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Evaluation of anthelmintic efficacy against gastrointestinal nematodosis in Cervids of Dehradun Zoo

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Abstract

Maintenance of wildlife population is an inevitable task. Endoparasitic diseases are one of the major problems in wildlife leading to morbidity and mortality. Indiscriminate use of anthelmintics for deworming of animals against gastrointestinal parasites has lead to development of anthelmintic resistance. Therefore, prompt decision on usage of right drug based on its efficacy is essential. The study to compare the efficacies of drugs namely Ivermectin and Fenbendazole was conducted in the Cervids of Dehradun Zoo. Animals were divided into an untreated, control group (I) and two treatment groups i.e. ivermectin (II) and fenbendazole (III). As per the routine Zoo regime, ivermectin was administered @0.2mg/kg body weight, orally and fenbendazole @7.5mg/kg body weight, orally. EPG (egg per gram) of collected faecal samples was determined on 0, 7th, 14th and 21st day post treatment, using the modified McMaster method to quantify nematode egg burdens. Faecal egg count reduction test (FECRT) showed 97 per cent efficacy for ivermectin, constantly reducing egg shedding by more than 95 per cent. Fenbendazole was also found 96.15% effective against nematodes, 14th day post-treatment. A greater than 95% reduction in faecal egg counts, in both the anthelmintic groups indicated that anthelmintics used are still beneficial. There was no significant (p>0.05) difference evident in efficacy of ivermectin and fenbendazole. Therefore, both the anthelmintics can be used in a strategic control program against gastrointestinal nematodosis in Cervids.

Keywords: Anthelmintic efficacy, fecal egg count, fenbendazole, ivermectin, Cervids

1. Introduction

Deer is a ruminant belonging to the family Cervidae and is included in the order Artiodactyla. The highest concentration of large deer species occurs in Southeast Asian countries like India and Nepal. Environmental changes and ecological disturbances, due to both natural phenomena and human intervention act like indiscriminate hunting and habitat destruction have contributed to the decline of this valuable creature of the country (Kanungo et al., 2011) [17]. Gastrointestinal (GI) nematodes have been shown to be a cause of morbidity and mortality in captive wild ungulates (Goossens et al., 2006) [12]. A major health issue in captive and wild deer is infection with helminthes (Thawait and Maiti, 2015) [27]. Abomasal damage comprised of varying degrees of hyperaemia, oedema, pitting and thickening of the abomasal mucosa. Banerjee et al. (2005) [2] in their study reported 17.7% samples of spotted deer, as positive for either single or mixed parasitic infection and Niranjan et al. (2022) [24] reported about 27.77% infection amongst captive herbivores in Uttarakhand. Strategic use of effective anthelmintics has been essential for the control of GI parasites in deers. Benzimidazoles are the most commonly used anthelmintics in captive wild ungulates owing to their wide margin of safety (Isaza et al., 1995) [14] and can be administered orally using different carriers such as feed or mineral licks. In the early 1980s ivermectin, a member of a new family of anthelmintics, the macrocyclic lactones (ML) or "mectins", was introduced and its efficacy in deers was investigated. The need for efficacy of anthelmintics arises from Agricultural Compounds and Veterinary Medicines (ACVM) Act, which provides for prevention or management of risks associated with the use of agricultural compounds including veterinary medicine (Geary et al., 2012)^[10]. Anthelmintic efficacies against helminthes in captive deer have been reported by various workers from time to time in different zoos of the country (Mason, 1994; Islam et al., 2003; Singh et al., 2006 and Castillo-Alcala et al., 2007) [20, 15, 26, 3]. But such information is very limited in Cervids of Uttarakhand.

Considering these facts, the present study was undertaken to evaluate the therapeutic efficacy of in-feed administration of fenbendazole and ivermectin against gastrointestinal nematode infections in members of Cervidae family.

2. Materials and Methods

2.1 Study area and animals

After getting approval from Chief Wildlife Warden, Uttarakhand and Institutional Animal Ethical Committee, the investigations were made in the Dehradun Zoo. The Zoo is located at Dehradun Mussoorie Highway. It is a part of reserve forest of Malsi Range and is dominated by coppice crop of Sal along with other species like Haldu, Rohini, Jhingan etc. The total area under this Zoo is 25 Ha (62.5 Acre). Spotted deer, Sambar, Barking deer belonging to family Cervidae, having egg counts more than 150epg were selected. Animals were randomly distributed into an untreated, control group (I) and two treatment groups i.e. ivermectin (II) and fenbendazole (III).

