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Effect of physiological parameter on the radial growth and sclerotial formation of *Rhizoctonia solani* Kuhn causing web blight of mungbean

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Abstract

Rhizoctonia solani Kuhn, is one of the most important pathogens of mungbean, causing aerial blight, which accounts for heavy losses in mungbean growing areas of MP. Three most common factors associated with the growth and sclerotia formation of pathogen is media, pH and temperature which has been studied in the present study. All three factors significantly affect the growth of the fungal mycelia and sclerotial production. In this investigation, The growth of *R. solani* was investigated using a total of seven solid mediums from which PDA media produced maximum mycelium growth (90.00 mm) and from seven different liquid media Potato dextrose broth (776.33 mg) produced the highest dry mycelial weight. The mycelium growth was tested at different temperatures *viz.*, 10, 15, 20, 25, 30, 35 and 40°C in *in vitro* conditions, among which maximum mycelium growth was observed at 30° C (88.33 mm) and among different pH *viz.*, 4.5, 5.0, 5.5, 6.0, 6.5, 7.0, 7.5, and 8.0 the highest mycelium growth was recorded at pH 7.0 (89.10 mm).

Keywords: Rhizoctonia solani, media, pH, temperature, web blight

Introduction

Mungbean (Vigna radiata (L.) Wilczek) is an important pulse crop In Madhya Pradesh. It belongs to family the Fabaceae or Leguminous. The origin of mungbean is the Indian subcontinent. It is also known as golden gram, mung and green gram. For India's vegetarian population, the mungbean is a significant source of nutritional proteins, vitamins and minerals. Mungbean seeds contain 22.9 gm protein, 61.8 gm carbohydrate, 1.2 gm fat, 4.4 gm crude fibre and 3.5 gm ash per 100 gm of the sample (Adsule et al., 1986)^[1]. Sprouting, there is an increase in thiamine, niacin, and ascorbic acid concentration. Mungbean has been demonstrated to have anti-hypertensive, anti-inflammatory, anti-diabetic and anti-tumour properties. (Matousek 2009)^[14]. The poor productivity of the mungbean has likely been caused by a number of biotic and abiotic factors. Among the various biotic factors, Web blight disease of the mungbean caused by Rhizoctonia solani Kühn [Teleomorph: Thanatephorus cucumeris (Fr.) Donk] is recognized as one of the most serious limitations. In MP, it affects mungbean productivity by up to 20–30%. Grain production and seed quality are seriously affected by this disease. The only sclerotia of R. solani undergo direct myceliogenic germination, through which vegetative hyphae capable of infecting the host, grow directly out of the sclerotium unlike many other sclerotia forming plant pathogens (Coley-Smith JR and Cooke RC.1971)^[3]. At present, very little information is available on the survival of R. solani isolated from mungbean. Therefore it was important to study the effect of pH, temperature and media on mycelia growth and sclerotia formation of the pathogen.

Materials and Methods Isolation of pathogen

The leaves of mungbean, which showed the characteristic symptoms of web blight disease was collected from research farm of Jawaharlal Nehru Krishi Vishwa Vidyalaya, Jabalpur M.P. For pathogen isolation, the lesions that displayed the first and most noticeable symptoms were selected. Using sterile water, infected leave was washed to remove any remaining soil. and control the surface contamination. With a disinfected blade, young diseased leaves were divided into 2 to 3 mm pieces (containing 2/3 healthy and 1/3 diseased sections).

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These leaf pieces were surface sterilized using a solution of 1% sodium hypochlorite and after sterilization, leaf pieces were washed with distilled water thrice. Then, under aseptic circumstances in the inoculation chamber, they were put on pre-sterilized blotter paper to make them free of any excessive moisture. The Pieces of the diseased leaves were transferred directly to the PDA (Potato Dextrose Agar) medium. Petri plates were kept in B.O.D. (Biological Oxygen Demand) Incubator at a temperature of $30\pm$ °C for 48 hrs.

