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Seed coating technology: A sustainable approach for improving seed quality and crop performance

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Abstract

The initial growth phase of agricultural plants, called crop establishment, is crucial and influenced by the quality of the seed used. Seed quality can be affected by various aspects such as genetics and environmental situations, foremost to financial losses for farmers. It is challenging for farmers to control all these factors, but seed coating technology can help by applying physical, chemical, or biological components to the seed surface to enhance its physical properties. Seed coating techniques such as pelleting, encrusting, and film coating increase seed weight and improve crop performance. Seed coating can be a useful means for addressing agricultural challenges and restoring degraded systems, ultimately contributing to improved food security and a more cost-effective ecosystem. This paper provides evidence on different seed coating technologies and discusses their possible benefits.

Keywords: Seed, seed quality, seed coating, seed invigoration, sustainable agriculture

Introduction

Farmers constantly seek high seed quality since it can enhance crop yields by up to 30%. High-quality seeds must be sown, but their use does not ensure effective standard establishment. (FAO, 2014, p. 4) ^[25]. A wide variety of biotic and abiotic stresses can affect seeds, lowering their expected performance. Yet, careful practice of chemical, biochemical, and biological seed treatments can safeguard and improve development, and prospective yield. (Taylor *et al.* 2020) ^[71]. With current conventional agricultural practices being unsustainable in addition to the impending impact of climate alteration, there is a essential solutions that can improve agricultural productivity while also promoting environmental and economic sustainability (FAO, 2017). Plant helpful microorganisms (PBMs) are being considered as a natural alternative to conventional farming practices to reduce environmental harm. PBMs can assist plants in maintaining or increasing yields though dipping the need for agrochemicals, restoring soil fertility, and addressing issues triggered by biotic and abiotic stresses. Although the use of PBMs has been gaining popularity, conventional agricultural practices such as soil tillage, fertilization, and pesticide application can negatively affect soil microorganisms and their interactions with plants, limiting the effectiveness of PBMs (Ma *et al.* 2019) ^[53]. Seed inoculation has been identified as an accurate and profitable method for delivering microbial inoculants, where a seed is coated with a carrier, binder, and an active agent such as a microbial inoculant before being planted (Rocha *et al.* 2019) ^[60].

Seed coating as a technology for enhancement of seed quality was first used for cereal seeds in the late 1930s by “Germins” a British corporation. After that, this technology was commercially accepted and started its application in the late 1960s (Ahmed *et al.*, 2021) ^[7]. This is one of the components of precision as it encourages quality seed sowing, Uniform size of the seed is maintained when we consider small sized seeds. The coating not only helps in enhancing good germination but also precision seed sowing as seed size is maintained due to uniform coating of every single seed as a result coated seeds are used through seed drill sowing which promotes sowing accuracy. Seed coating can be an effective technique for enhancing seed quality when it includes an active ingredient. This active ingredient can be a physical, chemical, or biological component such as a microbial inoculant that is adhered to the seed coat with the assistance of a binder or filler acting as a carrier. Biological seed treatments are commonly used for pest management and as bio-stimulants, which are natural substances that stimulate physiological and molecular processes that control crop yield and quality. There are various categories of plant bio stimulants, which are ordinary products or biologicals.

This review presents the most recent data on several seed coating techniques and seed coatings, as well as their possible impacts on crop performance.

Seed coating technology

Seed coating is a process of applying a thin layer of protective material to seeds to enhance their germination, growth, and survival. The practice has increased widespread status among farmers and horticulturists due to its ability to improve seed performance and plant establishment. Seed coating has been widely studied and developed over the years, with research focused on improving the effectiveness and efficiency of the process. According to a study by Hafeez *et al.* (2019) [76], seed coating can significantly progress seed quality also plant performance. The researchers found that seed coating with a

combination of fungicide and insecticide enhanced seedling emergence, growth, and yield in wheat crops. In one more study by Khan *et al.* (2020) [77], seed coating with plant growth-promoting rhizobacteria (PGPR) improved seed germination, growth, and stress tolerance in maize plants. The researchers concluded that seed coating with PGPR is a cost-effective and eco-friendly method to improve seed and plant performance. Amir *et al.*, (2021) [10] concluded from a field experiment that seed coating (on hydroprimed seed) with Bio NPK and drought alleviating bacteria is helpful in improving the planting value of lentil under sub-optimal condition.

Overall, seed coating is an important agricultural practice that has significant potential to progress crop productivity and sustainability. As such, it remains to be an active zone of study and development in the agricultural industry.

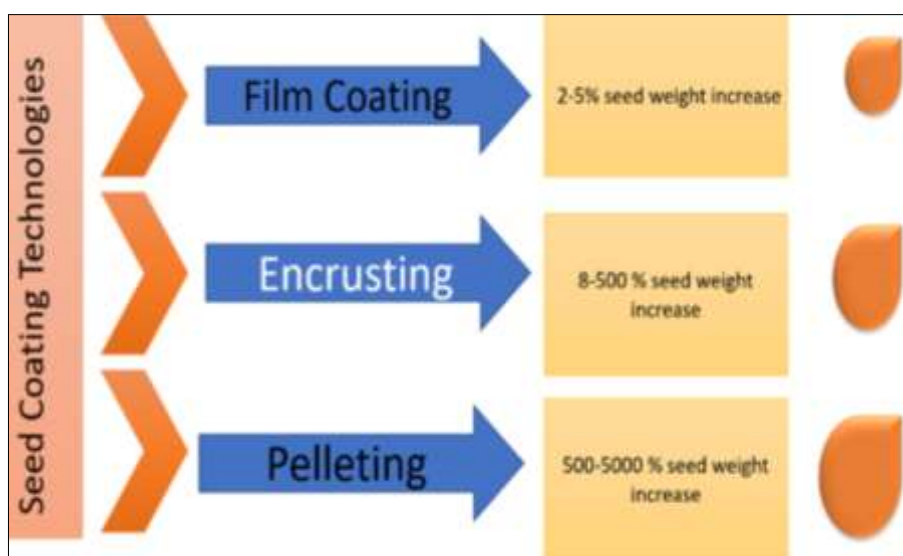


Fig 1: Types of seed coating

Seed film coating

Seed coating is a method that involves adding growth-promoting substances to seeds in order to improve their quality. This is done by applying a thin layer of a beneficial coating substance to the seed coat, which is typically less than 5% of the seed's weight. This approach is highly effective, with over 90% of treated seeds showing improved growth. (Pedrini *et al.*, 2017) [78]. By using coating technologies, seeds can be protected from various environmental factors that can harm them, such as biotic and abiotic stresses, from the time of seedling establishment to imbibition (Chandrika *et al.*, 2017) [19]. Waterlogging is a main stress that can inhibit plant growth, but coating *Brassica napus* L. seeds with a film containing uniconazole (0.0075%) has been found to significantly alleviate this stress and improve physiological and morphological characteristics. The coating also improved the activity of antioxidant enzymes such as peroxidase, catalase, and superoxide dismutase during waterlogging stress. However, the uniconazole coating did not affect germination under waterlogging stress. Another study found that canola seedlings coated with a polymer film had improved germination when subjected to moisture and low temperature stress. (Willenborg *et al.*, 2004) [78]. *Paraburkholderia phytofirmans* rhizobia has been used as a film coating agent to effectively treat wheat plants under drought stress. (Naveed *et*

