www.ThePharmaJournal.com

# The Pharma Innovation



ISSN (E): 2277-7695 ISSN (P): 2349-8242 NAAS Rating: 5.23 TPI 2023; 12(5): 1762-1765 © 2023 TPI www.thepharmajournal.com

Received: 02-03-2023 Accepted: 05-04-2023

### Karthik HN

Department of Forest Products and Utilization, College of Forestry, University of Agricultural Sciences, Dharwad, Karnataka. India

#### Ramana P

Department of Forest Products and Utilization, College of Forestry, Sirsi, University of Agricultural Sciences, Dharwad, Karnataka, India

**Corresponding Author:** Karthik HN

**Department of Forest Products** and Utilization, College of Forestry, University of Agricultural Sciences, Dharwad, Karnataka, India

### Organic acid profile of Garcinia gummi-gutta (L) Rob. from central Western Ghats of India

### Karthik HN and Ramana P

### Abstract

Forests are a rich reserve of compounds that can be used as pharmaceuticals and nutraceuticals. The genus Garcinia, which belongs to the Clusiaceae family, has about 200 species that are found throughout the world's tropics, mostly in Asia, Africa, and Polynesia Garcinia gummi-gutta (L) Rob. is the most commercially exploited for organic acids with its vast economic importance worldwide. Hydroxycitric acid (HCA) is known to have excellent anti-obesity property. Field study was conducted to determine the effect of different locations and its site factors on organic acid content of G. gummi-gutta fruits in 9 different sites of Uttara Kannada, Karnataka. A statistically significant influence of sites conditions on the organic acid content. The Organic acids were found higher in site  $S_1$  (with altitude 646.3 m MSL, high precipitation and average temperature of 24 °C), the highest of 7.85±0.23% (-)-Hydroxycitric acid lactone (HCA-L) with the average 7.15±0.57% and 21.52±1.28% of HCA with the average of  $18.47 \pm 1.50\%$  was found respectively. The HCA and HCA-L had very strong positive correlation (r = 0.84 to 0.98) with site factors (except mean annual temperature).

Keywords: Garcinia gummi-gutta, organic acid profile, locations, HPLC, central Western Ghats, Karnataka

### **1. Introduction**

Garcinia gummi-gutta (L) Rob. (syn. Garcinia cambogia). It belongs to family Clusiaceae, it is also known as Uppage (Kannada) locally. It is an evergreen tree of small to medium size, grows up to 12 m tall with leaves of shining dark green colour. The flowering season lasts from January to March, while fruit production begins in June and July. Fruits usually grow into ovoid shape with the diameter of 5 cm, ripe fruits are yellow, having 6 to 8 grooves and about 6 to 8 seeds covered by succulent aril (Singh, 1993)<sup>[20]</sup>.

Distribution ranges between southwards of Konkan to eastwards of Travancore in Western Ghats. Apart from home range it's been introduced in subtropical Malaysia, China and Philippines (Chuah et al., 2013)<sup>[3]</sup>. According to Lewis and Neelekantan (1965)<sup>[14]</sup>, hydroxycitric acid (HCA) is a key organic acid that predominates in the fruit rinds of G. gummi-gutta. HCA helps to reduce obesity by suppressing appetite (Preuss et al., 2004; Kovacs et al., 2006)<sup>[19, 12]</sup> and enhancing energy outlay (Leonhardt & Langhans, 2002)<sup>[13]</sup>. The fleshy fruit rind of the tree is the richest natural source of potential anti-obesity plant metabolite (-)-Hydroxycitric acid (HCA) along with minute quantity of citric, tartaric, and malic acids (Lewis & Neelekantan, 1965; Jayaprakash & Sakaraiah, 2000)<sup>[14, 8]</sup>.