Identification of the isolate of *R. solani*

Pathogens were identified based on cultural and morphological traits. Slide was prepared with lactophenol stain and for the morphological characteristics inspected under a compound microscope.

Effect of culture media

The seven cultures medium listed below were tested to see which was the best for sclerotia formation and mycelial growth.

- 1. Potato Dextrose agar medium (PDA) (peeled and sliced potato 200g, Agar-agar 20g, Dextrose 20g)
- Richard's agar (RA) medium (Potassium monobasic phosphate 5g, Potassium nitrate 10g, Magnesium sulphate 2.5g, Ferric chloride 0.02g, Sucrose 50g, Agaragar 20g)
- Czapek's Dox agar (CDA) medium (Sodium nitrate 2g, Magnesium sulphate 0.5g, Di potassium phosphate 1g, Ferrous sulphate 0.01g, Potassium chloride 0.5g, Sucrose 30g, Agar-agar 20g)
- 4. Asthana and Hawker's medium (D-Glucose 5g, potassium dihydrogen phosphate 1.75g, Potassium nitrate 3.50g, Magnesium sulphate 0.75g, Agar-agar 20g)
- Ashby's agar medium (Mannitol 20g, Magnesium sulphate 0.2g, Di potassium phosphate 0.2g, Potassium sulphate 0.1g, Sodium chloride 0.2g, Calcium carbonate 5g, Agar-agar 15g, final pH (at 25 °C) 7.4±0.2).
- 6. Brown agar (BA) medium (Dextrose 2g, Magnesium sulphate 0.75g, Tri basic potassium phosphate 1.25g, Agar-agar 20g)
- 7. Coon's agar (CA) medium (Sucrose 7.2g, Magnesium sulphate 1.23g, Potassium diphosphate2.72g, Potassium nitrate 2.02g, Dextrose 3.60g, Agar- agar 15g).

Effect of pH

Using a calibrated pH metre, potato dextrose agar's pH was adjusted to eight different values, including 4.5, 5.0, 5.5, 6.0, 6.5, 7.0, 7.5, and 8.0. Using 0.1 N NaOH or 0.1 N HCl solutions before the PDA medium was autoclaved. Three replications were kept for each pH value. 5mm mycelium discs were used to inoculate on Petri plates containing sanitized PDA media, which were then incubated at a temperature of 30 ± 1 °C. The mycelium growth was measured at48 hours and 96 hours. Sclerotia formation test ranges on various pH levels were reported.

Effect of temperature

The purpose of the studies was to determine the ideal temperature for mycelial growth and sclerotia formation of R. *solani*. The 5 mm disc of the test pathogen was inoculated on sterilized PDA medium, poured in petri discs. Three replications were kept for each different temperature. The petri dishes were incubated at seven different temperature 10

°C, 15 °C, 20 °C, 25 °C, 30 °C, 35 °C and 40 °C. The mycelium growth was recorded at 48 hours and 96 hours and sclerotia formation was also recorded.

Results and Discussion

Solid medium

The growth of Rhizoctonia solani was investigated using a total of seven solid mediums (Table 1). Maximum mycelium growth was found on PDA medium. Czapek's agar media came in second place with a colony diameter of 79.33 mm, followed by Richard's agar medium (56.83 mm). Minimum colony growth diameter was recorded in Ashby's agar medium. After 15 days of inoculation the formation of sclerotia was observed in which excellent sclerotia formation was seen in Potato dextrose agar medium and Czapek's Dox agar medium. Good sclerotia was seen in Richard's agar medium, fair in Brown's agar and Coon's agar medium. Asthana and Hawker's agar and Ashby's agar medium exhibited poor sclerotia formation. The present finding has been supported by other workers. According to Huang-Shi Chen and U-XiYing (2004)^[6], PDA was the best growing medium for *Rhizoctonia solani*. and Ritchie et al., (2009)^[17] also observed from research and conclude that in all the tested media for R. solani the number of sclerotia produced was more when they were grown on PDA.

