www.ThePharmaJournal.com

The Pharma Innovation



ISSN (E): 2277-7695 ISSN (P): 2349-8242 NAAS Rating: 5.23 TPI 2023; 12(6): 1345-1348 © 2023 TPI www.thepharmajournal.com

Received: 08-04-2023 Accepted: 19-05-2023

Savitha V

PG Student, School of Agricultural Sciences, Karunya Institute of Technology and Sciences, Coimbatore, Tamil Nadu, India

Anitha M

Assistant Professor, School of Agricultural Sciences, Karunya Institute of Technology and Sciences, Coimbatore, Tamil Nadu, India

Gopi V

Assistant Professor, School of Agricultural Sciences, Karunya Institute of Technology and Sciences, Coimbatore, Tamil Nadu, India

Vinu Radha R

Assistant Professor, School of Agricultural Sciences, Karunya Institute of Technology and Sciences, Coimbatore, Tamil Nadu, India

Jeevitha D

Assistant Professor, School of Agricultural Sciences, Karunya Institute of Technology and Sciences, Coimbatore, Tamil Nadu, India

Irine Chacko

PG Student, School of Agricultural Sciences, Karunya Institute of Technology and Sciences, Coimbatore, Tamil Nadu, India

Pavethra A

PG Student, School of Agricultural Sciences, Karunya Institute of Technology and Sciences, Coimbatore, Tamil Nadu, India

Anjana RJ

PG Student, School of Agricultural Sciences, Karunya Institute of Technology and Sciences, Coimbatore, Tamil Nadu, India

Naveen Kumar S

PG Student, School of Agricultural Sciences, Karunya Institute of Technology and Sciences, Coimbatore, Tamil Nadu, India

Corresponding Author: Anitha M

Assistant Professor, School of Agricultural Sciences, Karunya Institute of Technology and Sciences, Coimbatore, Tamil Nadu, India

Effect of biostimulants on growth of Red banana plantlets during the primary hardening phase

Savitha V, Anitha M, Gopi V, Vinu Radha R, Jeevitha D, Irine Chacko, Pavethra A, Anjana RJ and Naveen Kumar S

Abstract

An experiment was conducted to evaluate the effect of various biostimulants during the primary hardening phase of tissue culture plantlets of Banana cv. Red Banana at the North Farm, School of Agricultural Sciences, Karunya Institute of Technology and Sciences, Coimbatore in the year 2022-2023. Among the various treatments, the maximum survival percentage (100%) during the primary hardening phase was observed in Soil + Cocopeat + Chitosan (50 ppm), Soil + Cocopeat + Chitosan (100 ppm), Soil + Cocopeat + Moringa leaf extract (3%), Soil + Cocopeat + Moringa leaf extract (4%), Soil+ Cocopeat + Vermiwash (3%), Soil + Cocopeat + Fermented coconut water. Among the various treatments given the maximum plant height (12.65 cm), pseudostem girth (4.12 cm), leaf length (7.51 cm), leaf width (4.27 cm), leaf area (25.70 cm²) and number of leaves per plant (4.75) was observed in Soil + Cocopeat + Moringa leaf extract (4%). The SPAD value was highest (38.24) in the treatment of Soil + Cocopeat + Chitosan (50 ppm).

Keywords: Red banana, biostimulants, chitosan, hardening, tissue culture

Introduction

Bananas and plantains (*Musa* spp.) are the first crop plants domesticated by humans. Banana is a large monocotyledonous and monocarpic herbaceous perennial plant that belongs to the family of Musaceae and Zingiberales order. Bananas are otherwise called "Apple of Paradise" or "Kalpavriksh" which means the plant of virtue (Singh *et al.*, 2009) ^[1]. Banana is the most important tropical fruit next to mango. India is the largest producer of bananas in the world with an area of 880,000 ha and a production of 32454'000 MT (NHB, 2022) ^[2]. The Red banana (*Musa acuminata*) is the most relished and highly prized variety of bananas. It is a triploid cultivar (AAA) that is commercially grown in the Kanyakumari and Tirunelveli districts of Tamil Nadu.

Healthy and disease free planting material is an important factor for long term banana production. To meet the ever increasing demand for elite planting material, traditional propagation methods must be supplemented with modern propagation techniques. This increasing demand for high quality banana planting material can only be met if conventional and non-conventional banana propagation methods are used on a commercial scale, with need-based modifications involving modern propagation technologies.

