



ISSN (E): 2277-7695
 ISSN (P): 2349-8242
 NAAS Rating: 5.23
 TPI 2023; 12(8): 2359-2362
 © 2023 TPI

www.thepharmajournal.com

Received: 06-05-2023

Accepted: 10-06-2023

Author's details are given below
 the reference section

Effect of different weed management practices on weed dynamics of blackgram in Northeast India

Nancy Khwairakpam, Luikham Edwin, Jamkhogin Lhungdim, Herojit Singh Athokpam, N Brajendra Singh, N Okendro Singh, N Anando Singh, Herojit Khaidem, MM Rifana Ajam, Surbani Laishram, G Prameshchandra Sharma, Heisnam Henarita, B Keyeang, Thangneibong Serto, O Yaiphabee and M Puja Devi

Abstract

During the *kharif* season 2022, a field experiment was carried out at the College of Agriculture, Central Agricultural University, Imphal, Manipur. The experiment consisted of 8 treatments i.e. T₁ (hand weeding at 20 and 40 DAS), T₂ (rice husk @ 10 t/ha), T₃ (pendimethalin @ 1 kg/ha), T₄ (imazethapyr @ 0.07 kg/ha), T₅ (imazethapyr @ 0.08 kg/ha), T₆ (pendimethalin @ 1 kg/ha *fb* imazethapyr @ 0.07 kg/ha), T₇ (pendimethalin @ 1 kg/ha *fb* imazethapyr @ 0.08 kg/ha), T₈ (weedy check/control) was laid out in randomized block design replicated thrice. Blackgram variety Pant U31 was sown with the spacing 30 cm x 15 cm utilizing 20 kg of seed per hectare fertilized with the recommended dose of N:P₂O₅:K₂O i.e. 20:40:20 kg per hectare. The treatment pendimethalin @ 1 kg/ha *fb* imazethapyr @ 0.08 kg/ha followed by treatment T₁ (hand weeding at 20 and 40 DAS) which was statistically on par with each other were found out to be most effective weed management practices for blackgram in respect of obtaining the lowest weed count/m², highest WCE as well as lowest fresh and dry weight of the weeds.

Keywords: Blackgram, weed, hand weeding, pendimethalin, imazethapyr

Introduction

India is the world's largest producer, consumer, importer, and producer of pulses by area. About 20% of the area planted with food grains is dedicated to growing pulses, which produce 7-10 percent of the nation's total grain production. Pulses contain protein ranging from 17 to 27 percent. The amount of pulses produced falls considerably short of what is needed to even reach the very minimal level of per capita consumption. Contrary to FAO/WHO recommendations, which call for 104 g of pulses per person per day, the availability of pulses per person is just 45 g. Meeting the country's growing population's demand for pulses is therefore a significant challenge for agricultural scientists. Pulses as a leguminous crop, can fix and use atmospheric nitrogen besides helping to increase the fertility of soil. The total production of pulses in India during the past 15 years has impressively increased from 13.38 million MT in 2005-2006 to 25.58 million MT in 2020-2021 (Anon., 2020-2021) [3]. Production of blackgram has primarily been spread in tropical and subtropical countries. In India, it is grown during the *kharif*, *rabi*, and summer seasons. Infestation of the weed is the main reason for the reduction of the yield (i.e. low productivity of the crop) of blackgram grown during the *kharif* season. Weed losses are more than the entire yearly losses of agricultural products due to several categories such insects, illnesses, other pests, nematodes, etc. In general, the yield of the crops is reduced due to the presence of weeds around 31.5% i.e. 22.7% in the winter season and 36.5% in the summer and *kharif* season and sometimes it also causes complete devastation of the crop in India (Anon., 2007) [2]. Weeds usually compete with the crops in terms of nutrients, moisture and light thus, deplete crops environment of nutrients, waters, and light. During the growth period of the crop, if it receives frequent rainfall along with high temperature the weeds infest the crop growth heavily that leads in lowering the productivity of the crop. Against weed blackgram is not a good competitor at the early stages of the crop growth (Choudhary *et al.*, 2012) [5]. Weeds can be controlled by cultural, manual, biological, mechanical and chemical methods. In blackgram physical methods or the mechanical methods are the traditional method to control the weeds. Manual weeding is required to keep the crop weed-free, but due to time consumption, labour intensive as well as

Corresponding Author:
 N Anando Singh
 Scientist, Department of
 Agronomy, College of
 Agriculture, Central Agricultural
 University, Imphal, India

increasing labour crisis it requires other possible alternatives to control the weed population. Herbicides can be used to control weeds as an alternative to other weed management techniques. A single herbicide application might not be sufficient for broad-spectrum weed control instead application of pre- and post-emergence herbicide in sequence, mix or integrated with hand weeding's possibly a better option than single application. Application of a post-emergence herbicide is a suitable alternative to control the 2nd weed flush in pulses since it also minimizes the need for human labor. (Singh *et al.*, 2014) [11]. In light of these circumstances, the current study analyzed the effects of different weed management practices in order to determine the best performing course of action.