Liquid medium

Seven different liquid mediums were used to study the growth of Rhizoctonia solani. Potato dextrose broth (776.33 mg) produced considerably more dry mycelial weight in comparison to the other liquid media. Richard's broth (601.67 mg) was the next best medium in terms of merit. Dry weight of mycelium was good in Czapek's Dox broth (589.33 mg) and Coon's broth (382.67 mg) but very poor in Brown's broth, and Ashby's broth (Table 1). The development of sclerotia was seen after 15 days of inoculation. On Potato dextrose broth. Czapek's Dox and Richard's broth. R. solani produced excellent sclerotia. On Asthana and Hawker's broth, Coon's and Brown's broth it was found to be fair. Although on Ashby's broth, it was poor. These findings resemble with the observations of Khan et al., (2016)^[9] also found that the R. solani biomass produced on potato dextrose broth was the highest.

Temperature

The growth of *R. solani* was studied at seven different temperature levels viz. 10, 15, 20, 25, 30, 35 and 40 °C. the maximum radial growth was recorded at 30 °C (88.2mm), followed by 25 °C (79.2mm), 20 °C (72.06 mm), 35 °C (65.50 mm), and 40 °C (29.40 mm).and minimum radial growth was recorded at 10 °C (18.3 mm) (Table 2, Figure-1). After 15 days the formation of sclerotia was observed. Abundant sclerotia production was found at 30 °C and 25 °C. Sclerotia did not develop at 10 °C, 15 °C, or 40 °C. The present findings have been supported by other researchers Meena and Chattopadhyay (2002) ^[15] also found highest growth of *R. solani* found at 30 °C temperature. Grosch and Kofoet (2003) ^[7] also found that 20°-30 °C temperature was best for growth of *R. solani*.

pН

The effect of seven pH levels, namely 5.0, 5.5, 6.0, 6.5, 7.0, 7.5, and 8.0, on sclerotia formation and radial growth of *R*.

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solani was investigated (Table 3; Figure -2). The maximum colony diameter was found at pH 7.0 (89.10 mm). The next maximum growth was recorded at 6.5 pH (77.06 mm) followed by 7.5 pH (74.43 mm). Pathogen growth was reportedly inhibited by pH values of 5.0 and 8.0 and the growth was 17.06 mm and 23.40 mm, respectively. These results give conformance to the finding of Kumar et al., (2014)^[10] found that the optimum pH for mycelium growth and sclerotia production was pH 7.0 reported that best growth and slcerotia formation of R. solani occurred best at pH 6.0.

Table 1: Effect of solid and liq	id media on radial	growth and sclerotia	formation Rhizoctonia solani

Name of the medium	Solid media		Liquid media	
	Colony diameter (mm)	Sclerotia	Dry mycelium weight (mg) after	Sclerotia
	after 96 hrs*	formation	21 days	formation
Potato dextrose agar	90.00	++++	776.33	++++
Czapek's agar	88.17	++++	589.33	++++
Richard's agar	87.20	++++	601.67	+++
Coon's agar	72.27	+++	382.67	++
Brown's agar	65.40	++	147.10	+
Asthana and Hawker's agar	60.43	++	263.00	++
Ashby's agar	23.40	+	102.10	+
SE(m) ±	1.163		1.298	
CD(0.05)	3.562		2.342	

*mean of three replications

++++ excellent Sclerotia formation, +++ good Sclerotia formation, ++ Fair Sclerotia formation, + poor Sclerotia formation

Table 2: Effect of different temperature on radial growth and sclerotia formation Rhizoctonia solani

