



ISSN (E): 2277-7695
ISSN (P): 2349-8242
NAAS Rating: 5.23
TPI 2023; 12(6): 4833-4837
© 2023 TPI
www.thepharmajournal.com
Received: 13-03-2023
Accepted: 16-04-2023

Gaytri Soni
Department of Agricultural
Chemistry and Soil Science,
College of Agriculture, Junagadh
Agricultural University,
Junagadh, Gujarat, India

Dr. SG Savalia
Department of Agricultural
Chemistry and Soil Science,
College of Agriculture, Junagadh
Agricultural University,
Junagadh, Gujarat, India

Effect of simulated soil salinity conditions and varieties of pigeon pea (*Cajanus cajan* L.) on nutrient content, salt compositions and their ratios

Gaytri Soni and Dr. SG Savalia

Abstract

In this study, we investigated the effects of simulated saline soil on the accumulation of Na, K and Ca at 45 DAS and harvesting in shoot part is investigated in this study. The experiment was conducted at net house and the experiment constitutes five salinity levels (Control, 40, 60, 80 and 100 meq l⁻¹) and four pigeon pea varieties (V₁: GJP-1, V₂: Vaishali, V₃: BDN-2, V₄: AGT-2) in CRD (Factorial) replicated three times. The sodium concentration in leaves of pigeon pea observed lowest and highest potassium and calcium at 45 DAS with variety V₄ (AGT-2). The lowest Na⁺/K⁺ ratio and highest Ca⁺⁺/Na⁺ ratio were observed significant with variety V₄ (AGT-2) in leaves at 45 DAS and at harvest. Primary, secondary and macro nutrients were not affected by interaction of salinity levels and varieties.

Keywords: Pigeon pea, salinity levels, Na, K and Ca composition, Na⁺/K⁺ and Ca⁺⁺/Na⁺ ratio

Introduction

Salinity is widespread around the world and is usually a cause for concern because it affects plants negatively. Saline soils are common on Earth. Their formation can be due to natural or accelerated processes. Despite chloride being important as a micronutrient for all higher plants and sodium being important as a mineral nutrient for many halophytes and some C4 species, salt accumulation turns agricultural land into a hostile environment, reduce local biodiversity, restrict plant growth and reproduction, and are toxic in non-salt-tolerant plants, known as glycophytes (Marschner, 1995) [22]. Soil salinity is shown to increase P, Mn and Zn and drop in K and Fe (Turan *et al.*, 2007a) [26] concentrations in plants. Use of saline irrigation water and fertilizers are the main factors responsible for adding salinity in soil (Epstein *et al.*, 1980) [9]. Meanwhile, climate change is causing problems for agricultural production. Increased salinity hazards are associated with climate change impacts, especially in areas below sea level. Even in inland areas, including arid and semi-arid areas, salinity stress increasingly threatens crop production as salt builds up as a result of over-irrigation with poor quality water without proper drainage.

Materials and Methods

The experiment soil was silty clayey in texture and alkaline in reaction with pH 8.08, EC 0.30 dS m⁻¹, CaCO₃ 33.00% and CEC 35.20 cmol (p⁺) kg⁻¹. This trial consists of 20 treatments combinations, including five levels of salinity and four levels of varieties under the Factorial CRD design. The soil was low in available nitrogen (180.10 kg ha⁻¹) and medium in phosphorus (48.00 kg ha⁻¹), high in available potassium (407.00 kg ha⁻¹), and low in available sulphur (21.38 mg kg⁻¹). Dried samples were utilized for determination of calcium by Versenate EDTA method (Cheng and Bray, 1951) [6] and potassium & sodium by Flame photometer (Jackson, 1974) [15]. Na⁺/K⁺ and Ca⁺⁺/Na⁺ ratios were worked out by using values of content of potassium, sodium and calcium.

Chemical analysis of seed and straw were performed by taking representative samples from each pot at harvest. The samples were oven-dried at 60 °C for 24 hours and then ground with a pestle and mortar. Finally, the powdered samples were used for estimation of nitrogen, phosphorus, potassium, calcium, magnesium, sulphur, sodium, iron, zinc, manganese and copper. The nitrogen from plant samples were estimated separately by micro Kjeldahl's method as described by Kanwar and Chopra (1967) [17]. The phosphorus determined by Vanadomolybdo Phosphoric Yellow Colour method as described by Jackson (1974) [15]. The potassium and sodium by Flame Photometer as described by Jackson (1974) [15].