Materials and Methods

During the *kharif* season 2022, a field experiment was carried out at the College of Agriculture, Central Agricultural University, Imphal, Manipur. The experimental site is located 774.5 meters above mean sea level at latitude 24°45' N and longitude 93°54' E. The texture of the experimental soil was clay with sand 9.28%, silt 24.72% and clay 66%, highly acidic (pH 5.25), high organic carbon content with 1.11%, low available nitrogen content (263.42 kg/ha), low phosphorus content (20.82 kg/ha) and medium potassium content (232.31 kg/ha). The minimum and maximum temperatures that were logged while the crops were growing The temperatures (highest and lowest points) that were logged during the period of crop growth under review were 22.80 °C and 30.05 °C respectively with an average rainfall of 157.00 mm and average sunshine hours of 4.58 hrs. With eight treatments and three replications, the experiment was set out using randomized block design. The treatments were: T₁ (hand weeding at 20 and 40 DAS), T₂ (rice husk@10 t/ha), T₃ (pendimethalin @ 1 kg/ha), T₄ (imazethapyr @ 0.07 kg/ha), T₅ (imazethapyr @ 0.08 kg/ha), T₆ (pendimethalin @ 1 kg/ha fb imazethapyr @ 0.07 kg/ha), T₇ (pendimethalin @ 1 kg/ha fb imazethapyr @ 0.08 kg/ha), T₈ (weedy check). The blackgram variety Pant U31 was sown on 16th June 2022 with the spacing of 30cm x15cm using seed rate of 20kg/ha, the recommended fertilizer dose used was 20:40:20 N:P₂O₅:K₂O kg/ha respectively. The requisite amount of spray solution was sprayed with the help of hand sprayer. The quantity of

water used was 500 litres/ha. Pendimethalin was applied as pre-emergence (two DAS) and imazethapyr as post-emergence (nineteen DAS). Using a *khurpi*, hand weeding was carried out manually at 20 and 40 DAS as per specific treatment schedule. A quadrat of 0.25 square meters was used to measure the number of weeds in a confined area, randomly selected in each individual plot and converted into weed count per m². The weeds removed from the quadrat at 20, 40 and 60 DAS were cleaned, washed and weighed fresh weight of the weeds afterwards the weeds were weighed and expressed in g/m² as their dry weight after being placed in the oven for 72 hours at 60 °C. Using the formula suggested by Mani *et al.* (1973), weed control efficiency (%) was calculated.

$$WCE (\%) = \frac{DWC - DWT}{DWC} \times 100$$

Where, D.W.C is the dry weight of weeds in control and D.W.T is the dry weight of weeds in treated plots. The analysis of variance technique described by Gomez and Gomez (1984) [6] were used to compute the data obtained on the experiment. Square root transformation $(x+0.5)^{1/2}$ was applied to the data that displayed a high degree of variation, whenever the treatment differences were significant, for the comparison of the treatment critical difference at five percent probability level were worked out. While the non-significant were represented as NS.

Results and Discussion

Weed parameters

Weed flora

The predominant weed flora found in the experimental field during the *kharif* season 2022 was composed of sixteen species, belonging to eight different families. The dominant weeds were *Echinochloa colona*, *Digitaria sanguinalis*, *Cynodon dactylon*, *Galinsoga parviflora*, *Bidens pilosa*, *Euphorbia hirta*, *Spilanthes paniculata* (Table 1). Similar weed flora was also noticed by Jagadesh *et al.* (2019). Among which poaceae family weeds were dominated in the monocot weeds and asteraceae family weeds were dominated in the dicot weeds.