2.2 Treatment

As a part of the routine three monthly deworming strategies, animals were treated with anthelmintics, according to the manufacturer's recommended dose. Ivermectin (Bolus Neomac, Intas Pharmaceuticals Ltd., Ahmedabad, India) was administered at the dose rate of 0.2mg/kg body weight orally, mixed with concentrate feed and fenbendazole (Bolus Panacur 1.5VET, Intervet India Pvt. Ltd., Thane,

Maharashtra) at the dose rate of 7.5 mg/ kg body weight orally, mixed with concentrate feed.

2.3 Study design

The *in vivo* efficacy of the drugs was assessed by FECR tests. Fresh faecal samples were collected at 0, 7, 14, 21 days after treatment. About 5-10g of fresh faecal samples were collected randomly from the holding area and transported to the laboratory in separate sterile sealed containers. The faecal egg counts (FECs) were determined within 24 hours by a modified McMaster method, using a saturated solution of sodium chloride. The FECR was calculated using the formula by Young et al. (2000) [30]: {([pretreatment mean EPG] – [post-treatment mean EPG])/ (pretreatment mean EPG)} × 100 = % efficacy. For comparison, the FECR was also calculated as recommended by the World Association for the Advancement of Veterinary Parasitology (WAAVP) guidelines (Coles et al., 2006) [5], using the formula FECR% = $100\times(1-X_t/X_c)$, where X_t is the arithmetic mean EPG of the treated group and Xc is the arithmetic mean EPG of the control group.

2.4 Statistical analysis

Statistical analysis was made using Student's t test for comparing mean EPGs reduction between II (Ivermectin) and III (Fenbendazole) groups.

3. Results

Table 1: Average EPG, Geometric Mean, Median, Range, Variance of counts, Standard Deviation of Control (I Group)

Days	Arithmetic Mean/Average EPG*	Geometric Mean	Median	Range	Variance	Standard deviation
0	500	492.30	500	350-650	8333.34	91.29
7	505	498.92	500	400-650	6916.67	83.17
14	510	502.63	525	350-600	7666.67	87.56
21	515	512.10	500	450-600	3361.12	57.97

^{*}Not significant at 5% level of probability (p>0.05)

Table 2: Efficacy of Ivermectin (II Group) against Gastrointestinal Nematodosis in Cervids of Dehradun Zoo

Days	Arithmetic mean / Average EPG*	Geometric Mean	Median	Range	FEC Reduction (%)	Drug efficacy (%)	Variance	Standard deviation
0	500	484.27	500	300-700	-	-	16666.67	129.10
7	25	0	0	0-100	95.04	95	1250.00	35.36
14	15	0	0	0-50	97.05	97	583.34	24.15
21	20	0	0	0-50	96.12	96	666.66	25.81

^{*}Significant at 1% level of probability (p<0.01)

Table 3: Efficacy of Fenbendazole (III group) against Gastrointestinal Nematodosis in Cervids of Dehradun Zoo

Days	Arithmetic mean / Average EPG*	Geometric Mean	Median	Range	FEC Reduction (%)	Drug efficacy (%)	Variance	Standard deviation
0	520	512.67	525	350-650	1	1	7888.89	88.82
7	25	0	25	0-50	95.05	95.19	694.45	26.35
14	20	0	0	0-50	96.08	96.15	666.67	25.82
21	30	0	25	0-100	94.17	94.23	1222.22	34.96

^{*}Significant at 1% level of probability (*p*<0.01)

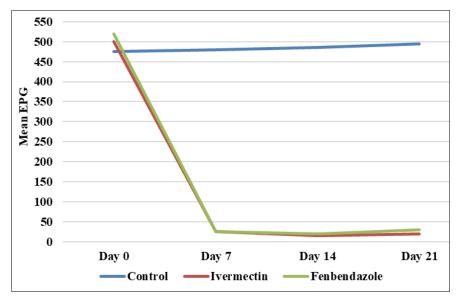


Fig 1: Comparative mean EPGs of Control (I), Ivermectin (II) and Fenbendazole (III) groups on 0, 7, 14, 21days