Lab rats (Ohia et al., 2001)<sup>[17]</sup> and other experimental animals with slower weight gain and less body fat accumulation have also been recorded after HCA usage (Kim et al., 2008; Amin et al., 2011; Kim et al., 2008)<sup>[10, 1, 11]</sup>. HCA has the property to lower blood lipids such as cholesterol and triglycerides. Additionally, there have been no reported negative consequences from human usage (Maia-Landim et al., 2021) [15]. There are wide variations in naturally distributed G. gummi-gutta populations. The present study was aimed to understand the organic acid chemical diversity existing among G gummi-gutta growing in Uttara Kannada district of Karnataka.

### 2. Material and Methods

### 2.1 Study area

The study was conducted in Uttara Kannada district of Karnataka, a part of Central Western Ghats. Nine sites were randomly selected, in each site five trees were chosen as replications. Trees of 50-80cm girth class were selected to maintain the uniformity among the samples. The rainfall in these sites ranged between 2300 to 4722 mm per year and the altitude ranges

between 418.7 to 646.3 m (Table 1).

 
 Table 1: Geographical and climatic information of study area in Uttara Kannada district, Karnataka

Site	Altitude (m)	Mean annual rainfall (mm)	Annual rainy days	Mean annual temperature (°C)
$S_1$	646.3	3800 - 4722	120	24
$S_2$	610.7	3800 - 4722	120	23
<b>S</b> <sub>3</sub>	623.3	3800 - 4722	120	24
$S_4$	516.2	2950 - 3667	100	25
<b>S</b> 5	508.9	2950 - 3667	100	25
<b>S</b> <sub>6</sub>	522.7	2950 - 3667	100	25
<b>S</b> <sub>7</sub>	460.8	2300 - 3263	95	27
<b>S</b> <sub>8</sub>	437.8	2300 - 3263	95	27
<b>S</b> 9	418.7	2300 - 3263	95	27

# **2.2 Estimation of Organic acid profile (High Performance Liquid Chromatography)**

Analysis was carried out using High Performance Liquid Chromatography (HPLC-Shimadzu chromatographic system -LC 8A) instrument.

### 2.2.1 Preparation of sample for analysis

Under forced air ventilation, the fruit rinds were dried in oven at 45 °C for 8 hours. The samples' dried rind was extracted with Soxhlet water extraction mentod by boiling 10 g of it in 50 ml of double-distilled water for 45 minutes at 100 °C. After the extract was dried, the residue was combined with 50 ml of 30% orthophosphoric acid.

## 2.2.2 Standards for High performance liquid chromatography

Citric acid, D-(+)-Glucuronolactone, and the ethylenediamine salt of HCA were purchased. The ethylene-diamine salt of HCA in 5 mg was dissolved in 5 mL of 50% H<sub>2</sub>SO<sub>4</sub>, which was then added to 10 mL of HPLC grade water to make the desired volume. A stock solution of 500 g/mL of HCA was created. By dissolving the lactone (D-(+)-Glucuronolactone) salts in HPLC grade water, lactone standards were created. Salts of citric acid were also dissolved in water of HPLC quality. The lactone and citric acid standard solvents were kept at a 500 g/mL concentration. (Jayaprakash and Sakariah, 2002)<sup>[9]</sup>.

### 2.2.3 Analysis using High performance liquid chromatography

A dual pump, photodiode array detector (SPDM10AVP), and LC10 software-equipped HPLC-Shimadzu Class chromatographic system (LC 8A model) were used to filter 200 µL of the standard samples through a 0.45-inch filter. Twenty litres of each were then fed into the system. Results from HPLC were obtained using a Waters Cortecs reversed phase C-18, 2.7 µm with sperical solid core, 4.6 mm 150 mm column, and a gradient elution solvent with 0.2 M sodium sulphate and a pH of 2.5 that was adjusted using diluted H<sub>2</sub>SO<sub>4</sub>. The reading was obtained using ultraviolet (UV) detection at 215 nm while the flow rate of 0.5 mL/min was maintained. To achieve accuracy in evaluation, various trial sets were inserted.

Based on the area obtained, the amounts were calculated using the lab solution software Shimadzu Analytical (India) Pvt. Ltd. Quantitative analysis was obtained by measuring the peak responses data of standards to that of the testing extracts. The values from two trials were recorded, and the average mean was obtained as the final amount (Ong, 2004; Nour *et al.*, 2013) <sup>[18, 16]</sup>.