S. No.	Temp.°C	Colony diameter (mm) after 96 hrs*	Sclerotia formation
1	10	18.33	+
2	15	56.50	+
3	20	72.06	++
4	25	79.06	++++
5	30	88.33	++++
6	35	65.50	+
7	40	29.40	+
S	$E(m) \pm$	0.525	
CD(0.05) 1.607		1.607	

*mean of three replications

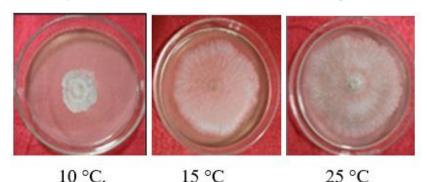
++++ excellent Sclerotia formation, +++ good Sclerotia formation, ++ Fair Sclerotia formation, + poor Sclerotia formation

Table 3: Effect of various pH levels on radial growth and sclerotia formation Rhizoctonia solani

S. No.	pН	Colony diameter (mm) after 96 hrs*	Sclerotia formation after 15 days
1	5.0	17.06	+
2	5.5	52.03	+
3	6.0	70.10	+++
4	6.5	77.06	++++
5	7.0	89.10	++++
6	7.5	74.43	+++
7	8.0	23.40	+
SE(m)) ±	0.719	
CD(0.	05)	2.202	

*mean of three replications

++++ excellent Sclerotia formation, +++ good Sclerotia formation, ++ Fair Sclerotia formation, + poor Sclerotia formation



10 °C. 15 °C

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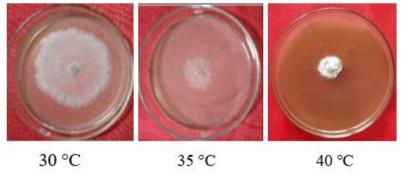


Fig 1: Effect of different temperatures on mycelium growth of R.solani



pH7

рН7.5 рН8

Fig 2: Effect of different pH on mycelium growth of R. Solani

Conclusion

Rhizoctonia solani Kuhn causes aerial blight, which contributes for significant losses in MP mungbean growing areas. The three most common parameters linked with pathogen growth and sclerotia formation addressed in this study are medium, pH, and temperature. All three of these parameters have a substantial impact on the proliferation of fungal mycelia and sclerotial formation. PDA media yielded the most mycelium growth (90.00 mm), whereas Potato dextrose broth (776.33 mg) yielded the greatest dry mycelial weight of the test pathogen. Mycelium growth was greatest at 30 °C (88.33 mm) and pH 7.0 (89.10 mm).

Reference

- Adsule RN, Kadam SS, Salunkhe DK, Luh BS. Chemistry and technology of green gram (*Vigna radiata* [L.] *Wilczek*). Critical Reviews in Food Science & Nutrition. 1986;25(1):73-105.
- 2. Bara B. Epidemiology and management of web blight disease of Urdbean (Doctoral dissertation, Ph.D. Thesis Birsa Agricultural University, Kanke, Ranchi, Jharkhand, 2007, p. 235.
- Coley-Smith JR, Cooke RC. Survival and germination of fungal sclerotia. Annual review of phytopathology. 1971;9(1):65-92.
- Datta S, Das S, Sarkar A, Tarafdar J, Chowdhury A. Assessing the effects of varied temperature and pH on the growth and sclerotial formation of Rhizoctonia solani Kuhn, isolated from paddy field: a case study. International Journal of Life Sciences. 2014;8:4-9.
- 5. Dwivedi RP, Saksena HK. Web blight disease of arhar (*Cajanus cajan* (L.) *Millsp.*) caused by *Thanatephorus cucumeris*. Indian journal of farm sciences; c1975.

