



ISSN (E): 2277-7695
ISSN (P): 2349-8242
NAAS Rating: 5.23
TPI 2023; 12(3): 1591-1594
© 2023 TPI
www.thepharmajournal.com
Received: 21-12-2022
Accepted: 27-02-2023

Gangadhar Kapase
Assistant Professor,
Department of Veterinary
Physiology and Biochemistry,
Veterinary College, Nandinagar,
Bidar, Karnataka, India

Shrikant Kulkarni
Professor and Head,
Department of Veterinary
Physiology and Biochemistry,
Veterinary College, Nandinagar,
Bidar, Karnataka, India

M Kiran
Assistant Professor, Department
of Livestock Products
Technology, Veterinary College,
Nandinagar, Bidar, Karnataka,
India

Srinivas Reddy Bellur
Assistant Professor, Department
of Veterinary Physiology and
Biochemistry, Veterinary
College, Nandinagar, Bidar,
Karnataka, India

Yogesh
Senior Veterinary Officer,
Department of Animal
Husbandry and Veterinary
Services, Bengaluru, Karnataka,
India

Sarita Patil
Assistant Professor,
Department of Veterinary
Physiology and Biochemistry,
Veterinary College, Nandinagar,
Bidar, Karnataka, India

Corresponding Author:
Gangadhar Kapase
Assistant Professor,
Department of Veterinary
Physiology and Biochemistry,
Veterinary College, Nandinagar,
Bidar, Karnataka, India

Profiling of physico-chemical and biochemical characteristics of chicken thigh (Iliotibialis) and breast (Pectoralis) Muscles

Gangadhar Kapase, Shrikant Kulkarni, MKiran, Srinivas Reddy Bellur, Yogesh and Sarita Patil

Abstract

The current study was carried to determine various physico-chemical and biochemical characteristics of Thigh (Iliotibialis) and Breast (Pectoralis) muscles of chicken. The pH was significantly ($p < 0.05$) higher in thigh muscle than breast muscle. R-value is indirect measure of ATP depletion indicates completion of rigor mortis. R-value, Myofibrillar Fragment Index (MFI) value and Muscle Fiber Diameter (MFD) value showed no significant ($p < 0.05$) difference between muscles of thigh and breast. Sarcoplasmic protein extractability was higher in thigh muscle whereas myofibrillar and total protein extractability were higher in breast muscle. TBARS and Myoglobin was significantly more in thigh muscles while % metmyoglobin was higher in breast muscle.

Keywords: Breast, R-value, pH, Myoglobin and MFI

Introduction

Short production cycle of poultry allows producers to respond quickly to market signals, while also allowing for rapid improvements in genetics, animal health, and feeding practices. Rapid expansion is also foreseen in Asia, led by China and India. Consumption of poultry meat increases regardless of region or income level. Per capita consumption will grow, even in the developed world, but growth rates will remain higher in developing regions. Among all the additional meat consumed over the next decade, poultry is expected to account for 44%. Meat imports into Asia account for 56% of global trade, and poultry will constitute more than half of this additional import demand (OECD-FAO, Agricultural Outlook 2018–2027) [17].

Chicken meat is considered as one of the most desirable meats all over the world. Chicken meat contains a high protein, low fat content and deliberated as the principal source of polyunsaturated fatty acids (PUFA) (Kamboh and Zhu, 2013) [14]. Meat quality is primarily important to consumers, and the demand for high-quality meat has increased globally (Joo, Kim, Hwang, & Ryu, 2013) [17]. Consumers, with increasing health consciousness, are becoming more aware of the nutritional value of the foods they eat. Over the last several decades, the proportion of poultry marketed as whole intact carcasses has declined due to increased consumer demands for cut-up carcass parts and further-processed poultry products (Bowker, 2017) [5]. The relationship between quality and eating experience of different cuts within the same carcass has gained significant attention (Huang *et al* 2018) [15]. Since consumer demand for specific meat cuts is shifting to smaller sized animals because of smaller household sizes (Fowler *et al.*, 2018) [12]. Marketing of chicken parts is one of the fastest growing segments of the food industry around the world. Special interest in this area is partially due to the convenience and nutritional value of cuts such as chicken breasts and thighs (Seabra *et al.*, 2001) [19].

Materials and Methods

The chicken meat required for the study was sourced from local slaughter house, Bidar, Karnataka, India, under chilled conditions. The boneless muscles from the thigh and breast region were cut into uniform sized pieces and packed under atmospheric conditions using low-density polyethylene bags kept at 4 ± 1 C in a domestic refrigerator.