al., 2014) [80]. In a study, it was found that cowpea plants subjected to water scarcity responded positively to microbial inoculation as a film coating (Rocha *et al.*, 2019 a, b) [60]. A significant improvement in disease control was observed when comparing coated seeds to untreated seeds. Pigeon pea seeds were coated with zinc and iron nanoparticles (750 ppm), resulting in a remarkable enhancement in seedling appearance, robustness, dehydrogenase, and amylase activities related to raw seeds. (Maity *et al.*, 2018) [61].

Seed encrusting

Encrusting is a seed coating process that rises seed weight by 8-100% without significantly altering the seed's shape (Pedrini *et al.*, 2017) [78]. The encrusting process involves two important phases: "before coating" (BC) and "coating phase" (CP). In the BC phase, the initial weights of the seed, precipitate, and binder are noted. The CP involves the actual coating process, where small amounts of precipitate and binder are applied to the seed to safeguard good binding (Rocha *et al.* 2019) [60]. Sunflower seeds that were coated with a seed coating material were able to protect the seeds from the harmful effects of herbicides on germination (Piveta *et al.* 2010) [67]. Encrusting fescue seeds before storage bring about in significantly higher germination and vigour compared to untreated seeds and seeds that were encrusted after storage.

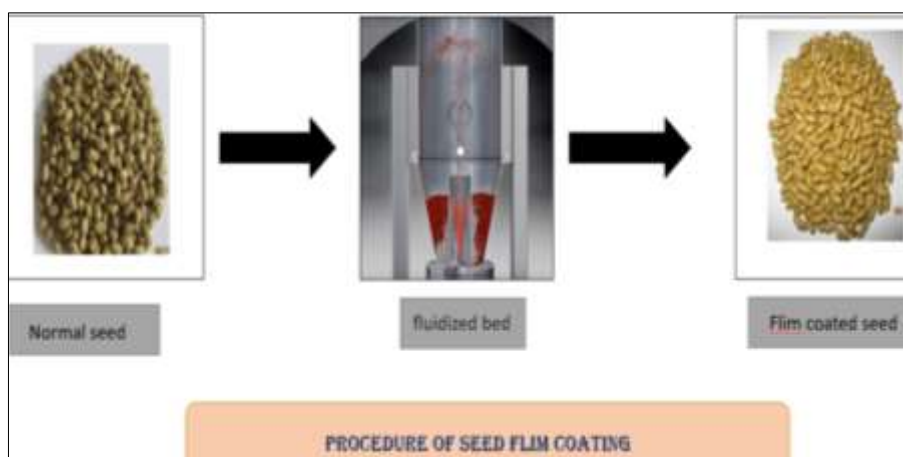


Fig 2: Procedure of seed film coating

Encrusting also had a positive effect on seedling growth and mitigated the negative effects of commercial herbicides. (Olivera *et al.*, 2017) ^[81]. Seed encrusting has been used to successfully deliver rhizobacteria or beneficial plant compounds near the sorption zone. In onion seeds coated with Thiram: Genus's coat, higher levels of germination, vigour index, root length, and shoot length were observed. Seed encrusting has been shown to progress the performance and survival rate of plant growth-endorsing rhizobacteria (PGPR) Zaheer *et al.*, 2019) ^[82]. PGPR-coated seeds have been observed to have higher viability and longer shelf life under varying temperatures and light conditions compared to untreated seeds. (Ma, 2019) ^[53]. In conclusion, seed encrusting is an effective technique for enhancing seed quality and improving the survival of beneficial microorganisms. It has also been demonstrated that seed encrusting can mitigate the harmful effects of various plant protection agents, particularly during storage.

Seed pelleting

Seed pelleting refers to a technique of adding natural or synthetic materials to seeds in a manner that increases their

size, resulting in a uniform shape that makes it difficult to distinguish the original seeds. (Pedrini *et al.*, 2017) ^[78]. Tomato seeds have been successfully improved in relations of vigor, germination, and preservation through the use of seed pelleting. (Javed and Afzal, 2020) ^[37]. Seed pelleting with a mixture of ammonium molybdate, ferrous sulphate, and rhizobium was found to enhance the growth and yield parameters of soybean (Ramesh and Thirumurugan, 2001) ^[58]. In another study, seed pelleting was shown to increase the effectiveness of strain *Paenibacillus polymyxa* E681 in biological control of plant diseases, when compared to non-pelleted seeds (Ryu *et al.*, 2006) ^[61]. For tomato seeds, their germination and emergence were improved through pelletization with a mix of talcum, calcium oxide, and bentonite (Javed and Afzal, 2020) ^[37]. Moreover, research on cowpea seeds demonstrated that seed pelleting led to improved seed quality compared to non-pelleted seeds. These studies highlight the valuable effects of seed pelleting in enhancing various aspects of seed performance and crop growth. Pelletized rice seeds exhibited superior physiological indicators and biochemical characteristics in contrast to non-pelletized rice seeds. (Sun *et al.*, 2019) ^[83].