- Shichen H, Xiying L. Study on biological characters of Rhizoctonia solani on Rhodiola sachalinen. *Jilin Nong ye da xue xue bao* Acta Agriculturae Universitatis Jilinensis. 2004;26(1):39-41.
- Grosch R, Kofoet A. Influence of temperature, pH and inoculum density on bottom rot on lettuce caused by *Rhizoctonia solani* Einfluss von Temperatur, pH und Inokulumdichte auf die Salatfäule verursacht durch *Rhizoctonia solani*. Zeitschrift für Pflanzenkrankheiten und Pflanzenschutz. Journal of Plant Diseases and Protection; c2003. p. 366-378.
- Jalali BL. Observation on Rhizoctonia leaf blight of rice bean from HAU, Hisar. Consolidated Report on Kharif Pulses; c1988. p. 144-145.
- Khan I, Hussain H, Shah B, Ullah W, Naeem A, Ali W, et al. Evaluation of phyto biocides and different culture media for growth, isolation and control of *Rhizoctonia* solaniin vitro. Journal of Entomology and Zoology Studies. 2016;4(2):417-420.
- Santosh K, Amarendra K, Gireesh C, Mehi L, Rakesh K. Dynamics of mycelial growth and sclerotia production of Rhizoctonia solani Kuhn (AG1-IB) of Urdbean. The Ecoscan. 2014;8(3&4):273-277.
- 11. Kumar S, Tripathi H, Singh D. Evaluation of Selected Systemic and Non-Systemic Fungicides *in vitro* and *in vivo* condition against Web Blight Disease of Urd Bean caused by *Rhizoctonia solani* Kuhn. Journal of Agriculture Search. 2014;1(1):45-48.
- 12. Kushwaha SK, Kumar S, Chaudhary B, Sahu R. Effect of different media, ph and temperature on growth and sclerotia formation of *Sclerotium rolfsii* Sacc. Causing

collar rot of lentil. Chemical Science Review and Letters. 2019;8(29):01-05.

- Lakshmanan P, Nair MC, Menon MR. collar rot and web blight of cowpea caused by *Rhizoctonia-solani* in Kerala, INDIA. Plant Disease Reporter. 1979;63(5):410-413.
- 14. Lee SJ, Lee JH, Lee HH, Lee S, Kim SH, Chun T, Imm JY. Effect of mung bean ethanol extract on proinflammtory cytokines in LPS stimulated macrophages. Food Science and Biotechnology. 2011;20, 519-524.
- 15. Matousek J, Podzimek T, Pouckova P, Stehlik J, Skvor J, Soucek J, *et al.* Antitumor effects and cytotoxicity of recombinant plant nucleases. Oncology Research Featuring Preclinical and Clinical Cancer Therapeutics. 2009;18(4):163-171.
- 16. Meena PD, Chattopadhyay C. Effect of some physical factors, fungicides on growth of *Rhizoctonia solani Kuhn* and fungicidal seed treatment on groundnut seed germination. Indian Journal of Plant Protection. 2002;30(2):172-176.
- 17. Muhsin TM, Selman MS. *In vitro*, optimization of growth and bioactivity of antibacterial metabolite produced by *Rhizoctonia solani* Kuhn. Journal of Basrah Researches (Sciences), 2013, 39(1).
- Ritchie F, Bain RA, McQuilken MP. Effects of nutrient status, temperature and pH on mycelial growth, sclerotial production and germination of *Rhizoctonia Solani* from potato. Journal of Plant Pathology. 2009, 589-596.
- Saksena HK, Dwivedi RP. Web blight of black gram caused by *Thanatephorus cucumeris*. Indian Journal Farm Science. 1973;1:58-61
- 20. Singh A, Malhotra SK. Host range studies of *Rhizoctonia Solani* causing web blight of winged bean. Bhartiya Krishi Anusandhan Patrika. 1994;9(2):113-116.
- 21. Tripathi HS. Current status of research on web blight disease of urd bean: a review. Agricultural Reviews. 2007;28(1):1-9.
- 22. Yao Y, Chen F, Wang M, Wang J, and Ren G. Antidiabetic activity of Mung bean extracts in diabetic KK-Ay mice. Journal of agricultural and food chemistry. 2008;56(19):8869-8873.