiya and Narendra Singh

Abstract

Calf diarrhea is one of the major health challenges in cattle herds. *Escherichia coli* is the most common cause of calf diarrhea. In the current study, we isolated a total of 27 isolates from diarrheic calves. The preliminary identification was carried out by the development of a green metallic sheen on the Eosin Methylene Blue agar plate. After that molecular level confirmation by amplification of 16S rRNA gene. The positive isolates were subjected to phylogroups. The B1 and A were most prevalent while phylogroup C, D, and E were less likely to be associated with calf diarrhea.

Keywords: Calf diarrhea, *E. coli*, Quadruplex PCR

Introduction

Newborn calves play an important role in animal production for meat production as well as breeding purposes worldwide. Calf diarrhea is a disease of neonatal and young animals and is well-known for productivity and economic loss to livestock owners throughout the world. The main causative agent for diarrhea includes enterotoxigenic *Escherichia coli* (*E. coli*), which causes noninflammatory secretory diarrhea to 3-5 days old calves mainly, *Salmonella* sp. causes foul-smelling feces containing blood and mucus which further leads to septicemia mainly to 2-12 weeks of calves, *Clostridium perfringens* which causes fatal hemorrhagic enteritis up to 8 weeks of age and another known causative agents are Bovine Rotavirus, Bovine Coronavirus and *Cryptosporidium parvum* (Cho *et al.*, 2014) [1].

E. coli consists of commensal and pathogenic strains of great diversity, therefore the classification of those microorganisms in phylogenetic groups through the combination of gene clusters is vital to understanding *E. coli* pathogenesis and interaction with its hosts (Croxen *et al.*, 2013) [2]. An understanding of the *E. coli* genomic structure showed that the strains belonging to the various phylogroups are also related to the source of isolation (Clermont *et al.*, 2013) [3]. Since its introduction in 2000 (Clermont *et al.*, 2000) [4], phylogenetic typing using PCR has become widely used because of its simplicity and speed, allowing the differentiation of virulent strains (B2 or D) and commensal lineages (A and B1). This method was improved in 2013 (Clermont *et al.*, 2013) [3] and a quadruplex PCR can identify seven phylogroups (A, B1, B2, C, D, E, and F). This study aimed to find an association of different phylogroups with calf diarrhea in the Bikaner district of Rajasthan.

Material and Methods

Sample collection and bacterial isolation: A total of 100 fecal samples were collected from diarrheic calves from private dairy farms near Bikaner, Rajasthan in the year 2018 and 2019. 1 gram of fecal sample was diluted in PBS followed by overnight incubated in with the nutrient broth. The enriched sample was streaked on MacConkey Agar Plate. The pink colonies (lactose fermentors) obtained from MacConkey Agar were subjected to further streaked on selective media EMB Agar. The characteristic Growth of colonies on EMB agar was a metallic sheen of colonies. The molecular levels of conformation of EMB positive isolates were carried out by performing 16sRNA-gene-specific PCR as described previously (Khalad *et al.*, 2010) [5] using forward primer 5'-GCTTGACACTGAACATT-3' and reverse primer 5'-GCACTTATCTCTTCCGCATT-3'.

Amplification of genes encoding for phylogenetic groups: After isolation of genomic DNA by using the phenol-chloroform method, the phylogroups of *E. Coli* isolates were carried out by performing Quadruplex-PCR as per described previously (Clermont *et al.* 2013) [3].

The PCR product was analyzed by agarose gel electrophoresis to examine the amplicon sizes and the phylogenetic group to which an isolate belonged was done as per the method of Clermont *et al.* (2013) [3].

Results and Discussion

Isolation and confirmation of *E. coli* isolate: Preliminary isolation and identification of *E. coli* isolates were carried out by culture characteristic (Differential media, Selective media) and gram staining. A total of 27 isolates were recovered in the study which showed pink colonies on MacConkey lactose Agar and also produces green metallic sheen on EMB agar which is characteristic of *E. coli*. Besides this, all 27 isolates were found positive for species-specific 16srRNA ribotyping (all isolates showed an amplicon of 662bp). The findings were similar to Meena *et al.*, 2015 [6] where 40 isolates of *E. coli* from healthy poultry were phenotypically confirmed by metallic sheen on EMB agar and all the isolates were genotypically confirmed by 16SrRNA gene.