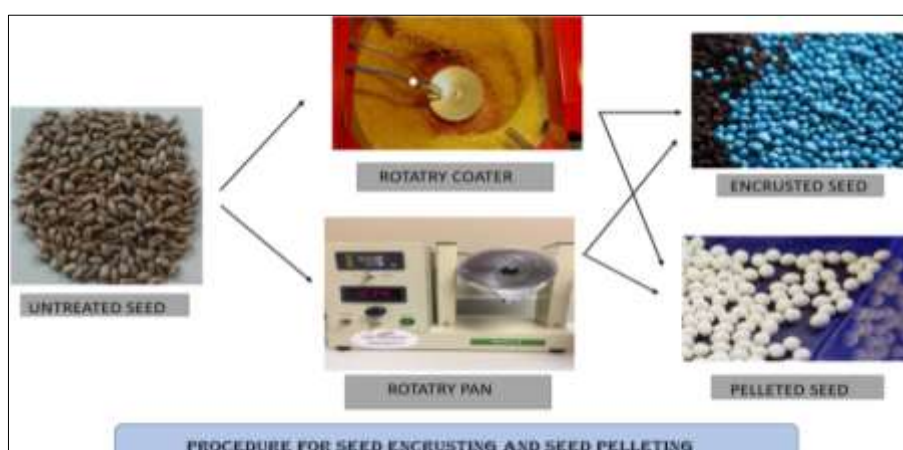


Fig 3: Procedure of seed encrusting and pelleting

Seed coating with plant beneficial microbes (PBM)

Plant beneficial microbes (PBM) play an indispensable role in fostering plant growth and preserving their well-being by establishing a mutualism bond with plants. Coating seeds with PBM is a common and real approach to promoting crop growth, increasing yield, and safeguarding plants from pathogens.

Research has demonstrated that seed coating with PBM can lead to improved seedling establishment, enhanced germination, and overall crop productivity and quality, while reducing the reliance on chemical fertilizers. Seed inoculation is an economical method for significant field applications, delivering PBM efficiently (John *et al.*, 2010; O'Callaghan,

2016) [84-85]. Compared to other methods like direct soil inoculation, foliar spray, or root dipping, seed inoculation proves to be more efficient, requiring a lower amount of microbial inoculum. By delivering PBM directly to the rhizosphere of the target crop during germination, seed inoculation establishes an intimate plant-microbe contact early on (Philippot *et al.*, 2013) [86]. Seed coating with PBM permits for a true application of a small quantity of inoculant at the seed-soil interface, confirming that beneficial microbes are available during the critical stages of germination and early plant development (Scott, 1989) [62]. This, in turn, promotes healthy and speedy plant establishment, ultimately maximizing crop production (Colla *et al.*, 2015a) [22]. This review paper focuses on various types of microorganisms known as PBM, which positively impact plant establishment, growth, and development through direct or indirect mechanisms. These microorganisms encompass plant growth-promoting bacteria (PGPB), arbuscular mycorrhizal (AM) fungi, and Trichoderma. Through their symbiotic interactions with plants, these PBM contribute significantly to agricultural sustainability and productivity.

Plant growth-promoting bacteria (PGPB)

In addition to protecting plants against biotic and abiotic challenges, plant growth-promoting bacteria (PGPB) also promote plant growth and performance through both direct and indirect methods (Lugtenberg and Kamilova, 2009). The facilitation of nutrient uptake, including nitrogen fixation, phosphorus, and potassium solubilization, are direct impacts of PGPB on plant growth. Along with helpful substances like ammonia, organic acids, and ACC deaminase, PGPB can also create plant hormones including indole-3-acetic acid and cytokinins. (Hayat, 2016) [37]. Protecting plants from harm brought on by numerous biotic stressors, such as bacteria, fungi, and nematodes, is one of the indirect impacts of PGPB on plant development and health. These protective effects are achieved primarily through the synthesis of allelochemicals, like antibiotics and hydrogen cyanide, as well as lytic enzymes. Furthermore, PGPB can activate induced systemic resistance, providing additional protection against pathogens (Glick, 2012) [88]. Among the PGPB, rhizobia are commonly employed for nitrogen fixation in pulse crops, contributing to a reduction in the need for chemical fertilizers. Amir *et al.*, (2021) [11] reported that seed coating (on hydroprimed seed) with drought alleviating bacteria + BioNPK can be used for higher seed production of lentil under sub optimal conditions. This sustainable practice not only enhances plant growth but also minimizes the reliance on synthetic fertilizers, promoting environmental and agricultural sustainability.

Arbuscular mycorrhiza and Trichoderma

Arbuscular mycorrhiza and Trichoderma are two types of fungi that have long been utilized to enhance nutrient uptake and acquisition in plants. For instance, Arbuscular mycorrhiza can contribute up to 80% of the phosphorus acquired by plants, while plants, in return, transfer about 20% of the fixed carbon to the fungi (Jeffries 2016) [46]. It has been demonstrated that using certain Arbuscular mycorrhizal fungi species, including *Glomus intraradices*, *Rhizophagus irregularis*, *Funneliformis mosseae* (previously known as *Glomus mosseae*), and *Rhizophagus fasciculatus*, can significantly increase crop growth and productivity, especially in difficult situations involving salinity and drought stress (Abdul Latef and

Chaoxing, 2011) [55]. These fungi are crucial in helping plants adapt to external stresses, which ultimately improves agricultural performance and output. When *Rhizobium* spp. are applied either alone or in conjunction with other plant growth-promoting microorganisms (PBMs), positive benefits in plant growth and production have been seen in several studies. However, due to the possibility of inconsistent results, care must be taken when mixing *Rhizobium* spp. seed inoculation with other chemical applications. For instance, when infected seeds were treated with lime, the survival of rhizobia was decreased in some species of clover. Similarly, when applied to inoculated seeds, certain fungicides including N-(trichloromethylthio)-4-cyclohexene-1,2-dicarboximide, metalaxyl-M, carbathiin, oxycarboxin, and thiram significantly affected the activity of *Rhizobium* on plant growth and function. These findings highlight the importance of doing in-depth research on the relationship between fungicides and bacterial inoculation agents. This will make it easier to choose suitable fungicides for seed coating treatments that are compatible with the particular *Rhizobium* strain. By using these measures, the effectiveness of the *Rhizobium* inoculants will be guaranteed, and successful plant growth will be encouraged.