The average EPG, geometric mean, median, range, variance of counts and standard deviation of untreated Cervids, designated as control (I group) is presented in Table1. The average EPG, geometric mean, median, range, anthelmintic efficacy (using the formula of Young et al., 2000) [30], FEC percent reduction (using the formula of Coles et al., 2006) [6], variance of counts and standard deviation in egg counts of ivermectin is illustrated in Table 2. The Cervids (Group T1) naturally infected with G.I. helminths were treated with ivermectin at the dose rate of 0.2 mg/kg body weight orally. Percent efficacy of drug was recorded on days 7th, 14th and 21st. The average EPG counts done on 7th day post treatment indicated 95% efficacy of the drug whereas 97% efficacy was observed on 14th day with 97.05% FEC reduction, lastly on 21st day, 96% anthelmintic efficacy was observed with 96.12% FEC reduction. Another group of Cervids (T2) naturally infected with G.I. helminths was treated with fenbendazole at the dose rate of 7.5mg/kg body weight orally. The average EPG, geometric mean, median, range, anthelmintic efficacy, FEC percent reduction, variance of counts and standard deviation in egg counts of fenbendazole is presented in Table 3. The drug was found to be 95.19% efficacious post 7th day with 95.05% reduction in EPG counts. On 14th day 96.08% reduction in EPG counts was recorded and anthelmintic efficacy of 96.15% was found. Reduced efficacy of 94.23% was observed by day 21st.

Compared to the control group, there was sharp decline of mean EPG in both the treated groups up to14th day post treatment. However, the mean EPG rose, post 14th day in both the groups (Fig. 1). Statistical analysis using Student's t-test revealed that, since p-value>0.05 at 95% CI, there is evidence to accept null hypothesis (H0), which explains that mean EPG reduction of both the drugs is similar. Therefore, there is no significant difference between efficacy of ivermectin and fenbendazole. Hence, both the treatments are equally effective against gastro-intestinal nematodes in Cervids of Dehradun Zoo.

4. Discussion

In the present study, efficacy of the two most commonly available and frequently used anthelmintics against gastrointestinal parasites of deer was evaluated. The infected

untreated control group continued to discharge the eggs during the entire treatment period, these observations were in accordance with Kanungo et al. (2011) and Islam et al. (2003) [17,15]. The percentage reduction in faecal egg count on day 7th and 14th for ivermectin was recorded 95.04%, 97.05% respectively and for fenbendazole 95.05% and 96.08%, respectively. As per Coles et al. (2006) [6] these values suggest that these two drugs are effective against gastrointestinal nematodes, as the percentage reduction in egg counts is more than 95.0%. In our study, ivermectin was found to be 95% effective on 7th day post treatment which is in close agreement with the observations made by Aman (2015) [1], who also found ivermectin to be 96.2% effective against naturally occurring GIN in deers of Zoological Park, Punjab. Moreover, efficacy of the drug increased to 97.0% on 14th day post treatment which is in agreement with Fathima et al. (2018) [9], who also reported 97.0% efficacy of ivermectin in spotted deer of Arignar Anna Zoological Park, Chennai. Studies of Jena et al. (2018) [16] showed ivermectin to be 100.0% effective on 14th day post treatment against gastrointestinal helminths of sheep in Jharkhand, whereas Mylrea et al. (1991) [22] suspected ivermectin resistance against gastrointestinal helminthosis in fallow deer. The faecal egg count reduction remained around 96.12% at day 21 post treatment, demonstrating higher efficacy of ivermectin. This is attributed to the usage of combination of anthelmintics, which aided in expansion of the spectrum of efficacy of drugs (Leathwick and Miller, 2013) [18]. Fenbendazole was found to be 95.19%, 96.15% effective against gastro-intestinal parasitism on 7th and 14th day post treatment, respectively. Our results were in agreement with Goossens et al. (2005) [11] who reported 94.0 to 98.0% reduction in faecal egg counts in captive wild ruminant after treatment with fenbendazole @ 7.5 mg/kg body weight in Belgium Zoo. These findings were not in agreement with Chintoan et al., (2014) [4] who reported resistance to benzimidazole and susceptible to ivermectin in wild roe deer at New Forest, UK using FECRT. However on the 21st day of post treatment there was rise in EPG leading to reduction in anthelmintic efficacy to 94.23%. These findings were in agreement with Das et al. (2009) [8] who in their study experienced parasitic build-up in enclosure, thus repeated exposure to infection. Nalubamba and Mudenda (2012) [23]

