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Effect of pre-sowing treatments on seed germination and vigour of seedlings of commercial acid lime cultivars of Andhra Pradesh

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Abstract

The present experiment was conducted at commercial nursery block, College of Horticulture, Anantharajupeta, Dr. Y.S.R. Horticultural University, Andhra Pradesh during the year 2022. The experiment was laid out in randomized block design with two factors and three replications. The first factor comprises of 2 levels of cultivars (Balaji and Petluru Pulusu Nimma) and the second factor comprises of 10 levels of different seed treatments viz. F₁ (GA₃ @ 250 ppm), F₂ (GA₃ @ 500 ppm), F₃ (NAA @ 25 ppm), F₄ (NAA @ 50 ppm), F₅ (KNO₃ @ 1%), F₆ (KNO₃ @ 2%), F₇ (KNO₃ @ 3%), F₈ (GA₃ @ 250 ppm + KNO₃ @ 1%), F₉ (NAA @ 25 ppm + KNO₃ @ 1%). F₁₀ (control) with a total of 20 treatment combinations. Result revealed that the acid lime seeds soaked in F₈ (GA₃@ 250 ppm + KNO₃ @ 1%) found minimum days for initiation of germination (12.26), minimum days for 50% germination (19.69), maximum germination percentage (78.34), maximum germination index (5.3), maximum survival percentage of germinated seedlings (95.97).

Keywords: GA₃, KNO₃, germination, growth regulators, seed treatment, Balaji, Petluru Pulusu Nimma

Introduction

Citrus fruits have a prominent place among popular and extensively grown tropical and subtropical fruits. Among them, acid lime (*Citrus aurantifolia* Swingle) is an important citrus crop which is grown on commercial scale in India and Andhra Pradesh. It is a member of the family Rutaceae and originated in the tropical and sub tropical regions of South East Asia, particularly India and China. In India, Acid lime covers an area of 333 thousand hectares with a production of 3750 thousand MT.

In India, Andhra Pradesh ranks first in production and it covers 30 thousand hectares with a production of 731 thousand MT. It is also known as Pati lime, Kagzi lime and Mexican lime etc. Acid lime is used for table purpose in daily life of Indians for flavouring vegetable curries, salad, fish and meat. It is also used in the preparation of refreshing cold drinks especially to beat the hot summer. Acid lime contains 6.3-6.6% citric acid. It is an appetizer, anti scorbutic, anti helminthic and it checks biliousness besides a good source of nutrients, vitamins and other antioxidant compounds. It is used in making candies, chocolates, ice-creams and pastries etc.

Acid lime is commercially propagated by seeds as it is the easiest and cheapest method of propagation. Acid lime propagation is hampered by high mortality at the nursery stage, moreover; seeds lose their viability very soon. In acid lime germination percentage is low and it varies between 27-58% and it takes about 3 weeks to germinate (Cheema *et al.*, 1954)^[10]. Nurserymen and growers thus face problems like lower seed germination and poor vigour of seedlings. The growth of acid lime seedlings is very slow and therefore, to raise seedlings with in the shortest possible time, growth has to be accelerated for which pre sowing treatments can be employed. The use of growth regulators in overcoming the inhibitory action of certain chemical substances that delay the germination has been reported by Abohassan *et al.* (1979)^[1] in kagzi lime and Chaudhari and Chakrawar (1980)^[12] in Rangpur lime. Nitrate ions stimulate germination of dormant seeds (Alboresi *et al.*, 2005)^[2]. Keeping this in view, an experiment entitled "Effect of pre-sowing treatments on seed germination and vigour of seedlings of commercial acid lime cultivars of Andhra Pradesh.