The presence of significant free-living fungi in the soil and rhizosphere has been linked to metabolites generated during interactions between plants and *Trichoderma*. *Trichoderma* treatment can have a considerable effect on a number of plant development factors, including growth, root shape, and nutritional status. It can boost nutrient uptake and solubilization, boost nitrogen usage effectiveness, stimulate systemic resistance, and provide biocontrol against infections while also detoxifying harmful substances in the root zone. *Trichoderma* has a noticeable impact on plant growth and seed germination when administered alone. (Nawar, 2007;) [64]. Furthermore, it shows efficacy in controlling pathogenic agents like *Rhizoctonia solani* (Mihuta-Grimm and Rowe, 1986) [62], *Pythium* spp. (Sivan *et al.*, 1984; Lifshitz *et al.*, 1986; Taylor *et al.*, 1998) [68, 69, 72], *Sclerotium cepivorum*, and *Fusarium* spp. (Sivan and Chet, 1986; Sivan *et al.*, 1987; Babychan and Simon, 2017) [67, 69, 17]. Depending on the specific interactions with other bacterial species when they are coated together, the effects of *Trichoderma* on plant development and disease incidence can vary. *Trichoderma* has been seen to have a major effect on plant growth and disease incidence in greenhouse and field settings. However, depending on the exact interaction with the species when coated alongside other bacterial species, the outcome may differ. In a 2016 study, *Trichoderma parareesei*, *Pseudomonas fluorescens*, *Bacillus subtilis*, and *Azotobacter chroococcum* were combined in a talc-based consortium formulation to treat tomato seeds. This treatment significantly reduced the incidence of wilt caused by *Ralstonia solanacearum*. The application of this consortium composition also produced a considerable improvement in fruit output. This research demonstrated the effectiveness of the talc-based seed treatment in promoting disease resistance and enhancing crop productivity in tomatoes. The co-inoculation of *Trichoderma* spp., *B. bassiana*, *Metarhizium anisopliae*, and AM fungi had a negative effect on lettuce seed growth, significantly reducing it. To gain a better understanding of the impact of seed coating with bacterial species on plant growth and development at every stage of crop development, it is necessary to analyse the interaction and relationship between different species of bacterial inoculants under both laboratory and field conditions.

This statement is a paraphrase of the importance of conducting studies to investigate the effects of different combinations of bacterial inoculants on plant growth and development, as highlighted in various research papers and reviews in the field of plant-microbe interactions.

Nutrient seed coating

Coating the seeds with the essential nutrients including macronutrients and micronutrients enhances the germination as well as do not allow weed growth which is a problem during the field application of major fertilizers. Also, seed coating technique with fertilizers reduces the cost of production. It has been seen that the fertilizer application at early phase of plant growth requires more amount of nutrients rather than later

stages depending upon the size of the seed. Larger seeds demand more application than the smaller seeds. Direct application of macronutrients to the seed coat can hamper the germination and subsequent processes therefore before the application of nutrients, it is recommended to coat the seeds with other inert material. The application of slow-release nutrients on the seed results in enhanced performance and increased yield in maize (Dong *et al.*, 2016) [25]. Some cereals like rice showed better performance when seed coated with more than one nutrient *viz* mixture of nitrogen, phosphorus and lime leading to well growing and yield. Improved physiological activity has been examined in Barley with seed coating of Phosphorus has been applied (Zełonka (2005) [94].

Table 1: The effect of different nutrient application for the different crops has been illustrated in the table below.

Source	Application rate	Crop	Main finding	Reference
Boron	1.5 g/kg seed	Chickpea	Seedling growing, grain yield improved also improvement in nodulation.	(Hussain <i>et al.</i> , 2020) [36]
Fertilizer mastermins	10 mL/100 g seed	Campo Grande <i>Stylosanthes</i>	Shoot dry mass improved.	(Baroni and Vieira 2020) [13]
ZnSO ₄	8.81 g/kg of seed	Rice	Advanced actions of sulfur-metabolism-enzyme.	(Da-Costa <i>et al.</i> , 2020) [24]
Calcium peroxide	6 g/20g of seed	Rice	Increase in crop establishment.	(Javed <i>et al.</i> , 2021) [38]
zinc	1.5 g/kg seed	Wheat	Enhanced seed germination.	(Mohammad and Peksen, 2020) [63]

The low-quality seeds can result into a good germinating seed if treated with optimum amount of nutrients including both macro and micro nutrients at correct dose. The seed coating does not only help with improving germination but also it improves seedling growth (seedling length, seedling DW, shoot length, shoot DW, root length, seedling nutrient). The nutrients applied spreads out to the early root system established by the plant thus improving seedling growth. The percentage of abnormal seedlings can be reduced through nutrient coating. The optimum plant nutrient formulation leads to proper development of emerging coated seeds. This was observed in one of the experiments showing higher germination and speed of seedling emergence when Tomato seeds treated with K₂SO₄ (0.5 mol dm⁻³), Ca(NO₃)₂ (2.5 mol dm⁻³), KH₂PO₄ (0.05 mol dm⁻³), MgSO₄ (0.3 mol dm⁻³), KCl (0.05 mol dm⁻³), FeEDTA (0.05 mol dm⁻³), H₃BO₃ (0.005 mol dm⁻³), MnSO₄·4H₂O (2.5 × 10⁻⁴ mol dm⁻³), CuSO₄·5H₂O (5 × 10⁻⁵ mol dm⁻³), ZnSO₄·7H₂O (1.5 × 10⁻⁴ mol dm⁻³), (NH₄)₆Mo7O₂₄·H₂O (2.5 × 10⁻⁶ mol dm⁻³) formulation of nutrients. This nutrient formulation also led to stimulation of total dehydrogenase activity at 48 hours after seed imbibition. Seeds coating with plant nutrient formulation had significantly higher absorption and retention of nutrients in the tomato seedlings at 14 and 30 DAS.

Conclusion

With the main goal of increasing yields through improved establishment, development, and overall crop growth, seed coating technology has shown significant potential for improving seed quality. A crop's ability to grow depends greatly on the quality of its seeds, which are judged by their vigour and potential for emergence. Better stand establishment and seedling vigour can result from the proper application of a suitable seed coating agent in the proper concentration. The difficulties that seedlings face during the development phase, however, cannot be solved with a one-size-fits-all approach, relying on a single or a few modest seed coatings as a universal cure for all limitations in early seedling growth and

development. It is necessary to build solid concepts and methodologies for seed coating research as well as a thorough understanding of the complex interactions that are present in seed coatings in order to make meaningful progress in the field. Collaboration amongst experts in a variety of disciplines, including chemical and mechanical engineering, agronomy, soil science, and microbiology, is crucial to achieving this. Such cross-disciplinary cooperation will aid in risk reduction and issue resolution related to seeds, ultimately resulting in higher seed quality. To understand the behaviour and performance of coating agents and to enable the creation of efficient guidelines and processes for seed coatings, it is crucial to have a strong foundation of fundamental knowledge. Additionally, more investigation through transcriptome and proteomic studies is necessary to advance our understanding of the molecular pathways underpinning seed coating-induced seed quality development. Exploring these systems will provide priceless information and boost seed coating technology.