Corresponding Author:
Gaytri Soni
Department of Agricultural
Chemistry and Soil Science,
College of Agriculture, Junagadh
Agricultural University,
Junagadh, Gujarat, India

Calcium and Magnesium content were determined by the method developed by Versenate EDTA method. Sulphur content was determined by the method developed by Williams and Steinberg's (1959) [31]. Sodium was determined by flame photometer method developed by Jackson (1974) [15]. Micronutrients viz., Fe, Zn, Mn and Cu were estimated by Atomic Absorption Spectrophotometer (AAS) as described by Jackson (1967) [14].

Results

1. Effect of salinity and varieties on salt (Na, K and Ca) composition at 45 DAS

Sodium content increased significantly with increasing levels of salt concentration. Significantly lower value of Na content was registered with salt concentration level S₁ (Control). K and Ca content in leaves of pigeon pea decreased with increasing level of salt stress and significantly the highest values were noted with salt concentration level S₁.

The concentration of Na in leaves of pigeon pea at 45 DAS was significantly affected by tested varieties and the lowest content of Na recorded with variety V₄ (AGT-2) and the highest content was found in V₁ (GJP-1). Significantly the highest K and Ca content observed with variety V₄ (AGT-2). Interaction effect of salinity and variety was obtained significant in K and Ca at 45 DAS. The maximum potassium content (1.88%) was perceived with treatment combination of S₁ (Control) x V₄ (AGT-2). The maximum calcium content (1.65%) was perceived with treatment combination of S₁ (Control) x V₄ (AGT-2).

2. Effect of salinity and varieties on nutrient content at harvest (in seed and straw)

The concentration of phosphorus, potassium and sulphur content in seed and straw of the plant did not significantly influence by different salinity levels at harvest of crop. While nitrogen content in seed and straw significantly affected by different salinity levels and the highest nitrogen content was found in S₁ (Control) level. At harvest of crop, the Mg content in seed and straw was significantly affected by different levels of salinity. Whereas Ca content was significantly affected by salinity levels in seed only. At harvest of crop, the concentration of Na in seed and straw of the plant were significantly influenced by different levels of saline irrigation water. The Na concentration was significantly higher found with salinity level S₅ (100 meq l⁻¹) in seed (0.101) and straw (0.099), respectively. The concentration of Mn and Zn in seed and straw of the plant did not significantly affected by different levels of saline irrigation water at harvest of crop. Whereas, Fe (seed) and Cu (seed and straw) were significantly affected by different salinity levels.

The concentration of N, P, K, S Ca, Mn and in seed and straw of the plant were not significantly influenced by different varieties of pigeon pea. Whereas, Mg and Na content were significantly affected in seed and straw of various varieties at harvest of crop. The Na content was observed the lowest with variety AGT-2 (V₄) in seed at harvest of pigeon pea crop. Fe and Cu in straw and seed respectively were not found significant. While Fe in seed and Cu in straw, were significantly influenced by different varieties of pigeon pea at harvest of crop.

Table 1: Effect of salinity levels and varieties on concentration (%) of Na, K and Ca content in leaves of pigeon pea at 45 DAS

Treatments	Na	K	Ca
Salt concentration(Salinity) (S)			
S ₁ : Control	0.058	1.68	1.39
S ₂ : 40 meq l ⁻¹	0.061	1.53	1.25
S ₃ : 60 meq l ⁻¹	0.064	1.49	1.23
S ₄ : 80 meq l ⁻¹	0.067	1.46	1.21
S ₅ : 100 meq l ⁻¹	0.073	1.39	1.14
S.Em. ±	0.001	0.02	0.02
C.D. (P=0.05)	0.004	0.06	0.05
Variety (V)			
V ₁ :GJP-1	0.071	1.48	1.18
V ₂ : Vaishali	0.064	1.49	1.22
V ₃ :BDN-2	0.066	1.50	1.27
V ₄ : AGT-2	0.057	1.57	1.29
S.Em. ±	0.001	0.02	0.01
C.D. (P=0.05)	0.003	0.05	0.04
S x V Interaction			
S.Em. ±	0.003	0.04	0.03
C.D. (P=0.05)	NS	0.12	0.09
C.V.%	7.26	4.90	4.49