Micropropagation or tissue culture is the only viable option for large scale banana multiplication. This method is free of seasonal bonds and allows for the multiplication of the chosen plants. Other benefits include product uniformity, disease free plants, and the ease of exchanging germplasm and planting material (Murashige *et al.*, 1974) ^[3]. Acclimatization or hardening is a critical step in the micropropagation process. It is a process of adaptation to the natural environment for various plant species that have undergone *in vitro* growth and development. Most *in vitro* species require an acclimatization process to ensure that a sufficient number of plants survive and grow vigorously when transferred to soil. Acclimatization of *in vitro* raised plants to *ex vitro* conditions is critical and a major determinant of these plants subsequent field establishment (Singh *et al.*, 2012) ^[4]. The benefit of any micropropagation system can only be realized if plantlets are successfully transferred to the soil through the acclimatization process, morphological characteristics can be visible (Singh *et al.*, 2017) ^[5]. The selection of suitable media and biostimulants are important during the acclimatization process in crops.

Natural biostimulants are gaining popularity in sustainable agriculture for improving crop productivity and quality (Abdalla, 2013)^[6]. Biostimulants are a diverse group of compounds capable of enhancing certain physiological processes that promote crop growth and development and also reduces the use of synthetic fertilizers while maintaining crop productivity. Plant biostimulant is also a substance or microorganism which is applied to plants to improve nutrition efficiency, abiotic stress tolerance, and crop quality traits, regardless of nutrient content. Biochar, chitosan derivatives, humic substances, microbial inoculants, moringa leaf extract, vermiwash, and seaweed extracts are the most widely used natural biostimulants in crop production (Jardin, 2015)^[7].

The plantlets when treated with various biostimulants may improve establishment and overall performance. A beneficial organic extract in commercial cultivation would reduce pesticides, saving costs and reducing pollution to the environment. With the above background, the experiment was done to the study the efficiency of various biostimulants on survival percentage and growth parameters of tissue cultured red banana plantlets.

Materials and Methods

The present investigation was taken up at the North Farm, School of Agricultural Sciences, Karunya Institute of Technology and Sciences, Coimbatore of Tamil Nadu. The experiment was laid out in a Completely Randomized Design with eleven treatments and three replications. The following treatments T₁: Soil + Cocopeat (Control), T₂: Soil + Cocopeat + Chitosan (50 ppm), T₃: Soil + Cocopeat + Chitosan (100 ppm), T₄: Soil + Cocopeat + Moringa leaf extract (3%), T₅: Soil + Cocopeat + Moringa leaf extract (4%), T₆: Soil + Cocopeat + Vermiwash (3%), T₇: Soil + Cocopeat + Vermiwash (4%), T₈: Soil + Cocopeat + Cow urine (10 times diluted), T₉: Soil + Cocopeat + Cow urine (5 times diluted), T₁₀: Soil + Cocopeat + Fermented coconut water and T₁₁: Soil + Cocopeat + Humic acid were given to the red banana plantlets at fortnight intervals.

Results and Discussion

The salient findings of the study are presented in Table 1 and discussed below.

Survival Percentage: The survival percentage of red banana plantlets during the primary hardening phase was recorded maximum of 100% in T₂: Soil + Cocopeat + Chitosan (50 ppm), T₃: Soil + Cocopeat + Chitosan (100 ppm), T₄: Soil + Cocopeat + Moringa leaf extract (3%), T₅: Soil + Cocopeat + Moringa leaf extract (4%), T₆: Soil+ Cocopeat + Vermiwash (3%) and T10: Soil + Cocopeat + Fermented coconut water whereas it was minimum in the control (66.67%). The higher survival percentage observed in the red banana plantlets treated with moringa leaf extract may be due to the presence of antioxidants in it. The above result is in accordance with the findings of Guler et al., (2021)^[8] and it is further supported by the research findings of Veraplakorn and Kudan (2021)^[9] using chitosan, Molnar et al., (2011)^[10] reported that the liquid endosperm in coconut water contains a variety of amino acids, organic acids, nucleic acids, vitamins, sugars, sugar alcohols, plant hormones (auxins and cytokinins), minerals that are entirely responsible for the growth promoting activity in plants.