Fresh meat samples were obtained in batches separately for each of the three replications ($n = 3$). The raw meat pieces were evaluated for pH, R- value, water holding capacity (WHC), protein extractability, muscle fiber diameter (MFD), myofibrillar fragmentation index (MFI), Myoglobin and % metmyoglobin.

pH value was measured using a digital pH meter homogenizer (Model: Z742486, Bench mark, D1000 Hand held homogenizer, Malaysia) Kiran *et al.* (2019) [20]. R-value was measured by using the method reported by Honikel and Fischer (1977) [13]. For determining water-holding capacity, centrifugal method of estimation was used (Wardlaw *et al.*, 1973) [34]. Myofibrillar fragmentation index was determined as per the procedure outlined by Davis *et al.* (1980) [8]. The MFI was reported as the weight of the residue in grams times one hundred. Muscle fibre diameter was determined as described by Tuma *et al.* (1962) [33], using calibrated micrometer. Protein extractability was determined according to procedure of Joo *et al.* (1999) [16]. The modified extraction method of Witte *et al.* (1970) [37] was followed to estimate TBARS. Myoglobin was extracted from raw pieces using a modified procedure of Warris (1979) [36]. The myoglobin concentration and % Met myoglobin were calculated according to Trout (1989) [32].

Results and Discussion

The change in physicochemical properties as influenced by muscle type is presented in Table 1. pH value of breast muscle was 5.67 which is significantly lower than leg muscle 6.03. The pH of muscle/meat is a measurement of acidity. In a normal living muscle the pH is approximately 7.2. Glycogen is broken down to lactic acid when muscle turns into meat. The highest quality products tend to fall in the pH range of 5.7 to 6.0. The pH may also affect the oxidation susceptibility of meat. The more acidic a meat is, the greater the risk of oxidation (Baeza, 2020) [4]. Farouk and Lovatt (2000) [11], who reported that white muscle fibres have a higher rate of glycolytic change than do red fibres and thus have a faster rate of pH decline. Lesiak *et al.* (1996) [21] reported in poultry meat, pH of leg muscle was higher than that of breast muscle about 0.2-0.3 unit. The pH of breast meat was found significantly ($p < 0.05$) lower than meat from leg and back portion in emu Raut *et al.* 2017) [29]. Choe *et al.* (2010) [7], stated pH of broiler thigh muscle was higher than that of breast muscle.

The R-value is used to observe development of rigor which indicates the degree of conversion of adenosine to inosine nucleotides (Papa and Fletcher, 1988). The R-value is an indirect measure of ATP degradation in the muscle (Mckee and Sams, 1998) [24], a high number means a relatively low ATP content (Honikel and Hamm, 1978) [14]. In the present experiment, R-value of the thigh and breast muscle was 1.31 and 1.36 respectively. The R-values of the breast and leg muscles were not significantly different ($p > 0.05$). The R-values of the breast and leg muscles were greater than 1.2 after 6 h PM, which indicated that rigor mortis was complete (Yu *et al.* 2011) [40]. Young and Lyon (1997) [39] suggested that 1.35 was the ultimate R-value. Song *et al.* (2020) [31] observed that the R-value of pre-rigor chicken breasts salted with KCl (1.41) was significantly higher than those salted with NaCl (1.35).

Table 1: Profile on physicochemical property of poultry meat

Parameters	Thigh	Breast
pH*	6.03±0.02 ^a	5.67±0.05
R-VALUE	1.31±0.02	1.36±0.04
WHC (%)*	26.63±0.59	23.36±0.49
MFI	71.65±1.89	71.20±1.32
MFD (μ)	60.33±1.05	62.60±1.58
SPE (mg/g)	82.85±0.85	78.61±0.47
MFPE (mg/g)	152.29±1.74	164.10±1.79
TPE (mg/g)	235.15±1.23	245.95±2.10
TBARS	0.36±0.02	0.25±0.01
Myoglobin (mg/g)	3.33±0.18	1.55±0.07
Metmyoglobin (%)	57.03±1.63	67.97±1.95

The WHC was influenced by the chicken muscle location, with a higher ($p < 0.05$) WHC in thigh samples with higher pH values than in breast samples with lower pH values. The current results were accorded with that of Kadioglu *et al.* (2019) [18] who found that thigh meat with relatively high pH values shows a higher WHC than that of breast meat with relatively low pH values. By contrast, Botka-Petrak *et al.* (2005) [5], reported that white muscle (M. Pectoralis superficialis) has a greater water holding capacity than does red muscle (M. iliotibialis lateralis). Farouk *et al.*, 2012 [10], hypothesized that the increase in WHC with ageing is due to the breakdown in meat structure and the creation of “sponge effect”, which disrupts the channels through which moisture is lost and physically entraps the free water in meat and reduce the amount that drips out.