### 2.3 Data analysis

The data were subjected to one factor ANOVA using both 'SPSS' and 'R-software', additionally 'Tukey's range test' was done, and 'Compact Letter Display' (CLD) was assigned to each variable. Further data visualizations were done using "ggplot2" package in R-software.

### 3. Results and discussion

### 3.1 Organic acid profile of fruit rind

The organic acids viz. (-)-Hydroxycitric acid lactone (HCA-L), Hydroxycitric acid (HCA), and Citric acid were estimated from dried rind samples (Figure 1) using HPLC. Percent share of these organic acids were presented in Table 2. The mean HCA of 18.47±1.90%, HCA-L of 7.15±0.57% and Citric acid of 0.13±0.04% were found respectively. The highest (-)-Hydroxycitric acid lactone (HCA-L) was found in site S<sub>3</sub>  $(7.94\pm0.43\% \text{ w/w})$  and has no statistically significant difference between site S<sub>1</sub>, maximum Hydroxycitric acid (HCA) in site  $S_1$  (21.52±1.28% w/w) and Citric acid content in S<sub>2</sub> (0.18 $\pm$ 0.03% w/w) was observed. In similar findings the range of HCA was at 10 to 30 percent (Anju & Rameshkumar, 2017; Lewis and Neelekantan, 1965; Devi et al., 2012)<sup>[2, 14, 5]</sup>, which in turn agrees with our results with an overall average HCA of 18.47±1.90%. However, There's been a report of up to 42 to 44 percent of HCA from G. gummi-gutta rind (Edirisinghe et al., 2015)<sup>[7]</sup>, which is way more than what we found from our study.

The observations indicated a higher percent presence of major organic acids (HCA and HCA-L) in site  $S_1$  of altitude 646.3 with lower temperature, high rainfall, and total rainy days compared to other study sites. A very strong positive correlation was observed between HCA and HCA-L with site factors (*viz.* altitude, mean annual rainfall and annual rainy days) except mean annual temperature, where strong negative correlation was observed in case of HCA. With respect to fruit and rind parameters, only the rind thickness had strong positive correlation with HCA and HCA-L (Figure 2).

The environmental factors might have some kind of influence on chemical composition. The process of metabolism and secondary metabolite accumulation will be greatly influenced by important environmental factors like altitude, temperature, incident light, humidity, and precipitation. These environmental differences in different locations directly contribute to the difference and variation in active ingredient contents (Dong *et al.*, 2011; Wang *et al.*, 2014)<sup>[6, 21]</sup>.

Hydroxycitric acid and citric acid is formed from carbohydrate in Garcinia fruit tissue and suggest that their initial accumulation and subsequent disappearance are related to the changes in the activity of the condensing enzyme and other enzymes of the citric acid cycle. However young fruits have more citric acid and mature fruits have more HCA. The decrease in citric acid in the ripe fruit suggests the possibility that this stage may perhaps be characterized by the presence of an enzyme system that affects the partial breakdown of citric acid (Deshpande & Ramakrishnan, 1960)<sup>[4]</sup>.

The different forms of organic acids such as HCA-L, HCA, and Citric acid may be due to the process of concentration along with evaporation in Garcinia species leading to lactonization of regular - HCA to HCA-L. HCA-L (lactone) is way less biologically active (Jayaprakash & Sakariah, 2002)<sup>[9]</sup>.