Plant height (cm)

Plant height recorded in the tissue cultured red banana plantlets during the primary hardening phase was maximum (12.65 cm) in the treatment of T₅: Soil + Cocopeat + Moringa leaf extract (4%) followed by T₉: Soil + Cocopeat + Cow urine (5 times diluted) (12.05 cm) whereas it was minimum in the control (9.32 cm). The increased plant height may be due to the presence of zeatin in moringa leaf extract (Culver et al., 2012) ^[11]. The above result is also supported by the findings of Sardar et al., (2021) [12] who opined that Moringa Leaf Extract (MLE) is effective in improving vegetative growth, vield, and quality of numerous crops around the world. It is also quite inexpensive compared to synthetic growth regulators. According to Bashir et al., (2014) ^[13], Moringa leaf extract increased the average plant height, number of leaves, branches, and yield parameters. The increased plant height observed in plantlets receiving the treatment of T₉: Soil + Cocopeat + Cow urine (5 times diluted) may be due to the presence of macro and micro nutrients present in it (Ambiga and Balakrishnan, 2015)^[14].

Pseudostem girth (cm)

Pseudostem girth was recorded maximum (4.12 cm) in the treatment of T_5 : Soil + Cocopeat + Moringa leaf extract (4%) followed by T_2 : Soil + Cocopeat + Chitosan (50 ppm) (3.85 cm) whereas it was recorded minimum in the control (2.40 cm). The increased pseudostem girth may be due to the presence of cytokinin present in moringa leaf extract which promoted the cell division and cell elongation (Yusuff *et al.*, 2020) ^[15]. The increase in pseudostem girth due to chitosan is supported by the findings of El-Miniawy *et al.*, (2013) ^[16] who observed that chitosan spray on strawberries after transplantation increased the vegetative growth and yield.

Leaf length (cm)

Leaf length was recorded maximum (7.51 cm) in the treatment of T_5 : Soil + Cocopeat + Moringa leaf extract (4%) followed by T_9 : Soil + Cocopeat + Cow urine (5 times diluted) (7.42 cm). The control plants which were not treated by any biostimulants recorded the least length of 5.03 cm. The above result is in accordance with the findings of Ahmed *et al.*, (2020) ^[17] and Jandaik *et al.*, (2015) ^[18]. Hasegawa *et al.*, (2000) ^[19] opined that by activating enzymes, osmoregulation, and photosynthesis, the presence of calcium and potassium in Moringa Leaf Extract can potentially promote plant growth and development.

Leaf width (cm)

Leaf width was recorded maximum (4.27 cm) in the treatment of T₅: Soil + Cocopeat + Moringa leaf extract (4%) followed by T₆: Soil + Cocopeat + Vermiwash (3%) (4.15 cm). The control plants which were not treated by any biostimulants recorded the least length of 2.13 cm. The results are in accordance with the findings of Foidle *et al.* (2001) ^[20]. According to Subasashri (2003) ^[21], vermiwash is an excellent liquid manure that greatly impacts crop growth and productivity when applied as a foliar spray.

Leaf area (cm²)

In the primary hardening phase of red banana plantlets, leaf area was found to be maximum (25.70 cm²) in the treatment of T₅: Soil + Cocopeat + Moringa leaf extract (4%) followed by T₉: Soil + Cocopeat + Cow urine (5 times diluted) (24.05

cm²). The control plants which were not treated by any biostimulants recorded a minimum leaf area of 8.60 cm². The results are in accordance with the findings of Rehman and Basra, (2010) ^[22] who stated that the foliar application of moringa leaf extract promotes earlier cytokinin synthesis, prevents premature leaf senescence, and results in greater leaf area with higher photosynthetic pigments.

Number of leaves per plant

The present study revealed that application of T_5 : Soil + Cocopeat + Moringa leaf extract (4%) increased the leaf production (4.75) in red banana plantlets. The above result is on par with T_4 : Soil + Cocopeat + Moringa leaf extract (3%), T_8 : Soil + Cocopeat + Cow urine (10 times diluted) and T_6 : Soil + Cocopeat + Vermiwash (3%). The above result is in accordance with the findings of Nasir *et al.*, (2016) ^[23], Sahu

et al., (2022) ^[24].