Availability of data and materials

As authors, we will ensure that the availability of data and materials used in the review article on is clearly stated for transparency and reproducibility.

Competing Interests

As a responsible author, I declare that there are no competing interests that may have influenced this manuscript. However, if there are any potential conflicts of interest that were inadvertently omitted, they will be disclosed to the journal immediately.

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References

- Accinelli C, Abbas HK, Little NS, Kotowicz JK, Mencarelli M, Shier WT. A liquid bioplastic formulation for film coating of agronomic seeds. *Crop Prot.* 2016;89:123-128. <https://doi.org/10.1016/j.cropro.2016.07.010>.
- Accinelli C, Abbas HK, Shier WT, Vicari A, Little NS, Aloise MR, *et al.*, Degradation of microplastic seed film-coating fragments in soil. *Chemosphere.* 2019;226:645-650.
- Afzal I, Basra SMA, Cheema MA, Farooq M, Jafar MZ, Shahid M, *et al.*, Seed priming: a shotgun approach for alleviation of salt stress in wheat. *Int. J Agric. Biol.* 2013;15(6):1199-1203.
- Afzal I, Javed T, Amirkhani M, Taylor AG. Modern seed technology: seed coating delivery systems for enhancing seed and crop performance. *Agriculture.* 2020;10(11):526. <https://doi.org/10.3390/agriculture10110526>.
- Afzal I, Rehman HU, Naveed M, Basra SMA. Recent advances in seed enhancements. New challenges in seed biology-basic and translational research driving seed technology. In *Tech*; c2016. p. 47-74. <https://doi.org/10.5772/6479110.5772/64791>
- Afzal I, Saleem S, Skalicky M, Javed T, Bakhtavar MA, *et al.*, Magnetic field treatments improve sunflower yield by inducing physiological and biochemical modulations in seeds. *Molecules.* 2021;26(7):2022. <https://doi.org/10.3390/molecules26072022>.
- Ahmed W, Anwar SA, Tariq M, Rehman R, Islam W. Seed Coating: A Promising Technology for Sustainable Agriculture. In *Biotechnology for Sustainable Agriculture.* Springer, Singapore; c2021. p. 123-141.
- Ali S, Khan AR, Mairaj G, Arif M, Fida M, Bibi S. Assessment of different crop nutrient management practices for yield improvement. *Aust. J Crop Sci.* 2008;2(3):150-157.
- Ambika S, Bhaskaran M, Manonmani V, Vanangamudi K. Storability of polymer coated CORH 3 hybrid rice seeds. *ORYZA- Int. J Rice.* 2014;51(2):125-130.
- Amir M, Singh CB, Jatav AL, Kumar M, Singh DP, Tripathi DK. Effect of seed priming technologies on planting value of lentil (*Lens culinaris* M) cultivars under sub-optimal condition. *The Pharma Innovation Journal.* 2021;10(12):673-678.
- Amir M, Singh CB, Jatav AL, Kumar M, Singh DP, Tripathi DK. Effect of seed enhancement techniques on yield and yield attributes of Lentil (*Lens culinaris* M.) cultivars under rainfed conditions. *SKUAST Journal of Research.* 2021;23(2):193-197.
- Amirkhani M, Netravali AN, Huang W, Taylor AG. Investigation of soy protein-based biostimulant seed coating for broccoli seedling and plant growth enhancement. *Hort. Sci.* 2016;51(9):1121-1126. <https://doi.org/10.21273/HORTSCI10913-16>.
- Baroni DF, Vieira HD. Coating seeds with fertilizer: a promising technique for forage crop seeds. *Ciência e Agrotecnologia.* 2020, 44. <https://doi.org/10.1590/1413-7054202044013720>
- Ben-Jabeur M, Adrian GR, Camilo LC, Rubén V, Zayneb K, Shawn CK, *et al.*, The promising Multispe Q device for tracing the effect of seed coating with biostimulants on growth promotion, photosynthetic state and water-nutrient stress tolerance in durum wheat. *Euro Mediterr. J. Environ. Integr.* 2021;6:1-11. <https://doi.org/10.1007/s41207-020-00213-8>.
- Berto B, Ritchie AL, Erickson TE. Seed-enhancement combinations improve germination and handling in two dominant native grass species. *Restor. Ecol.* 2021;29(1):13275. <https://doi.org/10.1111/rec.13275>.
- Bueno AN, Meyer TE, Anthony RJ. INCOTEC HOLDING BV, Seed coating composition. U.S. Patent Application 15/346,189. Biological Products Markets around the World; c2017. Available online: <http://www.bpia.org/wp-content/uploads/2018/03/Biological-Products-Markets-Around-The-World.pdf> (accessed on 30 October 2020).
- Babychan M. Efficacy of *Trichoderma* spp. against *Fusarium oxysporum* f. sp. lycopersici. (FOL) infecting pre-and post-seedling of tomato; c2017. <https://www.phytojournal.com/archives?year=2017&vol=6&issue=4&ArticleId=1405>.
- Buffington B, Beegle D, Lindholm C. Seed Treatment a National Pesticide Applicator Manual; Pesticide Educational Resources Collaborative (PERC), University of California Davis: Davis, CA, USA; c2018.
- Chandrika KP, Prasad RD, Godbole V. Development of chitosan-PEG blended films using *Trichoderma*: Enhancement of antimicrobial activity and seed quality. *Int. J. Biol. Macromol.* 2019;126:282-290. <https://doi.org/10.1016/j.ijbiomac.2018.12.208>.
- Chandrika KSVP, Singh A, Prasad RD, Yadav P. Prominence of seed coating for biotic and abiotic stresses. *Popular Kheti*; c2017. p. 1-3.
- Cho S, Seo H, Oh Y, Lee E, Choi I, Jang Y, *et al.*, Selection of coating materials and binders for pelleting onion (*Allium cepa* L.) seed. *Korean J. Hortic. Sci. Technol.* 2000;41(6):593-597.
- Chen Y, Turnblad KM. Insecticidal Seed Coating. U.S. Patent 0,177,526 A1, 28 November 2002.
- Colla G, Rouphael Y, Bonini P, Cardarelli M. Coating seeds with endophytic fungi enhances growth, nutrient uptake, yield and grain quality of winter wheat. *Int. J Plant Prod.* 2015;9(2):171-190.
- Costa DCA, Carvalho DLCRM, Medeiros DAF, Silva DLB, Romão MH, De Andrade Soares R, *et al.* Açai seed extract prevents the renin-angiotensin system activation, oxidative stress and inflammation in white adipose tissue of high-fat diet-fed mice. *Nutrition Research.* 2020;79:35-49. <https://doi.org/10.1016/j.nutres.2020.05.006>
- Dong Y, Jiang J. Improving seed germination and peanut yields by cold plasma treatment. *Plasma Science & Technology.* 2016;18(10):1027-1033. <https://doi.org/10.1088/1009-0630/18/10/10>
- Delin S, Engström L, Lundkvist A. Optimal placement of meat bone meal pellets to spring oats. *Front. Sustain. Food Syst.* 2018;2:27. <https://doi.org/10.3389/fsufs.2018.00027>
- Dimkpa CO, Andrews J, Fugice J, Singh U, Bindraban PS, Elmer WH, *et al.*, Facile coating of urea with low-dose ZnO nanoparticles promotes wheat performance and enhances Zn uptake under drought stress. *Front. Plant Sci.*