Material and Methods

The present experiment was carried out at commercial nursery block, College of Horticulture, Anantharajupeta, YSR Kadapa district, Andhra Pradesh during the year 2022 to study the effect of pre-sowing treatments on seed germination and vigour of seedlings of commercial acid lime cultivars of Andhra Pradesh which falls under southern agro-climatic zone of Andhra Pradesh at an elevation of 162 m (531 feet) above mean sea level; geographically it lies between 130 59' North latitude and 790 19' East longitude. The experiment was laid out in factorial randomized block design for germination parameters. The treatments comprised of different seed treatments viz. F₁ (GA₃ @ 250 ppm), F₂ (GA₃ @ 500 ppm), F₃ (NAA @ 25 ppm), F₄ (NAA @ 50 ppm), F₅ (KNO₃ @ 1%), F₆ (KNO₃ @ 2%), F₇ (KNO₃ @ 3%), F₈ (GA₃ @ 250 ppm + KNO₃ @ 1%) F₉ (NAA @ 25 ppm + KNO₃ @ 1%), F₁₀ (control). were soaked for 24 hr and with a total of 20 treatment combinations are employed in this seed treatments, the seeds were collected from disease free and fully ripened fruits of acid lime and uniform size, healthy seeds were selected and used for the experimental purpose. Seeds were sown during the last week of January in nursery beds, immediately watered by using rose cane and also maintained optimum moisture level when required. The observation on days taken for initiation of germination, Days to 50% germination, germination percentage, germination index, survival percentage, The data recorded from the present studies were subjected to analysis by using standard method suggested by Panse and Sukhatme (1967)^[22].

Results

Germination parameters

Days for initiation of seed germination

The information regarding the effect of seed treatments with growth regulators and chemicals on the days for initiation of seed germination presented in table 1 and figure 1.

Among the two cultivars significantly minimum days for initiation of seed germination was observed in C₂ (Petluru Pulusu Nimma) cultivar (14.76 days), than that of Balaji (18.91) cultivar.

The recorded data clearly indicated that the days for initiation of seed germination was significantly influenced by the seed treatment with plant growth regulators and various chemicals. Minimum number of days taken for seed germination was recorded when the seeds were treated with F₈ (GA₃ @ 250 ppm + KNO₃ @ 1%) (12.26 days), while maximum number of days for germination was recorded in F₁₀ (control) (20.88 days).

Among the interaction effect of cultivars and seed treatments was recorded significantly minimum number of days when the seeds were treated with (C₂F₈) (Petluru Pulusu Nimma treated with GA₃ @ 250 ppm + KNO₃ @ 1%) (11.03 days), which was on par with the (C₂F₉) (Petluru Pulusu Nimma treated with NAA @ 25 ppm + KNO₃ @ 1%) (11.67), while the maximum number of days for germination was recorded in (C₁F₁₀) (Balaji with control) (23.47 days).

Days for 50% seed germination

The information regarding the effect of seed treatments with growth regulators and chemicals on the days for 50% seed germination was presented in table 2.

Among the two cultivars, significantly minimum days for 50% seed germination was observed in C₂ (Petluru Pulusu

Nimma) cultivar (23.09 days), than that of Balaji cultivar (26.84).

The recorded data clearly indicated that the days for 50% germination was significantly influenced by the seed treatment with plant growth regulators and various chemicals. Minimum number of days taken for 50% germination was recorded when the seeds were treated with F₈ (GA₃ @ 250 ppm + KNO₃ @ 1%) (19.69 days), while maximum number of days for 50% germination was recorded in F₁₀ (control) (29.78 days).

Among the interaction effect of cultivars and seed treatment was observed significantly for 50% germination with minimum number of days when the seeds were treated with (C₂F₈) (Petluru Pulusu Nimma treated with GA₃ @ 250 ppm + KNO₃ @ 1%) (18.74 days), which was on par with the (C₂F₉) (Petluru Pulusu Nimma with NAA @ 25 ppm + KNO₃ @ 1%) (19.71), while maximum number of days for 50% germination was recorded in (C₁F₁₀) (Balaji with control) (30.41 days).