Table 1: Weed flora of blackgram field during the experimental season

| Category | Sl. No. | Scientific name | Common name | Family |
|--------------|---------|------------------------------------|---------------------|---------------|
| Monocot weed | 1 | <i>Echinochloa colona</i> | Jungle rice | Poaceae |
| | 2 | <i>Digitaria sanguinalis</i> | Hairy crabgrass | Poaceae |
| | 3 | <i>Juncus tenuis</i> | Path rush | Juncaceae |
| | 4 | <i>Juncus articulatus</i> | Jointleaf rush | Juncaceae |
| | 5 | <i>Setaria parviflora</i> | Marsh bristlegrass | Poaceae |
| | 6 | <i>Cynodon dactylon</i> | Bermuda grass | Poaceae |
| | 7 | <i>Cyperus rotundus</i> | Purple nutsedge | Cyperaceae |
| | 8 | <i>Sphenopholis obtusata</i> | Prairie wedge grass | Poaceae |
| Dicot weed | 9 | <i>Galinsoga parviflora</i> | Gallant soldier | Asteraceae |
| | 10 | <i>Cardamine hirsuta</i> | Hairy bittercress | Brassicaceae |
| | 11 | <i>Crassocephalum crepidioides</i> | Fireweed | Asteraceae |
| | 12 | <i>Euphorbia hirta</i> | Asthma weed | Euphorbiaceae |
| | 13 | <i>Bidens pilosa</i> | Black jack | Asteraceae |
| | 14 | <i>Ludwigia prostrata</i> | heusenkraut | Onagraceae |
| | 15 | <i>Aeschynomene indica</i> | Indian jointvetch | Leguminosae |
| | 16 | <i>Spilanthes paniculata</i> | Toothache plant | Asteraceae |

Weed count/m²

The total weed count/m² reduced acutely at all crop growth stages under various treatments (except 20 DAS) in comparison to weedy check. Analogous results were obtained by Yadav *et al.* (2014) [12], who concluded in comparison with weedy check the weed control techniques significantly reduce the weeds population in blackgram. Significantly, the lowest monocot weed count/m² was recorded under the treatment T₇ at 20, 40 and 60 DAS *fb* T₁ (Table 2). Regarding the dicot weed count/m² at 20, 40 and 60 DAS lowest was recorded

under the treatment T₇ which was statistically comparable with the value T₆ at 20 DAS. At 40 DAS and 60 DAS it was found to be on par with T₁. And significantly the lowest total weed count/m² were recorded under T₇ at 20, 40 and 60 DAS which was on par with T₁ except at 20 DAS, while the highest was noted under weedy check at all the stages except at 20 DAS which was on par with T₁ the trend follows the same for dicot as well as monocot. Identical set of results was also discovered by Balyan *et al.* (2016) [4].

Table 2: Effect of weed management practices on weed count/m² in blackgram

| Treatments | Monocot weed count/m ² | | | Dicot weed count/m ² | | | Total weed count/m ² | | |
|----------------|-----------------------------------|------------------|------------------|---------------------------------|---------------|---------------|---------------------------------|--------|--------|
| | 20 DAS | 40 DAS | 60 DAS | 20 DAS | 40 DAS | 60 DAS | 20 DAS | 40 DAS | 60 DAS |
| T ₁ | 20.33 (4.56)* | 13.33 (3.72)* | 16.33 (4.10)* | 22.67 (4.81)* | 12.33 (3.59)* | 16.33 (4.10)* | 43.00 | 25.67 | 32.67 |
| T ₂ | 18.33 (4.34)* | 28.33 (5.37)* | 39.67 (6.34)* | 16.00 (4.06)* | 36.00 (6.04)* | 41.33 (6.47)* | 34.00 | 64.33 | 81.00 |
| T ₃ | 18.67 (4.38)* | 26.33 (5.18)* | 33.00 (5.79)* | 13.67 (3.76)* | 28.00 (5.34)* | 37.33 (6.15)* | 32.33 | 54.33 | 70.33 |
| T ₄ | 18.33 (4.34)* | 22.67 (4.81)* | 28.00 (5.33)* | 17.00 (4.18)* | 24.33 (4.98)* | 37.00 (6.12)* | 35.33 | 47.00 | 66.00 |
| T ₅ | 16.33 (4.10)* | 22.33 (4.78)* | 27.67 (5.31)* | 16.67 (4.14)* | 22.33 (4.78)* | 35.33 (5.99)* | 33.00 | 44.67 | 63.00 |
| T ₆ | 14.00 (3.81)* | 13.67 (3.76)* | 17.67 (4.26)* | 13.00 (3.67)* | 15.33 (3.98)* | 18.67 (4.38)* | 27.00 | 29.00 | 36.33 |
| T ₇ | 11.00 (3.38)* | 12.00 (3.53)* | 14.67 (3.89)* | 11.67 (3.49)* | 12.33 (3.58)* | 16.33 (4.10)* | 22.67 | 24.33 | 31.00 |
| T ₈ | 21.33 (4.67)* | 41.67 (6.49)* | 44.33 (6.69)* | 22.67 (4.81)* | 42.33 (6.54)* | 44.67 (6.72)* | 44.00 | 84.00 | 89.00 |
| SEd (±) | 0.75 (0.10)* | 0.54 (0.06)* | 0.67 (0.07)* | 0.82 (0.11)* | 0.58 (0.06)* | 0.60 (0.05)* | 1.25 | 0.79 | 0.94 |
| C.D 5% | 1.59 (0.22)* | 1.15 (0.13)* | 1.46 (0.15)* | 1.77 (0.23)* | 1.23 (0.13)* | 1.29 (0.12)* | 2.68 | 1.7 | 2.03 |