Table 2: Interaction effect of salinity levels and varieties on concentration (%) of K content in leaves of pigeon pea at 45 DAS

	S ₁ : Control	S ₂ : 40 meq l ⁻¹	S ₃ : 60 meq l ⁻¹	S ₄ : 80meq l ⁻¹	S ₅ : 100 meq l ⁻¹	Mean
V ₁ :GJP-1	1.66	1.49	1.47	1.44	1.32	1.48
V ₂ : Vaishali	1.55	1.53	1.49	1.47	1.42	1.49
V ₃ :BDN-2	1.62	1.50	1.48	1.45	1.42	1.50
V ₄ : AGT-2	1.88	1.59	1.51	1.47	1.39	1.57
Mean	1.68	1.53	1.49	1.46	1.39	
S.Em. ±	0.04		C.D. (P=0.05)		0.12	

Table 3: Interaction effect of salinity levels and varieties on concentration (%) of Ca content in leaves of pigeon pea at 45 DAS

	S ₁ : Control	S ₂ : 40 meq l ⁻¹	S ₃ : 60 meq l ⁻¹	S ₄ : 80 meq l ⁻¹	S ₅ : 100 meq l ⁻¹	Mean
V ₁ :GJP-1	1.30	1.21	1.19	1.17	1.05	1.18
V ₂ : Vaishali	1.30	1.25	1.22	1.19	1.15	1.22
V ₃ :BDN-2	1.31	1.29	1.28	1.25	1.24	1.27
V ₄ : AGT-2	1.65	1.25	1.23	1.22	1.12	1.29
Mean	1.39	1.25	1.23	1.21	1.14	
S.Em. ±	0.03		C.D. (P=0.05)		0.092	

The interaction effect of salinity levels and varieties for all these nutrients were found non-significant.

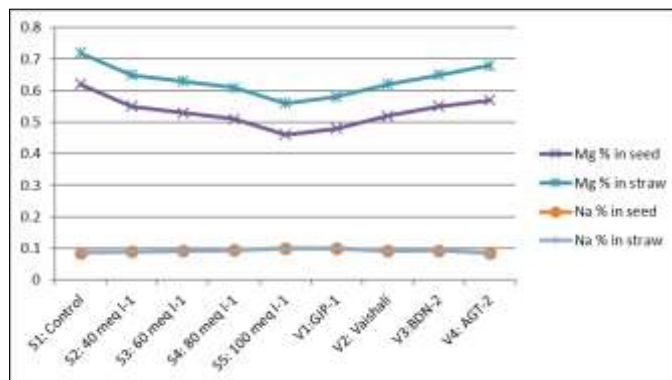


Fig 1: Effect of salinity levels and varieties on Mg and Na (%) content in seed and straw pigeon pea at harvest

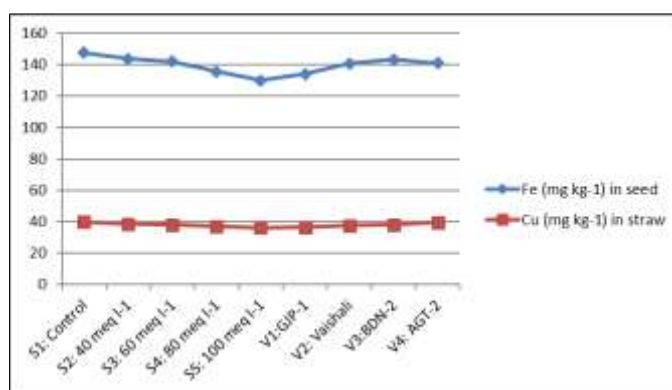


Fig 2: Effect of salinity levels and varieties on micronutrient (mg kg⁻¹) content in seed and straw pigeon pea at harvest

3. Na⁺/K⁺ and Ca⁺⁺/Na⁺ Ratio

3.1 Na⁺/K⁺ and Ca⁺⁺/Na⁺ ratio in leaves at 45 DAS

Various salinity levels produced significant result on Na⁺/K⁺ ratio and Ca⁺⁺/Na⁺ ratio. The lowest value of Na⁺/K⁺ ratio (0.033) recorded under S₁ (Control) in leaves of pigeon pea at 45 DAS. While Ca⁺⁺/Na⁺ ratio found significant by different varieties and salinity levels. The Highest Ca⁺⁺/Na⁺ ratio (24.27) observed at S₁ (Control) level of irrigation in pigeon pea leaves at 45 DAS.