Germination percentage

The information regarding the effect of seed treatments with growth regulators and chemicals on the germination percentage was presented in table 3 and figure 2.

Between the two cultivars, significantly highest germination percentage was observed in C₂ (Petluru Pulusu Nimma) cultivar (78.69%), than that of Balaji cultivar (69.18).

The recorded data clearly indicates that the germination percentage was significantly influenced by the seed treatment with plant growth regulators and various chemicals. Significantly highest germination percentage was observed when the seeds were treated with F₈ (GA₃ @ 250 ppm + KNO₃ @ 1%) (78.34%) while, minimum germination percentage was observed in F₁₀ (control) (67.42%).

Among the interaction effect of cultivars and seed treatment significantly maximum germination percentage was observed when the seeds were treated with (C₂F₈) (Petluru Pulusu Nimma treated with GA₃ @ 250 ppm + KNO₃ @ 1%) (83.14%), which was on par with the (C₂F₉) (Petluru Pulusu Nimma treated with NAA 25 ppm + KNO₃ @ 1%) (81.70) while, minimum seed germination percentage was observed in (C₁F₁₀) (Balaji with control) (64.57%).

Seed germination index

The information regarding the effect of seed treatments with growth regulators and chemicals on the seed germination index was presented in table 4.

Between the two cultivars, significantly highest seed germination index was observed in C₂ (Petluru Pulusu Nimma) cultivar (6.08), than that of Balaji cultivar (2.93).

The recorded data clearly indicated that the germination index was significantly influenced by the seed treatment with plant growth regulators and various chemicals. Significantly highest germination index was observed when the seeds were treated with F₈ (GA₃ @ 250 ppm + KNO₃ @ 1%) (5.30), which was on par with the F₉ (NAA @ 25 ppm + KNO₃ @ 1%) (5.18), while minimum germination index was observed in F₁₀ (control) (3.10).

Among the interaction effect of cultivars and seed treatments significantly affected the seed germination index. Maximum germination index was observed with (7.40) when the seeds were treated with (C₂F₈) (Petluru Pulusu Nimma treated with GA₃ @ 250 ppm + KNO₃ @ 1%) which was on par with the (C₂F₉) (Petluru Pulusu Nimma treated with NAA 25 ppm with

KNO₃ @ 1%) (7.26).

Final survival percentage of seedlings

The information regarding the effect of seed treatments with growth regulators and chemicals on the survival percentage of seedlings was presented in Table 5 and Figure 3.

Between the two cultivars, significantly highest survival percentage was observed in C₂ (Petluru Pulusu Nimma) cultivar (91.35), than that of Balaji.

The data recorded clearly indicates that the survival percentage was significantly influenced by the seed treatment with plant growth regulators and various chemicals. Significant highest survival percentage was observed when the seeds were treated with F₈ (GA₃ @ 250 ppm + KNO₃ @ 1%) (94.47), which were on par with the F₉ (NAA @ 25 ppm with KNO₃ @ 1%) (93.24), F₂ (GA₃ @ 500 ppm) (93.18), F₁ (GA₃ @ 250 ppm) (92.61), while minimum survival percentage was observed with control (85.13).

The interaction effect of cultivars and seed treatment significantly affected the highest survival percentage. The maximum survival percentage was observed when the seeds were treated with (C₂F₈) (Petluru Pulusu Nimma treated with GA₃ @ 250 ppm with KNO₃ @ 1%) (95.29), which were on par with the (C₂F₉) (NAA @ 25 ppm with KNO₃ 1%) (94.23), (C₂F₁) (Petluru Pulusu Nimma treated with GA₃ @ 250 ppm) (93.25), (C₂F₅) (Petluru Pulusu Nimma treated with Arka Citrus Special @ 2%) (94.31), (C₂F₂) (Petluru Pulusu Nimma treated with GA₃ @ 500 ppm) (94.23) while, minimum survival percentage was observed when seeds were treated with (C₁F₁₀) Balaji with control (82.60).