- 2020, 11. <https://doi.org/10.3389/fpls.2020.00168>.
28. Food and Agriculture Organization of the United Nations. Good Agricultural Practices for greenhouse vegetable crops: Principles for Mediterranean climate areas; c2014. Retrieved from <http://www.fao.org/3/i3382e/i3382e00.htm>
 29. Głodowska M, Schwinghamer T, Husk B, Smith D. Biochar based inoculants improve soybean growth and nodulation. *Agric. Sci.* 2017;8(9):1048-1064. <https://doi.org/10.4236/as.2017.89076>.
 30. Gorim L, Asch F. Effects of composition and share of seed coatings on the mobilization efficiency of cereal seeds during germination. *J Agron. Crop Sci.* 2012;198(2):81-91. <https://doi.org/10.1111/j.1439-037X.2011.00490.x>.
 31. Gorim L, Asch F. Seed coating increases seed moisture uptake and restricts embryonic oxygen availability in germinating cereal seeds. *Biol.* 2017;6(2):31. <https://doi.org/10.3390/biology6020031>.
 32. Govinden-Soulange J, Levantard M. Comparative studies of seed priming and pelleting on percentage and meantime to germination of seeds of tomato (*Lycopersicon esculentum* Mill.). *Afr. J Agric. Res.* 2008;3(10):725-731. <https://doi.org/10.5897/AJAR.9000117>.
 33. Guan Y, Song C, Gan Y, Li FM. Increased maize yield using slow-release attapulgit-coated fertilizers. *Agron. Sustain. Dev.* 2014;34(3):657-665. <https://doi.org/10.1007/s13593-013-0193-2>.
 34. Halmer P. Commercial seed treatment technology. In *Seed Technology and Its Biological Basis*; Black, M., Bewley, J. D., Eds.; Sheffield Academic Press: Sheffield, UK; c2000. p. 257-286.
 35. Halmer P. Seed technology and seed enhancement. In: XXVII International Horticultural Congress-IHC2006: International Symposium on Seed Enhancement and Seedling Production. 2006;771:17-26. <https://doi.org/10.17660/ActaHortic.2008.771.1>
 36. Hartley E, Gemell LG, Herridge DF. Lime pelleting inoculated serradella (*Ornithopus* spp.) increases nodulation and yield. *Soil Biol. Biochem.* 2004;36(8):1289-1294. <https://doi.org/10.1016/j.soilbio.2004.04.010>.
 37. Hayat R. Seed biopriming with plant growth promoting rhizobacteria. *FEMS Microbiology Ecology*, 2016, 92(8). <https://doi.org/10.1093/femsec/fiw112>
 38. Helal IM. Control of damping-off disease in some plants using environmentally safe biocides. *Pak. J Bot.* 2017;49(1):361-370.
 39. Hill HJ. Recent developments in seed technology. *J. New Seeds.* 1999;1:105-112.
 40. Hussain M, Mehboob N, Naveed M, Shehzadi K, Yasir TA. Optimizing boron seed coating level and boron-tolerant bacteria for improving yield and biofortification of chickpea. *J Plant. Nutr. Soil Sci.* 2020;20(4):2471-2478. <https://doi.org/10.1007/s42729-020-00313-y>.
 41. Javed T, Afzal I. Impact of seed pelleting on germination potential, seedling growth and storage of tomato seed. *Acta Hortic.* 2020;1273:417-424. <https://doi.org/10.17660/ActaHortic.2020.1273.54>.
 42. Javed T, Afzal I, Mauro RP. Seed coating in direct seeded rice: An innovative and sustainable approach to enhance grain yield and weed management under submerged conditions. *Sustainability.* 2021;13(4):2190. <https://doi.org/10.3390/su13042190>.
 43. Johnson SE, Lauren JG, Welch RM, Duxbury JM. A comparison of the effects of micronutrient seed priming and soil fertilization on the mineral nutrition of chickpea (*Cicer arietinum*), lentil (*Lens culinaris*), rice (*Oryza sativa*) and wheat (*Triticum aestivum*) in Nepal. *Exp. Agric.* 2005;41(4):427-448. <https://doi.org/10.1017/S0014479705002851>.
 44. Jeffries C. First report of 'Candidatus Liberibacter solanacearum' in parsley (*Petroselinum crispum*) seed. *New Disease Reports.* 2016;34(1):31. <https://doi.org/10.5197/j.2044-0588.2016.034.031>
 45. Kalaivani K, Kalaiselvi MM, Senthil-Nathan S. Effect of methyl salicylate (MeSA), an elicitor on growth, physiology and pathology of resistant and susceptible rice varieties. *Sci. Rep.* 2016;6:34498.
 46. Kanampiu F, Diallo A, Karaya H. Herbicide-seed coating technology: a unique approach for Striga control in maize. *Afr. Crop Sci. J.* 2015;8:1095-1098.
 47. Kanampiu FK, Kabambe V, Massawe C, Jasi L, Friesen D, Ransom JK, *et al.*, Multi-site, multi-season field tests demonstrate that herbicide seed coating herbicide-resistance maize controls *Striga* spp. and increases yields in several African countries. *Crop Prot.* 2003;22(5):697-706. [https://doi.org/10.1016/S0261-2194\(03\)00007-3](https://doi.org/10.1016/S0261-2194(03)00007-3)
 48. Kangsopa J, Hynes RK, Siri B. Lettuce seeds pelleting: A new bilayer matrix for lettuce (*Lactuca sativa*) seeds. *Seed Sci. Technol.* 2018;46:521-531.
 49. Kaufman G. Seed coating: a tool for stand establishment, a stimulus to seed quality. *Hort. Technol.* 1991;1(1):98-102. <https://doi.org/10.21273/HORTTECH.1.1.98>.
 50. Kimmelshue C, Goggi AS, Cademartiri R. The use of biological seed coatings based on bacteriophages and polymers against *Clavibacter michiganensis* subsp. *nebraskensis* in maize seeds. *Sci. Rep.* 2019;9:17950.
 51. Klessig DF, Manohar M, Baby S, Koch A, Danquah WB, Luna E, *et al.* Nematode ascarioside enhances resistance in a broad spectrum of plant-pathogen systems. *J. Phytopathol.* 2019;167:1-8.
 52. Kumar SV, Vyakaranahal B, Deshpande V, Raikar S, Nadaf H, Kumar BA. Effect of seed polymer coating on growth and yield of pigeonpea. *Karnataka J Agric. Sci.* 2014;27(4):469-471.
 53. Lamichhane JR, Dürr C, Schwanck AA, Robin MH, Sarthou JP, Cellier V, *et al.* Integrated management of damping-off diseases. A review. *Agron. Sustain. Dev.* 2017;37(2):10. <https://doi.org/10.1007/s13593-017-0417-y>.
 54. Lanka SK, Senthil-Nathan S, Blouin DJ, Stout MJ. Impact of thiamethoxam seed treatment on growth and yield of rice (*Oryza sativa*) *J Econ. Entomol.* 2017;110(2):479-486. <https://doi.org/10.1093/jee/tox043>.
 55. Latef AHA, He C. Effect of arbuscular mycorrhizal fungi on growth, mineral nutrition, antioxidant enzymes activity and fruit yield of tomato grown under salinity stress. *Scientia Horticulturae.* 2011;127(3):228-233. <https://doi.org/10.1016/j.scienta.2010.09.020>
 56. Lee MW, Huffaker A, Crippen D, Robbins RT, Goggin FL. Plant elicitor peptides promote plant defences against nematodes in soybean. *Mol. Plant Pathol.* 2018;19:858-869.
 57. Leetham D. The cement manufacturing process. Thermo Fisher Scientific; c2015. Retrieved: June 2019 from <https://www.thermofisher.com/blog/mining/thecement->