The myofibril fragmentation index is the process of breaking myofibrils into smaller segments at the Z-line or nearby during the animal post-mortem. Myofibrillar fragmentation index (MFI) and shear force values can be used to express meat tenderization chiefly caused by the proteolysis of myofibrillar protein (Kim *et al.*, 2013; Olson *et al.*, 1976). MFI increases continuously during aging of meat from ruminant animals, for example, but, to our knowledge, there are no standards established for MFI in chicken meat, which makes it difficult to rank the degree of tenderness using MFI. In current study MFI and MFD showed no significant difference between thigh and breast muscles. Mello *et al.*, 2017 [25] showed that during aging of breast meat, a reduction of MFI values also occurred.

MFD: There is no significant difference between breast and thigh muscles. A smaller diameter of muscle fibers is beneficial for meat juiciness, currently leg muscles showed lesser diameter than the breast muscle. The average muscle fiber diameter seen in the current study was similar to those noticed for chicken (Naveena and Mendiratta 2001) [26], and emu (Naveena *et al.* 2015) [27].

Protein extractability: Sarcoplasmic protein extractability was found higher in thigh muscle where as myofibrillar protein extractability was found to be higher in breast muscle. Total protein extractability was significantly more in breast muscle than the thigh muscle. Increased total protein extractability/solubility is due to degradation of myofibrillar proteins causing instability in the structure of myofibrils significant contributor to the variations of chicken quality in the early postmortem period (Warner *et al.* 2022) [35]. Xiong and Brekke (1991) [38] reported that post-rigor breast myofibrils showed a greater protein extractability and gel

strength than prerigor breast myofibrils, but the reverse was found for leg myofibrils. Salt-soluble protein was least extractable at pH 5.50 for both breast and leg myofibrils.

TBARS: The generally processes of cooking and storage heighten lipid peroxidation in poultry meat (Eder *et al.* 2005)^[9]. Lipid oxidation and protein oxidation are the chief causes of meat deterioration, which affects the nutritional value, physicochemical properties and shelf life of meat (Zhang *et al.* 2013)^[42]; Adeyemi *et al.* 2017)^[1]. Chicken thigh muscle showed significantly higher TBARS value than breast muscle. Al-Kelabi *et al.* 2020^[3] also reported lower TBARS values for breast than the thigh muscles. TBARS of 0.9 mg MDA/kg as a standard maximum acceptable limit for chicken meat. The TBARS values depends on the concentration of oxygen in the atmosphere, lesser the oxygen level lower will be the oxidation rate (Mao *et al.* 2023)^[23].

Myoglobin and Met myoglobin: The percentage of the presences of myoglobin, met myoglobin and oxymyoglobin significantly different in different samples. (Al-Husseiny and Khrebish, 2019)^[2]. Oxidation of oxymyoglobin produces brown discoloration as a result of Met myoglobin formation (Naveena *et al.* 2015)^[27]. The breast muscle showed lesser myoglobin content and more in metmyoglobin content and results were visa versa in thigh muscles. In beef, oxidation metabolism caused by the overabundance of proteins related to the TCA cycle and degradation of complexes in the electronic respiratory chain affects the reduction ability of MetMyoglobin (Yu *et al.* 2017)^[41].

To anticipate and respond to projected increases in the global demand of poultry, we must understand, in detail, those mechanisms responsible for optimizing meat quality. Current findings highlights difference in quality of meat between two major sources like breast and thigh. This clearly demonstrates need for muscle based strategy for better utilization of meat quality from poultry.