witnessed increase in EPG post 21 day of treatment. These observations were attributed to the rapid re-infection of the impala due to high pasture larval burdens and a large proportion of parasitically naïve young. The suckling impala, did not took in as much of the anthelmintic impregnated salt lick as the older ones but have increased EPGs until they attain 'self-cure' rendering the treatment to be less efficacious. A similar phenomenon has been shown by other researchers in Soay sheep in Belgium (Goossens et al., 2006) [12]. Arithmetic means may be more reliable when applied to small groups such as those involved in this study (Craven et al., 1998) [7], whereas geometric means may be more suitable in large herds (Vercruysse et al., 2001) [28]. Results from two of the studies in red deer (Mackintosh et al., 1985; Watson and Charleston, 1985) [19, 29] suggested that deer are able to metabolise and excrete benzimidazole compounds more quickly as compared to sheep and cattle, it is therefore proposed that anthelmintics should be administered to these captive Cervids for a longer period of 3 days using various delivery methods including feed and mineral licks.

5. Conclusion

It can be concluded that treatment with both ivermectin and fenbendazole against gastrointestinal nematodosis in Cervids was found to be equally effective. The finding of this research will be helpful to know about the anthelmintic therapy by the veterinarian and zoo keepers. As is apparent from our study, it was not possible to entirely eliminate the parasitic burden in animals which was evident by the reoccurrence of infection, which mostly occurs due to stress and close proximity of animals in captive conditions as well as mass deworming, where it becomes tough to ensure whether each animal has consumed the drug. In order to combat existing infection suitable measures like alternating of the anthelmintic class annually between a benzimidazole and macro-cyclic lactones should be employed along with pasture hygiene for small establishments, reducing stocking densities (Isaza et al.,1995) [14] and dietary modulation for increased immunity to GI nematodes (McClure, 2008) [21].

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7. References

- Aman DM. Studies on the prevalence and management of parasitic infections in zoo animals. Thesis, Doctor of Philosophy, Guru Angad Dev Veterinary and Animal Sciences University. Ludhiana, India; c2015. p. 185.
- 2. Banerjee PS, Garg R, Yadav CL, Ram H. Parasitic infections in some wild animals of Uttaranchal. Indian J. Anim. Sci. 2005;75:206-208.
- 3. Castillo-Alcala F, Wilson PR, Pomroy WE, Simone, Hoskin SO. A survey of anthelmintic use and internal parasite control in farmed deer in New Zealand. New Zealand Veterinary Journal. 2007;55(2):87-93.
- 4. Chintoan-Uta C, Morgan ER, Skuce PJ, Coles GC. Wild deer as potential vectors of anthelmintic-resistant abomasal nematodes between cattle and sheep farms.

- Biological Sciences. 2014;281(1780):20132985.
- 5. Coles GC, Bauer C, Borgsteede FHM, Geerts S, Klei TR, Taylor MA *et al.* World Association for the Advancement of Veterinary Parasitology (W.A.A.V.P.) methods for the detection of anthelmintic resistance in nematodes of veterinary importance. Vet. Parasitol. 1992;44:35-44.
- Coles GC, Jackson F, Pomroy WE, Prichard RK, von Samson-Himmelstjerna G, Silvestre A, et al. The detection of anthelmintic resistance in nematodes of veterinary importance. Vet. Parasitol. 2006;136(3-4):167-185
- Craven J, Bjorn H, Henriksen SA, Nansen P, Larsen M, Lendal S. Survey of anthelmintic resistance on Danish horse farms, using 5 different methods of calculating faecal egg count reduction. Equine Veterinary Journal. 1998;30:289-293.
- 8. Das GM, Srivastav A, Chakraborty D, Gupta SK, Nigam P. Stress free oral medication in captive cervids. J. Threat. Taxa. 2009;1(9):495-496.
- 9. Fathima JA, Palanivelrajan M, Gomathinayagam S, Jayathangaraj MG, Prathipa A. Evaluation of Anthelmintic Efficacy of Albendazole, Ivermectin and Levamisole in Captive Spotted Deer (*Axis Axis*). Indian Vet. J. 2018;95(5):45-48.
- Geary TG, Hosking BC, Skuce PJ, Holdsworth P, Pomroy W, Vercruysse J. World Association for the Advancement of Veterinary Parasitology (WAAVP) Guideline: Anthelmintic combination products targeting nematode infections of ruminants and horses. Vet. Parasitol. 2012;190:306-316.
- 11. Goossens E, Dorny P, Vercammen F, Vercruysse, J. Field evaluation of the efficacy of fenbendazole in captive wild ruminants. Vet. Rec. 2005;157:582-586.
- 12. Goossens E, Vercruysse J, Vercammen F, Dorny P. Evaluation of three strategic parasite control programs in captive wild ruminants. Journal of Zoo and Wildlife Medicine. 2006;37(1):20-26.
- 13. Goossensa E, Dorny P, Boomker J, Vercammen F, Vercruysse, J. A 12-month survey of the gastrointestinal helminths of antelopes, gazelles and giraffids kept at two zoos in Belgium. Vet. Parasitol. 2005;127:303-312.
- 14. Isaza R, Courtney CH, Kollias GV. The prevalence of benzimidazole-resistant trichostrongyloid nematodes in antelope collections in Florida. Journal of Zoo and Wildlife Medicine. 1995;26(2):260-264.
- 15. Islam SKMA, Ahmed S, Hoque MA, Alim MA, Hassan MM. Gastrointestinal parasites of captive deer and their response to selected anthelmintic. Bangladesh Veterinary Journal. 2003;37:63-66.
- Jena A, Kumari L, Deb AR, Joshi SK, Biswal SS. Efficacy of fenbendazole and ivermectin against naturally occurring gastrointestinal helminths of Chottanagpuri sheep at Ranchi, Jharkhand. The Pharma Innovation Journal. 2018;7(1):550-552.
- 17. Kanungo S, Das A, Das Gupta M, Huque Fazlul AKM. Haematological alterations and therapeutic efficacy of selected anthelmintics in gastrointestinal nematode infected Spotted Deer (*Cervus axis*). Wayamba Journal of Animal Science. 2011;3:107-109.
- 18. Leathwick DM, Miller CM. Efficacy of oral, injectable and pour-on formulations of moxidectin against gastrointestinal nematodes in cattle in New Zealand.