Different varieties of pigeon pea tested in experiment are produce significant effect on both Na⁺/K⁺ and Ca⁺⁺/Na⁺ ratio at 45 DAS. Significantly the lowest value of Na⁺/K⁺ ratio (0.037) and the highest value of Ca⁺⁺/Na⁺ ratio (23.27) were recorded with the same variety V₄ (AGT-2).

Significant effect was observed in context to combined effect of salinity levels and varieties on Na⁺/K⁺ and Ca⁺⁺/Na⁺. The minimum value of Na⁺/K⁺ ratio (0.024) and the highest value of Ca⁺⁺/Na⁺ (32.93) were found in salt concentration level S₁ (Control) in variety V₄ (AGT-2).

3.2 Na⁺/K⁺ and Ca⁺⁺/Na⁺ ratio in Seed and Straw at harvest

The results data depicted that increased trend was noted with respect to Na⁺/K⁺ ratio with increasing salt concentrations. On other hand, Ca⁺⁺/Na⁺ ratio was vice-versa of Na⁺/K⁺ ratio. At the same level of salinity S₁ (Control), the lowest value of Na⁺/K⁺ ratio in seed (0.067) and straw (0.211). Whereas, the highest value of Ca⁺⁺/Na⁺ ratio in seed (16.35) and straw (40.18) were recorded at salt concentration level of S₁ *i.e.* Control. Ca⁺⁺/Na⁺ ratio in straw remains statistically at par with S₁ (40 meq l⁻¹).

Different varieties of pigeon pea significantly influenced Na⁺/K⁺ ratio and Ca⁺⁺/Na⁺ ratio in seed and straw of pigeon pea at harvest. The significantly lower value of Na⁺/K⁺ ratio was (0.068 and 0.201) and significantly the highest of Ca⁺⁺/Na⁺ ratio was (15.45 and 40.88) in seed and straw were observed with variety V₄ (AGT-2), respectively.

The combined effect of salinity levels and varieties was found non-significant on Na⁺/K⁺ and Ca⁺⁺/Na⁺ ratio at harvest in seed and straw of pigeon pea.

Table 4: Effect of salinity levels and varieties on Na⁺/K⁺ and Ca⁺⁺/Na⁺ ratio in leaves of pigeon pea at 45 DAS

Treatments	Na ⁺ /K ⁺	Ca ⁺⁺ /Na ⁺
Salt concentration (Salinity) (S)		
S ₁ : Control	0.033	24.27
S ₂ : 40 meq l ⁻¹	0.041	20.82
S ₃ : 60 meq l ⁻¹	0.043	19.44
S ₄ : 80 meq l ⁻¹	0.046	18.35
S ₅ : 100 meq l ⁻¹	0.052	15.83
S.Em. ±	0.001	0.32
C.D. (P=0.05)	0.002	0.92
Variety (V)		
V ₁ : GJP-1	0.049	16.87
V ₂ : Vaishali	0.043	19.25
V ₃ :BDN-2	0.042	19.57
V ₄ : AGT-2	0.037	23.27
S.Em. ±	0.001	0.29
C.D. (P=0.05)	0.002	0.82
S x V Interaction		
S.Em. ±	0.002	0.64
C.D. (P=0.05)	0.005	1.84
C.V.%	6.61	5.64

Table 5: Interaction effect of salinity levels and varieties on Na⁺/K⁺ ratio in leaves of pigeon pea at 45 DAS

	S ₁ : Control	S ₂ : 40 meq l ⁻¹	S ₃ : 60 meq l ⁻¹	S ₄ : 80meq l ⁻¹	S ₅ : 100 meq l ⁻¹	Mean
V ₁ : GJP-1	0.039	0.045	0.047	0.052	0.061	0.049
V ₂ : Vaishali	0.040	0.041	0.043	0.045	0.049	0.043
V ₃ : BDN-2	0.028	0.043	0.044	0.046	0.052	0.042
V ₄ : AGT-2	0.024	0.033	0.039	0.041	0.047	0.037
Mean	0.033	0.041	0.043	0.046	0.052	
S.Em. ±	0.002		C.D. (P=0.05)		0.004	

Table 6: Interaction effect of salinity levels and varieties on Ca⁺⁺/Na⁺ ratio in leaves of pigeon pea at 45 DAS