	Organic acids (in percentage)							
Site	Hydroxycitric Acid (@ p<0.05)	Retention time (min)	Hydroxycitric acid lactone (@ p<0.05)	Retention time (min)	Citric Acid (@ <i>p</i> <0.05)	Retention time (min)		
$S_1$	21.52±1.28 <sup>a</sup>	5.263	7.85±0.23 <sup>a</sup>	4.690	0.13±0.05 ab	10.652		
$S_2$	20.61±1.04 <sup>a</sup>	5.268	7.23±0.29 <sup>ab</sup>	4.694	0.18±0.03 <sup>a</sup>	10.650		
<b>S</b> <sub>3</sub>	20.45±1.20 <sup>a</sup>	5.258	7.94±0.43 <sup>a</sup>	4.689	0.11±0.03 b	10.641		
<b>S</b> 4	17.61±0.59 <sup>b</sup>	5.264	6.94±0.60 <sup>b</sup>	4.691	0.15±0.02 ab	10.653		
<b>S</b> 5	17.38±0.43 <sup>b</sup>	5.266	7.24±0.41 <sup>ab</sup>	4.697	0.12±0.03 b	10.651		
S6	17.94±0.44 <sup>b</sup>	5.260	6.76±0.36 <sup>b</sup>	4.695	0.13±0.02 ab	10.657		
<b>S</b> <sub>7</sub>	16.58±0.35 <sup>b</sup>	5.257	6.72±0.41 <sup>b</sup>	4.689	0.1±0.02 b	10.649		
S <sub>8</sub>	16.89±0.41 <sup>b</sup>	5.259	6.94±0.38 <sup>b</sup>	4.686	0.09±0.02 b	10.654		
<b>S</b> 9	17.23±0.31 <sup>b</sup>	5.267	6.75±0.42 b	4.689	0.14±0.02 ab	10.656		
Mean	18.47±1.90		7.15±0.57		0.13±0.04			

Table 2: Percent (w/w) Organic acid content of Garcinia gummi-guttafruit rind



Fig 1: Overview of typical Garcinia fruit rind (Fresh and dried)

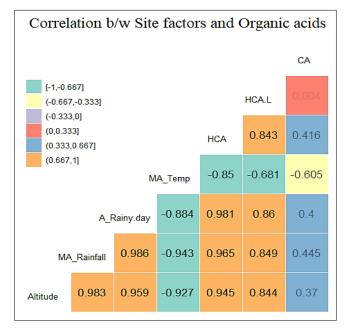


Fig 2: Correlation matrix of Organic acids with Site factors

### Conclusion

The environmental factors might have influence on organic acid profile of *Garcinia gummi-gutta*. HCA is the predominant commercially important organic acid observed through HPLC. The site  $(S_1)$  with high altitude and precipitation with lower temperature found to have higher percent availability of major organic acid content within the fruit rind. The HCA and HCA-L had very strong positive correlation with site factors (except mean annual temperature). The different environmental conditions might have a direct impact on organic acid composition within the plant in case of our study. Further study is needed to determine particular reason causing this phenomenon.

### References

effect of Garcinia against renal oxidative stress and biomarkers induced by high fat and sucrose diet. Lipids in health and disease. 2011;10(1):1-13.