References

- Adeyemi KD, Shittu RM, Sabow AB, Karim R, Sazili AQ. Myofibrillar protein, lipid and myoglobin oxidation, antioxidant profile, physicochemical and sensory properties of caprine longissimus thoracis during postmortem conditioning. *Journal of Food Processing and Preservation.* 2017;41(4):e13076.
- Al-Husseiny KS, Khrebish MT. Determination of Chemical Content and some Physical Properties and Meat Pigments (Myoglobin, Meta Myoglobin and Oxymyoglobin) in Different Parts of Slaughtered Animals. *Basrah J Agric. Sci.* 2019;32:302-319.
- Al-Kelabi TJK, Mohamed MF, Al-Karagoly H. The impact of sweet basil plant additive on biochemical parameters and organoleptic quality of meat of chicken. *Annals of Tropical Medicine & Public Health;* c2020.
- Baeza E. Factors Influencing the Processing Ability and Oxidation Susceptibility of Poultry Meat. *J Nutr Food Sci* 3: 012. Henry Publishing Groups Baeza E. 2020;3(1):100012.
- Botka-Petrak K, Vidaček S, Petrak T, Medić H. Influence of combined, continuous chilling on physical and chemical properties of white and red chicken muscles. *Veterinarski arhiv.* 2005 Oct 20;75(5):415-22.
- Bowker B. Developments in our understanding of water-holding capacity. In *Poultry Quality Evaluation.* Woodhead Publishing; c2017.p. 77-113.
- Choe JH, Nam KC, Jung S, Kim BN, Yun HJ, Jo CR. Differences in the quality characteristics between commercial Korean native chickens and broilers. *Food Science of Animal Resources.* 2010;30(1):13-9.
- Davis GW, Dutton TR, Smith GC, Carpenter ZL. Fragmentation procedure for bovine longissimus muscle as an index of cooked steak tenderness. *Journal of Food Science.* 1980;45(4):880-884.
- Eder K, Grünthal G, Kluge H, Hirche F, Spilke J, Brandsch C. Concentrations of cholesterol oxidation products in raw, heat-processed and frozen-stored meat of broiler chickens fed diets differing in the type of fat and vitamin E concentrations. *British Journal of Nutrition.* 2005;93(5):633-643.
- Farouk MM, Mustafa NM, Wu G, Krsinic G. The sponge effect hypothesis: An alternative explanation of the improvement in the water holding capacity of meat with ageing. *Meat Science.* 2012;90(3):670-677.
- Farouk MM, Lovatt SJ. Initial chilling rate of pre-rigor beef muscles as an indicator of colour of thawed meat. *Meat Science.* 2000 Oct 1;56(2):139-44.
- Fowler SM, Hoban JM, Melville G, Pethick DW, Morris S, Hopkins DL. Maintaining the appeal of Australian lamb to the modern consumer. *Animal Production Science.* 2018;58(8):1392-1398.
- Honikel KO, Fischer C. A rapid method for the detection of PSE and DFD porcine muscles. *Journal of Food Science.* 1977;42(6):1633-1636.
- Honikel KO, Hamm R. Measurement of water-holding capacity and juiciness. Quality attributes and their measurement in meat, poultry and fish products; c1994. p. 125-61.
- Huang S, Bohrer BM, Chalupa-Krebdak S, Wang LM, Mejia SV. Correlation of chicken breast (Pectoralis major) quality and sensory attributes with chicken thigh quality and sensory attributes. *Meat and Muscle Biology,* 2018, 2(2).
- Joo ST, Kauffman RG, Kim BC, Park GB. The relationship of sarcoplasmic and myofibrillar protein solubility to colour and water-holding capacity in porcine longissimus muscle. *Meat science.* 1999;52(3):291-297.
- Joo ST, Kim GD, Hwang YH, Ryu YC. Control of fresh meat quality through manipulation of muscle fiber characteristics. *Meat science.* 2013;95(4):828-836.
- Kadioğlu P, Karakaya M, Unal K, Babaoğlu AS. Technological and textural properties of spent chicken breast, drumstick and thigh meats as affected by marinating with pineapple fruit juice. *British poultry science.* 2019 Jul 4;60(4):381-7.
- Kamboh AA, Zhu WY. Effect of increasing levels of bioflavonoids in broiler feed on plasma anti-oxidative potential, lipid metabolites, and fatty acid composition of meat. *Poultry Science.* 2013;92(2):454-461.
- Kiran M, Naveena BM, Smrutirekha M, Reddy PB, Rituparna B, Kumar YP, *et al.* Traditional halal slaughter without stunning versus slaughter with electrical stunning of sheep (*Ovis aries*). *Meat science.* 2019;148:127-136.
- Lesiak MT, Olson DG, Lesiak CA, Ahn DU. Effects of postmortem temperature and time on the water-holding capacity of hot-boned turkey breast and thigh muscle. *Meat Science.* 1996 May 1;43(1):51-60.
- Ma Y, Wang Y, Wang Z, Chen B, Xie Y, Kong L, *et al.* Mechanisms of chicken processing quality changes