- Veterinary Parasitology. 2013;191:293-300.
- 19. Mackintosh CG, Mason PC, Manley T, Baker K, Littlejohn R. Efficacy and pharmacokinetics of febantel and ivermectin in red deer (*Cervus elaphus*). New Zealand Veterinary Journal. 1985;33:127-131.
- 20. Mason P. Parasites of deer in New Zealand. New Zealand Journal of Zoology. 1994;21:39-47.
- 21. McClure SJ. How minerals may influence the development and expression of immunity to endoparasites in livestock. Parasite Immunology. 2008;30(2):89-100.
- 22. Mylrea GE, Mulley RC, English AW. Gastrointestinal helminthosis in fallow deer (*Dama dama*) and their response to treatment with anthelmintics. Aust. Vet. J. 1991;68(2):74-75.
- 23. Nalubamba KS, Mudenda NB. Anthelmintic efficacy in captive wild impala antelope (*Aepyceros melampus*) in Lusaka, Zambia. Vet. Parasitol. 2012;186:532-537.
- 24. Niranjan AS, Singh VS, Vatsya S, Singh JL, Bharadwaj Y. Parasitic infection in captive wild animals of Nainital zoological park, India. The Pharma Innovation Journal. 2022;SP-11(7):3308-3313.
- Panda BB, Gaur K, Kori ML, Tyagi LK, Nema RK, Sharma CS, et al. Anti-Inflammatory and analgesic activity of *Jatropha gossypifolia* in experimental animal models. Global Journal of Pharmacology. 2009;3(1):1-5.
- 26. Singh P, Gupta MP, Singla LD, Sharma S, Sandhu BS, Sharma DR. Parasitic infections in wild herbivores in the Mahendra Choudhury zoological park, Chhatabir, Punjab. Zoos print journal. 2006;21(11):2459-2465.
- 27. Thawait VK, Maiti, SK. Prevalence of Gastro-intestinal Parasites in Captive Wild Animals of Kanan Pandari Zoo, Bilaspur. J Anim Res. 2015;5(1):199-202.
- 28. Vercruysse J, Holdsworth P, Letonja T, Barth D, Conder G, Hamamoto K, *et al.* International harmonisation of anthelmintic efficacy guidelines. Veterinary Parasitology. 2001;96:171-193.
- 29. Watson TG, Charleston WAG. The significance of parasites in farmed deer. In: Fennessy PF, Drew KR. ed. Biology of deer production. The Royal Society of New Zealand bulletin. 1985;22:105-117.
- 30. Young KE, Jensen JM, Craig TM. Evaluation of anthelmintic activity in captive wild ruminants by fecal egg reduction tests and a larval development assay. J. Zoo Wildl. Med. 2000;31:348-352.