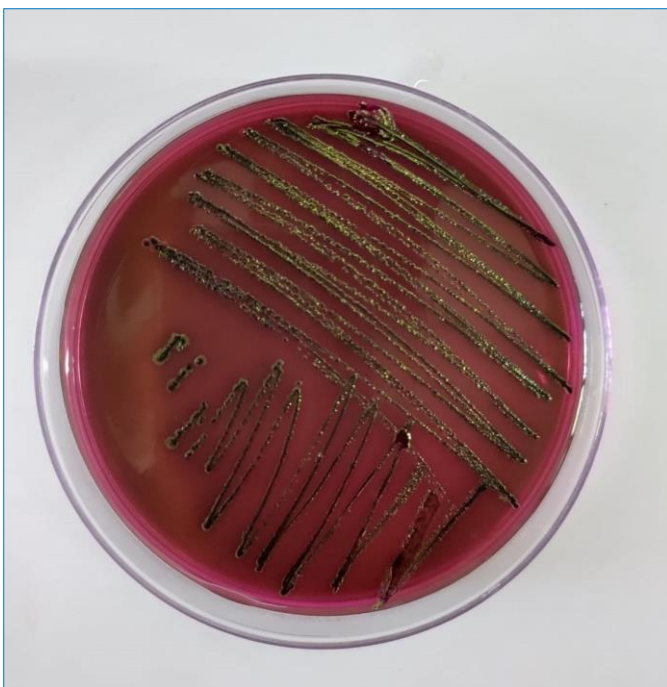


Fig 1: Metallic sheen colony of *E. coli* on EMB agar plate

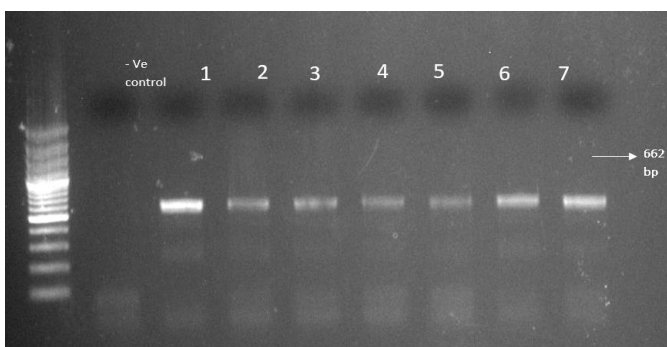


Fig 2: PCR based species-specific 16 S r RNA ribotyping of *E. coli* isolates

Quadruplex PCR-based Phylogenetic Grouping of *E. Coli*: The phylogrouping of *E. Coli* isolates was carried out by performing Quadruplex-PCR as per described previously described by Clermont *et al.*, 2013. The phylogenetic group analysis revealed that the most prevalent group B1 had 11

(40.75%) isolates followed by group A having 7 (25.92%) isolates. Only one isolate belongs to Phylogroup C, D, and E each. The 5 (18.51%) isolates were found to not be associated with any phylogenetic groups (Table-1). The findings were almost similar to Müştak *et al.*, 2015 [7], in which groups A1 and B2 were found the most prevalent by triplex and quadruplex PCR assays in *E. coli* isolates from mastitis samples and also most of the urinary tract isolates belonged to A1 by triplex and B2 by quadruplex PCR assays.

Table 1: Quadruplex PCR-based Phylogenetic Grouping of *E. Coli*

Phylogroup	Number of <i>E. Coli</i> isolates
Phylogroup-A	7
Phylogroup-B1	11
Phylogroup-C	2
Phylogroup-D	1
Phylogroup-E	1
Phylogroup-U	5

Conclusion

E. coli is the most common causative agent of calf diarrhea. Easy and early identification of pathogens plays an important role in the treatment of such diseases. In the present study, all the isolates from calf diarrhea samples showed a specific metallic sheen colony on EMB agar which is characteristic of *E. coli* isolates, and showed 16s r RNA specific gene amplification. In phylogenetic grouping by quadruplex highest isolates belonged to B1 followed by A, U, C, D, and, E.

Reference

1. Cho YI, Yoon KJ. An overview of calf diarrhea-infectious etiology, diagnosis, and intervention. *Journal of veterinary science.* 2014;15(1):1-17.
2. Croxen MA, Law RJ, Scholz R, Keeney KM, Wlodarska M, Finlay BB. Recent advances in understanding enteric pathogenic *Escherichia coli*. *Clinical microbiology reviews.* 2013;26(4):822-880.
3. Clermont O, Christenson JK, Denamur E, Gordon DM. The Clermont *Escherichia coli* phylo-typing method revisited: improvement of specificity and detection of new phylo-groups. *Environmental microbiology reports.* 2013;5(1):58-65.
4. Clermont O, Bonacorsi S, Bingen E. Rapid and simple determination of the *Escherichia coli* phylogenetic group. *Applied and environmental microbiology.* 2000;66(10):4555-4558.
5. Khalad J, Jeremy B, Pinyon L, Anantham S, Ruth M. Distribution of Human Commensal *Escherichia coli* Phylogenetic Groups. *Journal of Clinical Microbiology.* 2010;48(9):3455-3456.
6. Meena RH, Mir IA, Maherchandani S, Jangir K, Purohit N, Kashyap SK. Antibiotic Resistance Pattern and Phylogenetic Analysis of Commensal *Escherichia coli* Isolated from Poultry. *Journal of Pure and Applied Microbiology.* 2015;9(1):657-662.
7. Müştak HK, Günaydin E, Kaya İB, Salar MÖ, Babacan O, Önat K, *et al.* Phylo-typing of clinical *Escherichia coli* isolates originating from bovine mastitis and canine pyometra and urinary tract infection by means of quadruplex PCR. *Veterinary Quarterly.* 2015;35(4):194-199.