	S ₁ : Control	S ₂ : 40 meq l ⁻¹	S ₃ : 60 meq l ⁻¹	S ₄ : 80meq l ⁻¹	S ₅ : 100 meq l ⁻¹	Mean
V ₁ : GJP-1	20.26	18.06	17.25	15.81	12.96	16.87
V ₂ : Vaishali	21.67	20.56	19.27	18.35	16.43	19.25
V ₃ : BDN-2	22.20	20.38	19.69	18.89	16.71	19.57
V ₄ : AGT-2	32.93	24.28	21.54	20.34	17.23	23.27
Mean	24.27	20.82	19.44	18.35	15.83	
S.Em. ±	0.64		C.D. (P=0.05)		1.84	

Table 7: Effect of salinity levels and varieties on Na⁺/K⁺ and Ca⁺⁺/Na⁺ ratio in seed and straw of pigeon pea at harvest

Treatments	Na ⁺ /K ⁺		Ca ⁺⁺ /Na ⁺	
	Seed	Straw	Seed	Straw
Salinity (Salinity) (S)				
S ₁ : Control	0.067	0.211	16.35	40.18
S ₂ : 40 meq l ⁻¹	0.070	0.213	15.01	38.57
S ₃ : 60 meq l ⁻¹	0.072	0.220	14.37	37.22
S ₄ : 80 meq l ⁻¹	0.075	0.230	13.75	36.07
S ₅ : 100 meq l ⁻¹	0.081	0.253	12.44	34.37
S.Em. ±	0.001	0.006	0.31	0.63
C.D. (P=0.05)	0.003	0.016	0.90	1.79
Variety (V)				
V ₁ : GJP-1	0.078	0.254	12.93	33.55
V ₂ : Vaishali	0.073	0.222	14.26	37.14
V ₃ :BDN-2	0.074	0.225	14.90	37.55
V ₄ : AGT-2	0.068	0.201	15.45	40.88
S.Em. ±	0.001	0.005	0.28	0.56
C.D. (P=0.05)	0.003	0.014	0.80	1.60
S x V Interaction				
S.Em. ±	0.002	0.011	0.63	1.25
C.D. (P=0.05)	NS	NS	NS	NS
C.V.%	4.60	8.56	7.57	5.81

Discussion

Sodium is not considered an essential element for plants and plants accumulate Na⁺ at the expense of Ca⁺⁺ and K⁺ under saline conditions (Kuiper, 1984) [19]. According to Ioneva (1988) [13], the increased in Na⁺ contents, decreased in K⁺ contents and K⁺/Na⁺ ratios in plant leaves are attributed to the effects of competition between Na⁺ and K⁺ ions at absorption sites in plant roots. Mahmood *et al.* (2008) [21] also reported that with the addition of salt in the medium, Na content of shoot increased. Patel *et al.* (2010) [23] reported that Na⁺ increases with increasing salinity. According to Greenway and Munns (1980) [10], lower K⁺ concentrations may inhibit growth by reducing the capability to regulate osmotic pressure and maintain turgor pressure, and may negatively affect metabolic function. Mahmood *et al.* (2008) [21] also reported that K content dropped with the addition of salt to the medium. Essa (2002) reported that the main response of the plant to salt stress is a change in Ca⁺⁺ homeostasis and

attributed that the salt tolerance of plants is their ability to avoid Na⁺ toxicity and to maintain Ca⁺⁺ and K⁺ concentrations.

Similar results were found in studies done by Kurban *et al.* (1999) [20], Brown *et al.* (2006) [4], Waheed *et al.* (2006) [30], Alam *et al.* (2004) [1] and Turan *et al.* (2007b) [27] for N content. This result is consistent with Chakrabarti and Mukherji (2002) [5], who found that K decreased with increasing salinity. Turan *et al.* (2009) [28] reported that the application of NaCl to soil reduced the K content of plants. The decrease in K is thought to be due to its antagonistic relationship with Na. These investigators reported that high salt (NaCl) intake competes with the intake of other nutrient ions, especially K⁺, leading to depletion of K⁺ and other ions, as reported by Shokri and Maadi (2009) [25]. Antagonistic relation between Na and K may reduce the K concentration. This result is consistent with Chakrabarti and Mukherji (2002) [5] that Mg decreases with increasing salinity. As reported by Turan *et al.* (2009) [28], increasing NaCl levels increased Na uptake in plants. Na⁺ is also thought to maintain turgor, but cannot replace certain functions of Ca⁺⁺ and K⁺ (Patel *et al.* 2010) [23].