Discussion

Early germination in seeds treated with GA₃ may be attributable to the compound's direct action in releasing the embryo from its state of dormancy by encouraging protein synthesis, elongating the coleoptile, primary leaves, and aiding in the formation of ethylene. GA₃ stimulates the manufacture of hydrolases, particularly amylase enzyme, which promotes early seed germination. Several studies also confirmed the GA₃ treated seeds shown same results. Kumari *et al.*, 2007^[19] in Aonla, Garge *et al.*, 2011^[15] in custard apple, Athani *et al.*, 2013^[4] in guava and Lalitha *et al.*, 2020^[20] in aonla, Anusha 2009^[3] in avocado, chaudary 2020^[9] in Kagzi lime, Kadam *et al* 2010^[18] in kagzi lime, Joshi *et al.* (2015)^[17] in acid lime, Chiranjeevi *et al.* (2017)^[11] in Aonla, Dilip *et al.* (2017)^[13] in rangapur lime and Panda *et al.* (2018)^[21] in Kagzi lime.

KNO₃ promotes germination by lowering the endogenous abscisic acid levels in embryos due to increased oxidation of NADPH to NADP and enzymes that have been allocated to provide nutrients for germination, resulting in cell shrinkage and weakening of the aleurone layer's cell wall (Bethke *et al.* 2007, participation in the pentose phosphate pathway)^[7]. (Finkelstein *et al.* 2008)^[15], the buildup of nitrogen and potassium ions in seeds may also contribute to the rise in germination. (Hegazi *et al.* 2011 and Banik *et al.* 2015)^[16, 5] increasing the production of amino acids, which may have helped in enhancing germination and growth in plant tissues. promote germination among different species and same results observed in Bindu *et al.* (2014)^[8] in mango, Patel *et al.* 2018^[23] in jamun. Barathkumar^[6], 2019 in Aonla and Sheoran *et al.* 2019^[24] in ber.

Table 1: Days for initiation of seed germination as influenced by pre sowing treatments of growth regulators and chemicals in two commercial cultivars of acid lime.

Pre-sowing treatments	Days for initiation of germination		
	Cultivars		
	C ₁ (Balaji)	C ₂ (Petluru Pulusu Nimma)	Mean
F ₁ (GA ₃ @ 250 ppm)	17.74	13.62	15.68
F ₂ (GA ₃ @ 500 ppm)	15.84	13.41	14.62
F ₃ (NAA@ 25 ppm)	21.37	16.28	18.82
F ₄ (NAA@ 50 ppm)	22.48	17.51	19.99
F ₅ (KNO ₃ @ 1%)	19.69	14.64	17.17
F ₆ (KNO ₃ @ 2%)	20.89	16.25	18.57
F ₇ (KNO ₃ @ 3%)	19.57	15.10	17.31
F (GA ₃ @ 250 pp+KNO ₃ @1%)	13.49	11.03	12.26
F ₉ (NAA@ 25 pp+KNO ₃ @1%)	14.62	11.67	13.14
F ₁₀ (Control)	23.47	18.29	20.88
Mean	18.91	14.76	
Factors	C.D (5%)		S.E(m)±
C	0.24		0.08
F	0.53		0.18
C X F	0.75		0.26