- Anju V, Rameshkumar KB. Phytochemicals and bioactivities of *Garcinia gummi-gutta* (L.) N. Robson- A review. In K. B. Rameshkumar, (Eds.), Diversity of Garcinia species in the Western Ghats: Phytochemical perspective. Jawaharlal Nehru Tropical Botanic Garden and Research Institute; c2017. p. 151-161.
- 3. Chuah LO, Ho WY, Beh BK, Yeap SK. Updates on Antiobesity effect of *Garcinia* origin (–)-HCA. Evidence-Based Complementary and Alternative Medicine. 2013 Jan 1;2013.
- Deshpande WM, Ramakrishnan CV. Formation and breakdown of Citric acid in Garcinia fruit. Journal of Biological Chemistry. 1960;236:2377-2380.
- Devi SP, Balamohan TN, Thangam M, Kumar JA, Ramachandrudu K, Korikanthimath VS. A study on diversity and distribution of kokum (*Garcinia indica* (Choisy) Thouars) using DIVA-GIS in Goa with respect to fruit characters. Indian Journal of Horticulture. 2012;69(2):156-62.
- 6. Dong J, Ma X, Wei Q, Peng S, Zhang S. Effects of growing location on the contents of secondary metabolites in the leaves of four selected superior clones of *Eucommia ulmoides*. Industrial Crops and Products. 2011;34(3):1607-14.
- 7. Edirisinghe EA, Nawarathna SB, Marapana R, Jaysinghe J. Extraction, Crystallization, Preservtion, Pelletizing and Quantification of Hydroxy Citric Acid from *Garcinia cambogia*. International Journal of Innovative Research in Technology. 2015;2(4):3-10.
- 8. Jayaprakash GK. Sakaraiah KK. Determination of (-) Hydroxycitric acid in commercial samples of *Garcinia cambogia* extract by liquid chromatography with ultraviolet detection. Journal of Liquid Chromatography and Related Technologies. 2000;23(6):915-923.
- Jayaprakash GK, Sakariah KK. Determination of organic acids in leaves and rinds of *Garcinia indica* (Desr.) by LC. Journal of pharmaceutical and biomedical analysis. 2002;28(2):379-384.
- Kim KY, Lee HN, Kim YJ, Park T. Garcinia cambogia extract ameliorates visceral adiposity in C57BL/6J mice fed on a high-fat diet. Bioscience, biotechnology, and biochemistry. 2008;72(7):1772-80.
- 11. Kim YJ, Kim KY, Kim MS, Lee JH, Lee KP, Park T. A mixture of the aqueous extract of *Garcinia cambogia*, soy peptide and L-carnitine reduces the accumulation of visceral fat mass in rats rendered obese by a high fat diet.

<sup>1.</sup> Amin KA, Kamel HH, Abd Eltawab MA. Protective

Genes & nutrition. 2008;2(4):353-8.

- 12. Kovacs EM, Westerterp-Plantenga MS. Effects of (-)hydroxycitrate on net fat synthesis as de novo lipogenesis. Physiology & behavior. 2006;88(4-5):371-81.
- 13. Leonhardt M, Langhans W. Hydroxycitrate has long-term effects on feeding behavior, body weight regain and metabolism after body weight loss in male rats. The Journal of nutrition. 2002;132(7):1977-82.
- 14. Lewis YS, Neelakantan S. (-)-Hydroxycitric acid—the principal acid in the fruits of *Garcinia cambogia*desr. Phytochemistry. 1965;4(4):619-25.
- 15. Maia-Landim A, Lancho C, Poblador MS, Lancho JL, Ramírez JM. *Garcinia cambogia* and Glucomannan reduce weight, change body composition and ameliorate lipid and glucose blood profiles in overweight/obese patients. Journal of Herbal Medicine. 2021;26:100424.
- Nour V, Trandafir I, Cosmulescu S. HPLC determination of phenolic acids, flavonoids and juglone in walnut leaves. Journal of chromatographic science. 2013;51(9):883-90.
- Ohia SE, Awe SO, LeDay AM, Opere CA, Bagchi D. Effect of hydroxycitric acid on serotonin release from isolated rat brain cortex. Research Communications in Molecular Pathology and Pharmacology. 2001 Mar 1;109(3-4):210-6.
- 18. Ong ES. Extraction methods and chemical standardization of botanicals and herbal preparations. Journal of Chromatography B. 2004;812(1-2):23-33.
- 19. Preuss HG, Bagchi D, Bagchi M, Rao CS, Dey DK, Satyanarayana S. Effects of a natural extract of (–)hydroxycitric acid (HCA-SX) and a combination of HCA-SX plus niacin-bound chromium and *Gymnema sylvestre* extract on weight loss. Diabetes, Obesity and Metabolism. 2004;6(3):171-80.
- 20. Singh NP. Clusiaceae (Guttiferae nom. alt.) In: Sharma, BD and Balakrishnan NP (eds.), Flora of India 3. Botanical Survey of India, Kolkatta. 1993, 86-151.
- 21. Wang D, He F, Lv Z, Li D. Phytochemical composition, antioxidant activity and HPLC fingerprinting profiles of three Pyrola species from different regions. PLoS One. 2014;9(5):e96329.