These results were supported by the study of Brown *et al.* (2006) [4], Turan *et al.* (2007b) [27], Hasan and Miyake (2017) [12] and Rodda (2018) [24] for Fe content. Similar result was found in the studies of Rodda (2018) [24] for Mn and Zn content.

The K⁺/Na⁺ ratios in plant leaves is attributed to the effect of competition between Na⁺ and K⁺ ions at the absorption sites in plant roots. As Patel *et al.* (2010) [23] reported increasing soil salinity decreased the K⁺/Na⁺ ratio. The result is in agreement with the studies of Waheed *et al.* (2006) [30] and Kholovo *et al.* (2010) [18].

High salt accumulation can disrupt the normal balance of plant physiological processes and lead to starvation. These results are in consistent with work of Essa (2002) [8], Dogan (2011) [7] and Bazrafshan and Ehsanzadeh (2014) [3]. These results are consistent with those reported by Jat *et al.* (2011) [16], Uddin *et al.* (2011) [29], Babu *et al.* (2012) [2] and Hakim *et al.* (2014) [11] for Na⁺/K⁺ and Ca⁺⁺/Na⁺ ratios at harvest in seed and straw.

Conclusion

Overall, variety AGT-2 showed lower values of Na⁺/K⁺ ratio and higher Ca⁺⁺/Na⁺ ratios in leaves at 45 DAS as well as in seed and straw at harvest. Therefore, AGT-2 was shown to be resistant to salt stress. The K⁺/Na⁺ ratio decreased in the straw and seeds of the tested cultivars, whereas the Na content increased under saline conditions. Higher Ca⁺⁺/Na⁺ ratios at higher EC values indicated greater tolerance to salinity than lower Ca⁺⁺/Na⁺ ratios at lower levels of EC.

Reference

1. Alam MZ, Stuchbury T, Naylor REL, Rashid MA. Effect of salinity on growth of some modern rice cultivars. *Journal of Agronomy*. 2004;3(1):1-10.
2. Babu MA, Singh D, Gothandam KM. The effect of salinity on growth, hormones and mineral elements in leaf and fruit of tomato cultivar PKM1. *Journal of Animals and Plant Science*. 2012;22(1):159-164.
3. Bazrafshan AH, Ehsanzadeh P. Growth, photosynthesis and ion balance of sesame (*Sesamum indicum* L.) genotypes in response to NaCl concentration in