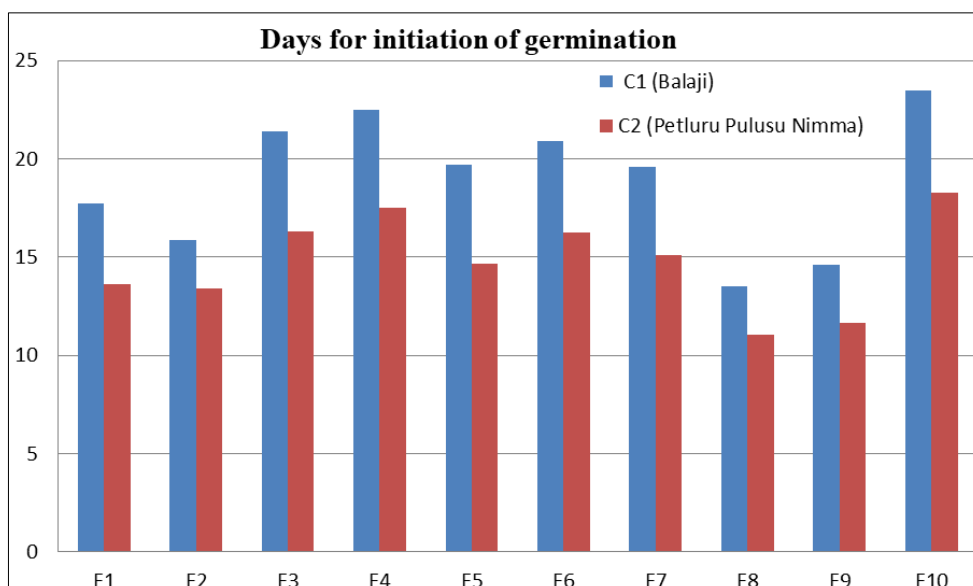


Fig 1: Days for initiation of germination

Table 2: Days for 50 percentage of seed germination as influenced by pre sowing treatments of growth regulators and chemicals in two commercial cultivars of acid lime.

Pre-sowing treatments	Days for 50 seed percentage		
	Cultivars		
	C ₁ (Balaji)	C ₂ (Petluru Pulusu Nimma)	Mean
F ₁ (GA ₃ @ 250 ppm)	25.84	22.19	24.02
F ₂ (GA ₃ @ 500 ppm)	24.47	21.31	22.89
F ₃ (NAA@ 25 ppm)	29.63	24.64	27.12
F ₄ (NAA@ 50 ppm)	29.74	25.46	27.60
F ₅ (KNO ₃ @ 1%)	27.71	22.47	25.09
F ₆ (KNO ₃ @ 2%)	29.43	24.14	26.78
F ₇ (KNO ₃ @ 3%)	27.97	23.08	25.52
F (GA ₃ @ 250 ppm +KNO ₃ @1%)	20.64	18.74	19.69
F ₉ (NAA@ 25 ppm +KNO ₃ @1%)	22.57	19.71	21.14
F ₁₀ (Control)	30.41	29.17	29.78
Mean	26.84	23.09	
Factors	C.D (5%)		S.E(m) ±
C	0.31		0.11
F	0.71		0.24
C X F	0.99		0.34

Table 3: Seed germination percentage as influenced by pre sowing treatments of growth regulators and chemicals in two commercial cultivars of acid lime

Pre-sowing treatments	Seed germination percentage		
	Cultivars		
	C ₁ (Balaji)	C ₂ (Petluru Pulusu Nimma)	Mean
F ₁ (GA ₃ @ 250 ppm)	71.63	76.58	75.10
F ₂ (GA ₃ @ 500 ppm)	70.58	79.95	75.27
F ₃ (NAA@ 25 ppm)	67.86	77.61	72.73
F ₄ (NAA@ 50 ppm)	65.92	76.50	71.13
F ₅ (KNO ₃ @ 1%)	68.55	80.03	74.29
F ₆ (KNO ₃ @ 2%)	66.14	79.07	72.61
F ₇ (KNO ₃ @ 3%)	66.43	80.06	74.25
F (GA ₃ @ 250 ppm +KNO ₃ @1%)	73.55	83.14	78.34
F ₉ (NAA@ 25 ppm +KNO ₃ @1%)	72.62	81.70	71.16
F ₁₀ (Control)	64.57	70.26	67.42
Mean	69.18	78.69	
Factors	C.D (5%)		S.E(m) ±
C	0.181		0.063
F	0.405		0.141
C X F	0.573		0.199