SPAD value

The chlorophyll content was measured by SPAD chlorophyll meter (model 502; Minolta Corp., Ramsey, N.J) and presented in Table 1. The SPAD value was maximum (38.24) in the plants which received the treatment of T₂: Soil + Cocopeat + Chitosan (50 ppm) followed by T₃: Soil + Cocopeat + Chitosan (100 ppm) (37.32) and it was minimum in control plantlets (29.20). The result is in accordance with the findings of Gornik *et al.*, (2008) ^[25] where dipping stem cuttings of grapevines in chitosan improved the subsequent rooting and increased the chlorophyll content. The above results are also supported by Van *et al.*, (2013) ^[26] who reported that foliar spray of chitosan enhanced leaf chlorophyll content, net photosynthesis rate, and nutrient uptake.

Table 1: Effect of biostimulants on growth of Red banana plantlets during the primary hardening phase

Treatments	Survival	Plant height	Pseudostem girth	Leaf length	Leaf width	Leaf area	Number of leaves	SPAD
	Percentage (%)	(cm)	(cm)	(cm)	(cm)	(cm ²)	per plant	value
T1	66.33	9.32	2.40	5.03	2.13	8.61	3.22	29.27
T2	100.00	11.50	3.85	6.89	3.43	18.96	4.21	38.24
T ₃	100.00	11.77	3.27	6.37	3.97	20.24	4.38	38.10
T 4	100.00	11.80	3.67	6.93	4.03	22.36	4.69	35.16
T5	100.00	12.65	4.12	7.51	4.27	25.70	4.75	36.60
T ₆	100.00	10.43	3.61	6.12	4.15	20.34	4.58	33.03
T 7	95.00	11.27	3.49	6.85	3.80	20.86	3.52	36.06
T8	95.00	10.86	3.79	6.32	3.74	18.93	4.62	37.32
T9	95.00	12.05	3.69	7.42	4.05	24.05	3.61	34.82
T10	100.00	11.33	3.12	6.24	2.58	12.94	3.68	35.45
T ₁₁	90.00	10.23	3.32	5.93	3.26	15.49	3.82	32.26
SE.d	4.71	0.17	0.16	0.18	0.11	0.79	0.13	0.51
CD (0.05)	9.82	0.37	0.32	0.38	0.23	1.64	0.28	1.07

Conclusion

From the present study it was concluded that Soil + Cocopeat + Moringa Leaf Extract (4%) (T_5) treatment recorded the maximum survivability, plant height, pseudostem girth, maximum leaf length, leaf width, leaf area and number of leaves per plant in red banana plantlets during the primary hardening phase. It was also found that red banana plantlets which received the treatment of Soil + Cocopeat + Chitosan (50 ppm) (T_2) and Soil + Cocopeat + Chitosan (100 ppm) (T_3) recorded the maximum SPAD values. Hence the effect of various biostimulants on the growth of red banana plantlets during the primary hardening phase were studied.

Acknowledgment

The authors are thankful to the School of Agricultural Sciences, Karunya Institute of Technology and Sciences, for their immense support and facilities provided during this work.

Reference

- 1. Singh HP, Uma S, Selvarajan R, Karihaloo JL. Micropropagation for production of quality banana planting material in Asia-pacific. Asia Pacific Consortium on Agricultural Biotechnology, New Delhi. 2009, 92.
- 2. NHB. 2022. https://nhb.gov.in/report_files/banana/BANANA.htm
- Murashige TM, Serpa, Jones JB. Clonal multiplication of Gerbera through tissue culture. Horticultural Science. 1974;9:175-180.