- hydroponics solutions. *Photosynthetica*. 2014;52(1):134-147.
4. Brown CE, Pezeshki SR, Delaune RD. Effects of salinity and soil drying on nutrient uptake and growth of *Spartina alterniflora* in a simulated tidal system. *Environmental and Experimental Botany*. 2006;58(3):140-148.
 5. Chakrabarti N, Mukherji S. Growth regulator mediated changes in leaf area and metabolic activity in mung bean under salt stress condition. *Indian Journal Plant Physiology*. 2002;7(3):256-263.
 6. Cheng KL, Bray RH. Determination of calcium and magnesium in soil and plant material. *Indian Society of Soil Science*. 1951;72:449-458.
 7. Dogan M. Antioxidative and proline potentials as a protective mechanism in soybean plants under salinity stress. *African Journal of Biotechnology*. 2011;10(32):5972-5978.
 8. Essa TA. Effect of soil salinity on yield and quality of soybean. *Journal of Agronomy and Crop Science*. 2002;188(1):86-93.
 9. Epstein E, Norlyn JD, Rush DW, Kinsbury RW, Kelly DB, Cunningham GA, *et al.*, Saline culture of crops: A genetic approach. *Science*. 1980;210:399-404.
 10. Greenway H, Munns R. Mechanisms of salt tolerance in nonhalophytes. *Annual Review of Plant Physiology*. 1980;31:149-190.
 11. Hakim MA, Juraimi AS, Hanafi MM, Ismail MR, Rafii MY, Islam MM, *et al.* The effect of salinity on growth, ion accumulation and yield of rice varieties. *Journal of Animal and Plant Science*. 2014;24(3):874-885.
 12. Hasan R, Miyake H. Salinity stress alters nutrient uptake and causes the damage of root and leaf anatomy in maize. *International Conference on Biological Science*. 2017;3(4):219-225.
 13. Ioneva ZS. Effect of potassium ion Na⁺ uptake by plants in conditions of chloride salinity. *Biological Reviews*. 1988;14:42-47.
 14. Jackson ML. *Soil Chemical Analysis*. Prentice Hall of India Pvt. Ltd., New Delhi. 1967. p. 327-350.
 15. Jackson ML. *Soil Chemical Analysis*. Prentice Hall of India Pvt. Ltd., New Delhi. 1974.
 16. Jat SR. Phosphorus requirement of chickpea (*Cicer arietinum* L.) irrigated with chloride and sulphate dominated saline water. M.Sc. (Ag.) Thesis, SKRAU, Bikaner. 2011.
 17. Kanwar JS, Chopra SL. *Analytical agricultural chemistry*. Kalyani publishers, New Delhi, 1967. p. 518.
 18. Kholova J, Sairam RK, Meena RC. Osmolytes and metal ions accumulation, oxidative stress and antioxidant enzymes activity as determinants of salinity stress tolerance in maize genotypes. *Acta Physiologiae Plantarum*. 2010;32(3):477-486.
 19. Kuiper PJC. *Salinity Tolerance in Plants*. Wiley-Interscience, New York. 1984. p. 77-91.
 20. Kurban H, Saneoka H, Nehira K, Adilla R, Premachandra GS, Fujita K. Effect of salinity on growth, photosynthesis and mineral composition in leguminous plant *Alhagi pseudoalhagi* (Bieb.). *Soil science and plant nutrition*. 1999;45(4):851-862.
 21. Mahmood A, Athar M, Qadri R, Mahmood N. Effect of NaCl salinity on growth, nodulation and total nitrogen content in *Sesbania sesban*. *Agriculturae Conspectus Scientificus*. 2008;73(3):137-141.
 22. Marschner H. *Mineral nutrition of higher plants*. 2nd ed., p. 889. Academic Press, London, New York; c1995.
 23. Patel P, Kajal S, Patel VR, Patel VJ, Khristi S. Impact of salt stress on nutrient uptake and growth of cowpea. *Brazilian Society of Plant Physiology*. 2010;22(1):43-48.
 24. Rodda CD. Soil properties and yield of wheat (*Triticum aestivum* L.) as influenced by use of waters having variable salinity and sodicity. M.Sc. thesis, (Unpublished). Department of Agriculture Chemistry and Soil Science, College of Agriculture, JAU, Junagadh. 2018.
 25. Shokri S, Maadi B. Effects of arbuscular mycorrhizal fungus on the mineral nutrition and yield of *Trifolium alexandrinum* plants under salinity stress. *Journal of Agronomy*. 2009;8(2):79-83.
 26. Turan MA, Türkmen N, Taban N. Effect of NaCl on stomatal resistance and proline, chlorophyll, Na, Cl and K concentrations of lentil plants. *Journal of Agronomy*. 2007a;6:378-381.
 27. Turan MA, Katkat V, Taban S. Variations in proline, chlorophyll and mineral elements contents of wheat plants grown under salinity stress. *Journal of Agronomy*. 2007b;6(1):137.
 28. Turan M, Elkarim A, Taban N, Taban S. Effect of salt stress on growth, stomatal resistance, proline and chlorophyll concentrations on maize plant. *African Journal of Agricultural Research*. 2009;4(9):893-897.
 29. Uddin MK, Juraimi AS, Ismail MR, Hossain MA, Othman R, Rahim AA. Effect of salinity stress on nutrient uptake and chlorophyll content of tropical turfgrass species. *Australian Journal of Crop Science*. 2011;5(6):620.
 30. Waheed Abdul, Hafiz IA, Qadir Ghulam, Murtaza Ghulam, Mahmood T, Ashraf Muhammad. Effect of salinity on germination, growth, yield, ionic balance and solute composition of pigeon pea (*Cajanus cajan* (L.) Mill Sp.). *Pakistan journal of botany*. 2006;38(4):1103.
 31. Williams CH, Steinbergs A. Soil sulphur (Heat soluble sulphur or available sulphur) fractions as chemical indices of available sulphur in some Australian soils. *Australian Journal of Agricultural Research*. 1959;10(3):340-352.