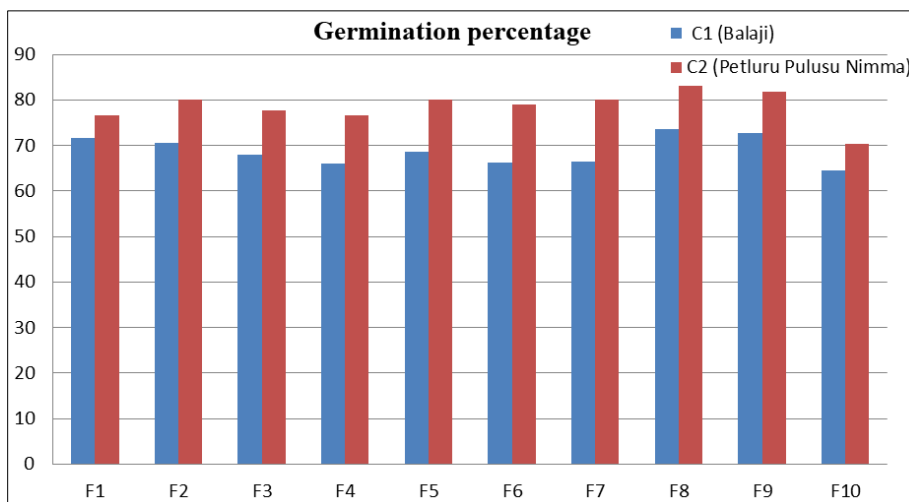


Fig 2: Germination percentage

Table 4: Seed germination percentage as influenced by pre sowing treatments of growth regulators and chemicals in two commercial cultivars of acid lime

Pre-sowing treatments	Germination index		
	Cultivars		
	C ₁ (Balaji)	C ₂ (Petluru Pulusu Nimma)	Mean
F ₁ (GA ₃ @ 250 ppm)	3.16	5.60	4.38
F ₂ (GA ₃ @ 500 ppm)	3.26	6.50	4.88
F ₃ (NAA@ 25 ppm)	2.80	5.30	4.05
F ₄ (NAA@ 50 ppm)	2.70	6.42	4.56
F ₅ (KNO ₃ @ 1%)	3.00	5.00	4.30
F ₆ (KNO ₃ @ 2%)	2.70	6.30	4.47
F ₇ (KNO ₃ @ 3%)	2.80	6.90	4.85
F (GA ₃ @ 250 ppm +KNO ₃ @1%)	3.20	7.40	5.30
F ₉ (NAA@ 25 ppm +KNO ₃ @1%)	3.11	7.26	5.18
F ₁₀ (Control)	2.60	3.60	3.10
Mean	2.93	6.08	
Factors	C.D (5%)		SE(m) ±
C	0.03		0.01
F	0.08		0.02
C X F	0.12		0.04

Table 5: Survival percentage as influenced by pre sowing treatments of growth regulators and chemicals in two commercial cultivars of acid lime

Pre-sowing treatments	Survival percentage		
	Cultivars		
	C ₁ (Balaji)	C ₂ (Petluru Pulusu Nimma)	Mean
F ₁ (GA ₃ @ 250 ppm)	91.97	93.25	92.61
F ₂ (GA ₃ @ 500 ppm)	92.14	94.23	93.18
F ₃ (NAA@ 25 ppm)	89.57	90.55	90.06
F ₄ (NAA@ 50 ppm)	86.52	86.57	91.15
F ₅ (KNO ₃ @ 1%)	87.92	94.31	89.81
F ₆ (KNO ₃ @ 2%)	89.40	90.38	89.89
F ₇ (KNO ₃ @ 3%)	90.06	92.18	91.12
F (GA ₃ @ 250 ppm +KNO ₃ @1%)	93.65	95.29	94.47
F ₉ (NAA@ 25 ppm +KNO ₃ @1%)	92.26	94.23	93.24
F ₁₀ (Control)	82.60	84.23	85.13
Mean	89.62	91.35	
Factors	C.D (5%)		S.E(m)±
C	1.24		0.43
F	2.78		0.97
C X F	3.94		0.37