- during the early postmortem time: the role of the changes of myofibrillar protein. *International Journal of Food Science & Technology*. 2023 January.
23. Mao Y, Yang S, Zhang Y, Luo X, Niu L, Holman BW. High-pressure processing and modified atmosphere packaging combinations for the improvement of dark, firm, and dry beef quality and shelf-life. *Meat Science*. 2023 Jan 14:109113.
 24. McKee SR, Sams AR. Rigor mortis development at elevated temperatures induces pale exudative turkey meat characteristics. *Poultry Science*. 1998 Jan 1;77(1):169-74.
 25. Mello JL, Souza RA, Paschoalin GC, Ferrari FB, Machado BM, Giampietro-Ganeco A, Souza PA, Borba H. A comparison of the effects of post-mortem aging on breast meat from Cobb 500 and Hubbard ISA broilers. *Animal Production Science*. 2017 May 22;58(10):1922-31.
 26. Naveena BM, Mendiratta SK. Tenderisation of spent hen meat using ginger extract. *British poultry science*. 2001 Jul 1;42(3):344-9.
 27. Naveena BM, Muthukumar M, Kulkarni VV, Praveen Kumar Y, Usha Rani K, Kiran M. Effect of aging on the Physicochemical, Textural, Microbial and Proteome Changes in Emu (*Dromaius novaehollandiae*) Meat Under Different Packaging Conditions. *Journal of Food Processing and Preservation*. 2015;39(6):2497-2506.
 28. Organisation for Economic Co-operation and Development. OECD-FAO agricultural outlook 2018-2027. OECD Publishing; c2018.
 29. Raut SS, Pathera AK, Sharma DP, Yadav S, Singh PK. Physico-chemical and Instrumental Colour Properties of Emu Meat. *Journal of Animal Research*. 2017;7(3):591-595.
 30. Seabra LM, Zapata JF, Fuentes MF, Aguiar CM, Freitas ER, Rodrigues MC. Effect of deboning time, muscle tensioning, and calcium chloride marination on texture characteristics of chicken breast meat. *Poultry Science*. 2001;80(1):109-112.
 31. Song DH, Ham YK, Ha JH, Kim YR, Chin KB, Kim HW. Impacts of pre-rigor salting with KCL on technological properties of ground chicken breast. *Poultry Science*. 2020;99(1):597-603.
 32. Trout GR. Variation in myoglobin denaturation and color of cooked beef, pork, and turkey meat as influenced by pH, sodium chloride, sodium tripolyphosphate, and cooking temperature. *Journal of Food Science*. 1989;54(3):536-540.
 33. Tuma HJ, Venable JH, Wuthier PR, Henrickson RL. Relationship of fiber diameter to tenderness and meatiness as influenced by bovine age. *Journal of Animal Science*. 1962;21(1):33-36.
 34. Wardlaw FB, McCaskill LH, Acton JC. Effect of postmortem muscle changes on poultry meat loaf properties. *Journal of Food science*. 1973 Mar;38(3):421-3.
 35. Warner RD, Wheeler TL, Ha M, Li X, Bekhit AE, Morton J, *et al.* Meat tenderness: Advances in biology, biochemistry, molecular mechanisms and new technologies. *Meat Science*. 2022 Mar 1;185:108657.
 36. Warris PD. The extraction of haem pigments from fresh meat. *International Journal of Food Science & Technology*. 1979;14(1):75-80.
 37. WITTE VC, Krause GF, Bailey ME. A new extraction method for determining 2-thiobarbituric acid values of pork and beef during storage. *Journal of food Science*. 1970;35(5):582-585.
 38. Xiong YL, Brekke CJ. Protein extractability and thermally induced gelation properties of myofibrils isolated from pre-and post-rigor chicken muscles. *Journal of Food Science*. 1991;56(1):210-215.
 39. Young LL, Lyon CE. Effect of calcium marination on biochemical and textural properties of peri-rigor chicken breast meat. *Poultry Sci*. 1997;76:197-201.
 40. Yu LH, Lee ES, Chen HS, Jeong JY, Choi YS, Lim DG, *et al.* Comparison of physicochemical characteristics of hot-boned chicken breast and leg muscles during storage at 20°C. *Korean J. Food Sci*. 2011;31(5):676-683.
 41. Yu Q, Wu W, Tian X, Jia F, Xu L, Dai R, Li X. Comparative Proteomics to Reveal Muscle-Specific Beef Color Stability of Holstein Cattle during Post-Mortem Storage. *Food Chem*. 2017;229:769-778. DOI: 10.1016/j.foodchem.2017.03.004.
 42. Zhang W, Xiao S, Ahn DU. Protein oxidation: basic principles and implications for meat quality. *Critical Reviews in Food Science and Nutrition*. 2013;53(11):1191-1201.