- manufacturing-process/
58. Lopisso DT, Kühlmann V, Siebold M. Potential of soil-derived fungal biocontrol agents applied as a soil amendment and a seed coating to control *Verticillium* wilt of sugar beet. *Biocontrol Sci. Technol.* 2017;27:1019-1037.
 59. Ma Y. Seed coating with beneficial microorganisms for precision agriculture. *Biotechnol. Adv.* 2019;37:107423.
 60. Madsen MD, Petersen S, Taylor AG. Seed Coating Compositions and Methods for Applying Soil Surfactants to Water-Repellent Soil. U.S. Patent, 9,554,502 B2, 31. Organic Materials Review Institute; c2017. Available online: <https://www.omri.org/>
 61. Maity A. Synthetic polymer based coating of fodder cowpea seeds enhances germination and vigour. *Uknowledge*; c2018. <https://uknowledge.uky.edu/igc/23/2-7-1/20/>.
 62. Mihuta-Grimm. *Trichoderma* spp. as biocontrol agents of *Rhizoctonia* damping-off of radish in organic soil and comparison of four delivery systems. *Disease control phytopathology journal*; c1986.
 63. Mohammad, Peksen. Seed coating technology: An innovative and sustainable approach for improving seed quality and crop performance. *Journal of the Saudi Society of Agricultural Sciences.* 2020;21(8):536-545. <https://doi.org/10.1016/j.jssas.2022.03.003>
 64. Nawar L. Pathological and rhizospheric studies on root-rot disease of squash in Saudi Arabia and its control; c2007. www.ajol.info. <https://doi.org/10.4314/ajb.v6i3.56140>
 65. Pedrini S, Balestrazzi A, Madsen M, Bhalsing K, Hardegree S, Dixon KW, *et al.* Seed enhancement: Getting seeds restoration-ready. *Restor. Ecol.* 2020;28:S266-S275.
 66. Porter FE, Scott JM. Seed coating methods and Purposes: A status report. In *Proceedings of the Short Course for Seedsmen; Mississippi Agricultural and Forestry Experiment Station: Prairie, MS, USA; c1979.*
 67. Piveta G, Menezes VO, Pedroso DC, Muniz MFB, Blume E, Wielewicky AP. Superação de dormência na qualidade de sementes e mudas: influência na produção de Senna multijuga (L. C. Rich.) Irwin & Barneby. *Acta Amazonica.* 2010;40(2):281-288. <https://doi.org/10.1590/s0044-59672010000200006>.
 68. Qiu Y, Amirkhani M, Mayton H, Chen Z, Taylor AG. Biostimulant seed coating treatments to improve cover crop germination and seedling growth. *Agronomy.* 2020;10(2):154. <https://doi.org/10.3390/agronomy10020154>
 69. Ramesh K, Thirumurugan V. Effect of seed pelleting and foliar nutrition on growth of soybean. *Madras Agric. J.* 2001;88(7):465-468.
 70. Rehman A, Farooq M. Zinc seed coating improves the growth, grain yield and grain biofortification of bread wheat. *Acta Physiol. Plant.* 2016;38(10):1-10. <https://doi.org/10.1007/s11738-016-2250-3>
 71. Rocha IDS, Ma Y, Souza-Alonso P, Vosátka M, Freitas H, Oliveira RS. Seed coating: A tool for delivering beneficial microbes to agricultural crops. *Front. Plant Sci.* 2019;10:1357.
 72. Ryu CM, Kim J, Choi O, Kim SH, Park CS. Improvement of biological control capacity of *Paenibacillus polymyxa* E681 by seed pelleting on sesame. *Biol. Control.* 2006;39:282-289.
 73. Scott JM. Seed coatings and treatments and their effects on plant establishment. *Adv. Agron.* 1989;42:43-83.
 74. Sikes BA. When do arbuscular mycorrhizal fungi protect plant roots from pathogens? *Plant Signaling Behav.* 2010;5:763-765. doi: <https://doi.org/10.4161/psb.5.6.11776>.
 75. Sim JB, Chung IM, Ku HM, Choi HW, Lee JM, Chun SC. Enhancing the biological control of rice seedling disease by adding specific carbon sources into the *Bacillus cereus* D324 formulation in water-seeded rice. *Plant Pathol. J.* 2008;24:58-62. doi: <https://doi.org/10.5423/PPJ.2008.24.1.058>.
 76. Singh A, Jain A, Sarma BK, Upadhyay RS, Singh HB. Rhizosphere competent microbial consortium mediates rapid changes in phenolic profiles in chickpea during *Sclerotium rolfsii* infection. *Microbiol. Res.* 2014;169:353-360. doi: <https://doi.org/10.1016/j.micres.2013.09.014>
 77. Singh V, Mawar R, Lodha S. Combined effects of biocontrol agents and soil amendments on soil microbial populations, plant growth and incidence of charcoal rot of cowpea and wilt of cumin. *Phytopathol. Mediterr.* 2012;51:307-316. doi: https://doi.org/10.14601/Phytopathol_Mediterr-9474
 78. Sivan A, Chet I. Biological control of *Fusarium* spp. in cotton, wheat and muskmelon by *Trichoderma harzianum*. *J Phytopathol.* 1986;116:39-47. doi: <https://doi.org/10.1111/j.1439-0434.1986.tb00892.x>
 79. Sivan A, Elad Y, Chet I. Biological control effects of a new isolate of *Trichoderma harzianum* on *Pythium aphanidermatum*. *Phytopathology.* 1984;74:498-501. doi: <https://doi.org/10.1094/Phyto-74-498>
 80. Sivan A, Ucko O, Chet I. Biological control of *Fusarium* crown rot of tomato by *Trichoderma harzianum* under field conditions. *Plant Dis.* 1987;71:587-592. doi: <https://doi.org/10.1094/PD-71-0587>
 81. Smith RS. Legume inoculant formulation and application. *Can. J Microbiol.* 1992;38:485-492. doi: <https://doi.org/10.1139/m92-080>
 82. Taylor AG. Seed storage, germination, quality and enhancements. In *The Physiology of Vegetable Crops*, 2nd ed.; Wien, H.C., Stutzel, H., Eds.; CAB International: Wallingford, UK; c2020. p. 1-30.
 83. Taylor AG, Allen PS, Bennett MA, Bradford KJ, Burris JS, Misra MK. Seed enhancements. *Seed Sci. Res.* 1998;8:245-256.
 84. Taylor AG, Trimmer M. Dunham Trimmer International Bio Intelligence, Lakewood Ranch, Florida. Global chemical and biological seed treatments market. Personal communication; c2020.
 85. Tayyab N, Naz R, Yasmin H, Nosheen A, Keyani R, Sajjad M, *et al.* Combined seed and foliar pre-treatments with exogenous methyl jasmonate and salicylic acid mitigate drought induced stress in maize. *PLoS ONE.* 2020;15:e0232269.
 86. Javed T, Afzal I, Shabbir R, Ikram K, Zaheer MS, Faheem M, *et al.* Seed coating technology: An innovative and sustainable approach for improving seed quality and crop performance, *Journal of the Saudi Society of Agricultural Sciences.* 2022;21(8):536-545, <https://doi.org/10.1016/j.jssas.2022.03.003>
 87. Hafeez M, Yuan C, Shahzad K, Aziz B, Iqbal K, Raza S. An empirical evaluation of financial development-carbon