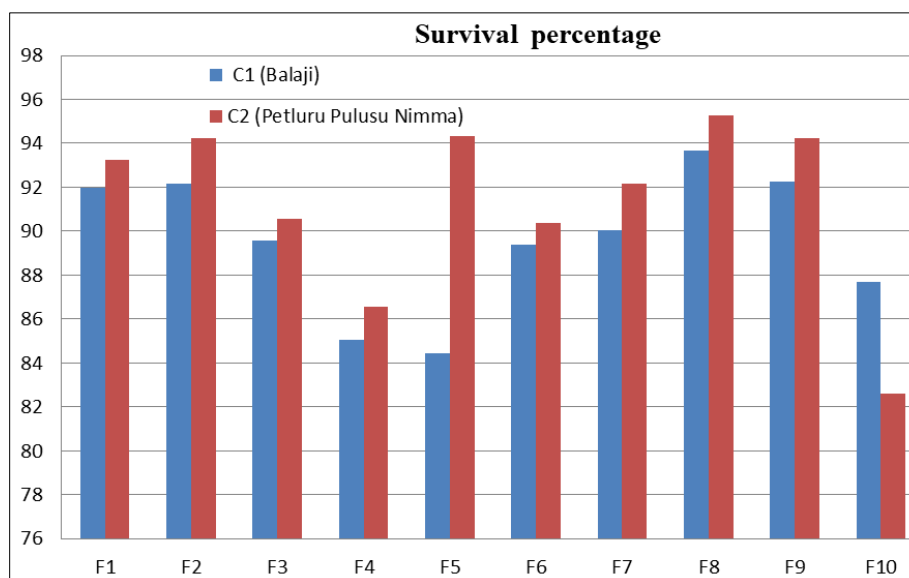


Fig 3: Survival percentage

Conclusion

By the present experimental results, it can be concluded that seed soaking with GA₃ @ 250 ppm + KNO₃ 1% has showed best germination attributes of acidlime cultivars, Between two cultivars Petluru Pulusu Nimma cultivar has recorded the best performance.