- Singh NV, Singh SK, Singh AK, Meshram DT, Suroshe SS, Mishra DC. *Arbuscular mycorrhizal* fungi (AMF) induced hardening of micro propagated pomegranate (*Punica granatum* L.) plantlets. Scientia Horticulture. 2012;136:122-7.
- Singh VK, Prasad VM, Kumari S, Rajoria P, Misra P. Identification of the suitable hardening protocol and hardening medium in micropropagation of gerbera (*Gerbera jamesonii* Bolus). International Journal of Current Microbiology and Applied Sciences. 2017;6(7):2476-2484.
- 6. Abdalla MM. The potential of *Moringa oleifera* extract as a biostimulant in enhancing the growth, biochemical and hormonal contents in rocket (*Eruca vesicaria* subsp. *sativa*) plants. International Journal of Plant Physiology and Biochemistry. 2013;5:42-49.
- Jardin PD. Plant biostimulants: Definition, concept, main categories and regulation. Scientia Horticulture. 2015;196:3-14.
- 8. Guler G, Gubbuk H, Arslan MA. The effects of antioxidants on micropropagation of Avocado by nodal segments. Horticultural Studies. 2021;38(1):50-55.
- Veraplakorn V, Kudan S. Chitosan elicitor stimulation of *invitro* growth and *ex vitro* acclimatization of *Lantana camara* L. Agriculture and Natural Resources. 2021;55:431-439.
- Molnar Z, Virog E, Ordog V. Natural Substances in tissue culture media of higher plants. Acta Biol Szegediesis. 2011;55:123-7.
- 11. Culver M, Fanuel T, Chiteka AZ. Effect of moringa

extract on growth and yield of tomato. Greener Journal of Agricultural Sciences. 2012;2(5):207-211.

- 12. Sardar H, Nisar A, Anjum MA, Naz S, Ejaz S, Ali S, *et al.* Foliar spray of moringa leaf extract improves growth and concentration of pigment, minerals and stevioside in stevia (*Stevia rebaudiana* Bertoni). Industrial Crops and Products. 2021;166:113485.
- Bashir KA, Bawa JA, Mohammed I. Efficacy of leaf extract of drumstick tree (*Moringa Oleifera* Lam.) on the growth of local tomato (*Lycopersicon esculentum*). IOSR Journal of Pharmacy and Biological Sciences. 2014;9(4):74-77.
- Ambika S, Balakrishnan K. Enhancing germination and seedling Vigour in cluster bean by organic priming. Academic Journal. 2015;10(8):298-301.
- Yussuf AQ, Adedeji MS, Falana AR, Majekodunmi OA. Efficacy of moringa extract on growth and yield of Okra. Journal of Agricultural Research Advances. 2020;2(1):1-7.
- El Miniawy S, Ragab M, Youssef S, Metwally A. Response of strawberry plants to foliar spraying of chitosan. Journal of Agriculture and Biological Sciences. 2013;9:366-372.
- 17. Ahmed MEM, Al-Ballat IA, Elzaawely AA. Moringa leaf extract stimulates growth and yield of cucumber (*Cucumis sativus* L.). Menoufia Journal of Plant Production. 2020;5:63-75.
- Jandaik S, Thakur P, Kumar V. Efficacy of cow urine as plant growth enhancer and antifungal agent. Advances in Agriculture. 2015. https://doi.org/10.1155/2015/620368
- Hasegawa PM, Bressan RA, Zhu JK, Bohnert HJ. Plant cellular and molecular responses to high salinity. Annual Review of Plant Physiology. 2000;51:463-499.
- Foidle N, Makkar HPS, Becker K. The potential of Moringa oleifera for agricultural and industrial uses, the miracle tree: The multipurpose attributes of moringa. CTA publications; c2001. p. 45-76.
- 21. Subasashri M. Vermiwash collection and its pesticidal properties. The Hindu, 2003;17:1-2.
- 22. Rehman H, Basra SMA. Growing *Moringa oleifera* as a multipurpose tree; some agro-physiological and industrial perspectives. American Chronicle, 2016. http://www. American chronicle.com/articles/view/159447
- 23. Nasir M, Khan AS, Basra SMA, Malik AU. Foliar application of moringa leaf extract, potassium and zinc influence yield and fruit quality of 'Kinnow' mandarin. Scientia Horticulturae. 2016;210:227-235.
- 24. Sahu K, Sahu GD, Khunte SD. Effect of different sources of organic fertilizers on vegetative growth of Banana (*Musa paradisiaca* L.) under precision farming. The Pharma Innovation Journal. 2022;11(10):317-319.
- 25. Gornik K, Grzesik M, Romanowska Duda B. The effect of chitosan on rooting of grapevine cuttings and on subsequent plant growth under drought and temperature stress. Journal of Fruit and Ornamental Plant. 2008;16:333-343.
- 26. Van SN, Minh HD, Anh DN. Study on chitosan nanoparticles on biophysical characteristics and growth of Robusta coffee in green house. Biocatalysis and Agriculture Biotechnology. 2013;2:289-294.