(* - The values in the parenthesis are square root transformed)

Fresh and dry weight of weed (g/m²)

All the treatments remarkably lessened the fresh and dry weight of the weeds over weedy check. Among the treatments, minimum fresh and dry weight of weed was recorded with the treatment T₇ at 20, 40 and 60 DAS, which was discovered to be on par with T₁ at 40 and 60 DAS (Table 3). However, at 20 DAS T₇ was found on par with T₆ for fresh weight and T₃ for dry weight of the weed. Under weedy check, the maximum fresh and dry weight of the weed was

reported at 20, 40 and 60 DAS. The same outcomes were likewise attained by Ali *et al.* (2011) [1] and Painkra *et al.* (2021) [9]. Data disclosed that different weed management practices controlled weeds effectively in comparison with weedy check. At different period of the crop growth variation in weight of the weeds can be observed this was because of the effect of weed management practices as for herbicidal treatment the dose applied and the type of herbicide used leads to variation.

Table 3: Effect of weed management practices on fresh and dry weight of the weeds and WCE in blackgram

| Treatments | Fresh weight of weed (g/m ²) | | | Dry weight of weed (g/m ²) | | | WCE (%) | | |
|----------------|------------------------------------------|--------|--------|----------------------------------------|--------|--------|---------|--------|--------|
| | 20 DAS | 40 DAS | 60 DAS | 20 DAS | 40 DAS | 60 DAS | 20 DAS | 40 DAS | 60 DAS |
| T ₁ | 20.72 | 19.56 | 28.32 | 5.35 | 4.35 | 10.34 | 4.91 | 86.24 | 85.11 |
| T ₂ | 7.63 | 36.17 | 89.34 | 2.95 | 18.27 | 41.73 | 47.66 | 42.15 | 39.92 |
| T ₃ | 6.54 | 31.50 | 75.43 | 1.15 | 10.12 | 29.42 | 79.57 | 67.95 | 57.64 |
| T ₄ | 8.36 | 28.36 | 57.32 | 2.83 | 9.11 | 25.37 | 49.73 | 71.16 | 63.47 |
| T ₅ | 6.78 | 25.67 | 53.52 | 2.00 | 7.38 | 20.52 | 64.41 | 76.62 | 70.45 |
| T ₆ | 6.09 | 23.68 | 34.45 | 1.88 | 5.74 | 13.53 | 66.65 | 81.82 | 80.51 |
| T ₇ | 5.54 | 19.43 | 29.20 | 1.04 | 4.14 | 10.73 | 81.58 | 86.88 | 84.54 |
| T ₈ | 22.15 | 65.09 | 112.46 | 5.63 | 31.59 | 69.45 | 0 | 0 | 0 |
| SEd (±) | 0.39 | 0.29 | 0.11 | 0.03 | 0.10 | 0.19 | 0.59 | 0.32 | 0.27 |
| C.D 5% | 0.83 | 0.62 | 0.25 | 0.07 | 0.22 | 0.42 | 1.25 | 0.69 | 0.58 |

Weed control efficiency

The treatment T₇ recorded the highest value of WCE at 20, 40 and 60 DAS (81.58%, 86.88% and 84.54%) which was

statistically at par with T₁ at 40 and 60 DAS (86.24% and 85.11%) and the next superior treatment was T₃ (79.57%) and T₆ (81.82%) respectively at 20 and 40 DAS (Table 3). Lowest

value was observed under rice husk @ 10 t/ha at 40 and 60 DAS (42.15% and 39.92%). The highest value of WCE may have resulted from a reduction in weed spectrum during the early stages of crop growth period that results in reduction of biomass of weed. Analogous findings were reported by Rai *et al.* (2016) [10] and Painkra *et al.* (2021) [9].