References

1. Abohassan AA, Hamady AAM, Hamouda MA. Effect of GA and kinetin on germination of apricot and lime seeds and subsequent seedling growth. Proceedings of Saudi Biological. Society, 1978;3:1-6.
2. Alboresi A, Gestin C, Leydecker MT, Bedu M, Meyer C. Trong HN. Nitrate, a signal revealing seed dormancy in Arabidopsis. Plant Cell Environment. 2005;28(4):500-512.
3. Anusha MD, Swamy GSK, Honnabyraiah MK, Vasudeva KR, Venkat Rao, Pallavi HM. Effect of growth regulators and chemicals on germination of avocado seeds. International Journal of Chemical Studies. 2019;7(4):3204-07.
4. Athani SI, Allolli TB, Gopali JB, Kotikal YK. Studies on effect of seed treatment on germination of guava (*Psidium guajava* L.) seeds. Abstract Book of National Seminar on Tropical and Subtropical Fruits, held at Navsari. 2013;36:9-11.
5. Banik BC, Dey AN, Pradhan S, Thapa N, Bhowmick N. Seed germination of karonda (*Carissa carandas* L.). Acta Horticulture. 2015;1074:23-26.
6. Barathkumar TR. Studies on influence of different seed treatments on dormancy breaking in aonla (*Phyllanthus embolic* L.). Journal of Pharmacognosy and Phytochemistry. 2019;2:131-33.
7. Bethke PC, Libourel IGL, Aoyama N, Chung Y, Still DW, Jones RL. The Arabidopsis aleuronic layer responds to nitric oxide, gibberellins, and abscisic acid and is sufficient and necessary for seed dormancy. Plant Physiology. 2007;143(3):1173-88.
8. Bindu A, Srihari D. Influence of pre-sowing treatments on germination, growth and vigour of mango cv. Alphonso. Asian Journal of Horticulture. 2013;8(1):122-25.
9. Chaudhary A, Ahlawat TR, Kumar S, Patel D, Jena S. Promoting seedling growth in kagzi lime through pre-sowing treatments. International Journal of Chemical Studies. 2020;8(1):2815-19.
10. Cheema GS, Bhat SS, Naik KC. Commercial fruits of India. Macmillan and cooperative limited; c1954, p. 153-156.
11. Chiranjeevi MR, Muralidhara BM, Sneha MK, Shivan, Hongal. The influence of growth regulators, chemicals and biofertilizers on germination, seedling growth and vigour attributes of Aonla. International Journal of Current Microbiology and Applied Sciences. 2017;6(12):1320-26.
12. Choudhari BK, Chakrawar VR. Effect of some chemicals on the germination of Kagzi lime (*Citrus aurantifolia Swingle*) seeds. Journal of Maharashtra agriculture luniver sities. 1980;5:173-174.
13. Dilip WS, Singh D, Moharana D, Rout S, Patra SS. Effect of gibberellic acid (GA) different concentrations at different time intervals on seed germination and seedling growth of rangpur lime. Journal of Agroecology and Natural Resource Management. 2017;4(2):157-162.
14. Finkelstein R, Reeves W, Ariizumi T, Steber C. Molecular aspects of seed dormancy. Annual Review of Plant Biology. 2008;59:387-15.
15. Garge VR, Kadam AS, Patil VK, Lakade SK, Dhomane PA. Effect of various concentrations of GA₃ and soaking period on seed germination of custard apple (*Annona squamosa* L.). Green Farming. 2011;2(5):550-51.
16. Hegazi ES, Mohamed SM, El-Sonbaty MR, Abd El-Naby SKM, El-Sharony TF. Effect of potassium nitrate on vegetative growth, nutritional status, yield and fruit quality of olive cv. Picual. Journal of Horticultural Science and Ornamental Plants. 2011;3(3):252-58.
17. Joshi PS, Sahoo AK, Bhojar RK, Meshram PC. Effect of various plant growth promoting substances on seedling growth of acid lime. Trends in Biosciences. 2015;8(19):5222-25.
18. Kadam AB, Singh DB. Effect of plant growth regulators and potassium nitrate on growth of seedlings of kagzi lime. The Asian Journal of Horticulture. 2010;5(2):431-34.

19. Kumari R, Sindhu SS, Sehwat SK, Dudi OP. Germination studies in Aonla (*Emblica officinalis* Gaertn.). Haryana Journal of Horticultural Sciences. 2007;36(1-2):9-11.
20. Lalitha KR, Tank RV, Chawla SL, Jena S. Effect of chemicals on seed germination and seedling growth of Aonla (*Emblica officinalis* Gaertn.). The Pharma Innovation Journal. 2020;9(12):239-43.
21. Panda PA, Karna AK, Sinha K. Effect of gibberellic acid (GA3) on germination, growth and survival of kagzi lime. International Journal of Chemical Studies. 2018;6(5):2803-05.
22. Panse VG, Sukhatme PV. Statistical Methods for Agricultural Workers, 1967.
23. Patel H, Tank RV, Bennurmath P, Patel M. Effect of seed treatment on germination and seedling growth of jamun (*Syzygium cumini* L.). International Journal of Current Microbiology and Applied Sciences. 2018;7(2):2654-59.
24. Sheoran V, Kumar M, Sharma JR, Gaur RK, Saini H. Effect of scarification treatments on growth parameters of ber seedling. Journal of Pharmacognosy and Phytochemistry. 2019;8(1):658-661.