- footprint nexus in One Belt and Road region. *Environmental Science and Pollution Research*. 2019 Aug 1;26:25026-25036.
88. Khan SA, Yu Z, Belhadi A, Mardani A. Investigating the effects of renewable energy on international trade and environmental quality. *Journal of Environmental management*. 2020 Oct 15;272:111089.
89. Pedrini S, Merritt DJ, Stevens J, Dixon K. Seed coating: science or marketing spin?. *Trends in plant science*. 2017 Feb 1;22(2):106-116.
90. Willenborg CJ, Gulden RH, Johnson EN, Shirtliffe SJ. Germination characteristics of polymer-coated canola (*Brassica napus* L.) seeds subjected to moisture stress at different temperatures. *Agronomy journal*. 2004 May;96(3):786-791.
91. Naveed M, Mitter B, Reichenauer TG, Wieczorek K, Sessitsch A. Increased drought stress resilience of maize through endophytic colonization by Burkholderia phytofirmans PsJN and Enterobacter sp. FD17. *Environmental and Experimental Botany*. 2014 Jan 1;97:30-39.
92. Olivera P, Danese S, Peyrin-Biroulet L. Next generation of small molecules in inflammatory bowel disease. *Gut*. 2017 Feb 1;66(2):199-209.
93. Zaheer H, Breyer Y, Dumay J. Digital entrepreneurship: An interdisciplinary structured literature review and research agenda. *Technological Forecasting and Social Change*. 2019 Nov 1;148:119735.
94. Zelonka. Effect and after-effect of barley seed coating with phosphorus on germination, photosynthetic pigments and grain yield *Acta Universitatis Latviensis, Biology*. 2005;691:111-119.
95. Sun J, Dai X, Wang Q, Van Loosdrecht MC, Ni BJ. Microplastics in wastewater treatment plants: Detection, occurrence and removal. *Water research*. 2019 Apr 1;152:21-37.
96. John OP, Robins RW, Pervin LA, editors. *Handbook of personality: Theory and research*. Guilford Press; 2010 Nov 24.
97. O'Callaghan M. Microbial inoculation of seed for improved crop performance: issues and opportunities. *Applied microbiology and biotechnology*. 2016 Jul;100(13):5729-5746.
98. Philippot L, Raaijmakers JM, Lemanceau P, Putten VDW. Going back to the roots: The microbial ecology of the rhizosphere. *Nature reviews microbiology*. 2013 Nov;11(11):789-799.
99. Hayat R, Ali S, Amara U, Khalid R, Ahmed I. Soil beneficial bacteria and their role in plant growth promotion: A review. *Annals of microbiology*. 2010 Dec;60:579-598.
100. Glick BR. Plant growth-promoting bacteria: mechanisms and applications. *Scientifica*. 2012 Oct 11; 2012.
101. Bulgarelli D, Schlaeppi K, Spaepen S, Themaat VEV, Schulze-Lefert P. Structure and functions of the bacterial microbiota of plants. *Annual review of plant biology*. 2013 Apr 29;64:807-838.