Conclusion

Considering the aforementioned results it could be resolved that the treatment T₇ followed by treatment T₁ which did not differ from each other were observed to be most effective weed management practices for blackgram in respect of obtaining the lowest weed count/m², highest WCE as well as lowest fresh and dry weight of the weeds.

References

1. Ali S, Patel JC, Desai LJ, Singh J. Effect of herbicides on weeds and yield of rainy season green gram (*Vigna radiata* L. Wilczek). Legum. Res. 2011;34(4):300-303.
2. Anonymous. Vision 2025. NRCWS Perspective Plan. Indian Council of Agricultural Research (ICAR), New Delhi, India. 2007.
3. Anonymous. A Brief Handbook, Directorate of Pulses Development, Government of India, Bhopal; c2020-2021.
4. Balyan JK, Choudhary RS, Kumpawat BS, Choudhary R. Weed management in blackgram under rainfed conditions. Indian J Weed Sci. 2016;48(2):173-177.
5. Choudhary VK, Kumar PS, Bhagawati R. Integrated weed management in blackgram (*Vigna mungo*) under mid hills of Arunachal Pradesh. Indian J Agron. 2012;57(4):382-385.
6. Gomez KA, Gomez AA. Statistical Procedures for Agricultural Research. 2nd edn., John Wiley and Sons, New York; c1984.
7. Jagadesh M, Raju M, Rahale CS. Influence of different weed management practices on growth and yield attributes of irrigated blackgram under Cauvery delta zone of Tamil Nadu. J Pharmacogn. Phytochem. 2019;8(3):608-611.
8. Mani VS, Pandita ML, Gautam KC, Das B. Weed killing chemicals in potato cultivation. Proc. Natl. Acad. Sci. U.S.A. 1973;23:17-18.
9. Painkra D, Gupta DK, Sinha AK, Paliwal AK, Chouksey N, Singh SK. Effective weed management strategy for blackgram (*Vigna mungo* L. Hepper) under north hill situation of Chattisgarh. Pharma Innov. 2021;10(7):85-88.
10. Rai CL, Sirothia P, Tiwari R, Panday S. Weed dynamics and productivity of blackgram (*Vigna mungo* L.) as influenced by pre and post emergence herbicides. Res. Crops. 2016;17(1):58-62.
11. Singh RP, Verma SK, Singh RK, Idnani LK. Influence of sowing dates and weed management on weed growth and nutrients depletion by weeds and uptake by chickpea (*Cicer arietinum*) under rainfed condition. Indian J. Agric. Sci. 2014;84(4):468-472.
12. Yadav SC, Singh UP, Padmavati J, Kumar SS, Singh L, Singh H. Influence of weed management practices in blackgram under guava based agri-horti system in Vindhyan region. Biennial Conference of Indian Society of Weed Science on Emerging Challenges of Weed Management, DWSR, Jabalpur; c2014. p. 149.

Author's Details

Nancy Khwairakpam

Scientist, Department of Agronomy, College of Agriculture, Central Agricultural University, Imphal, India

Luikham Edwin

Scientist, Department of Agronomy, College of Agriculture, Central Agricultural University, Imphal, India

Jamkhogin Lhungdim

Scientist, Department of Agronomy, College of Agriculture, Central Agricultural University, Imphal, India

Herojit Singh Athokpam

Scientist, Department of Agronomy, College of Agriculture, Central Agricultural University, Imphal, India

N Brajendra Singh

Scientist, Department of Agronomy, College of Agriculture, Central Agricultural University, Imphal, India

N Okendro Singh

Scientist, Department of Agronomy, College of Agriculture, Central Agricultural University, Imphal, India

N Anando Singh

Scientist, Department of Agronomy, College of Agriculture, Central Agricultural University, Imphal, India

Herojit Khaidem

Scientist, Department of Agronomy, College of Agriculture, Central Agricultural University, Imphal, India

MM Rifana Ajam

Scientist, Department of Agronomy, College of Agriculture, Central Agricultural University, Imphal, India

Surbani Laishram

Scientist, Department of Agronomy, College of Agriculture, Central Agricultural University, Imphal, India

G Prameshchandra Sharma

Scientist, Department of Agronomy, College of Agriculture, Central Agricultural University, Imphal, India

Heisnam Henarita

Scientist, Department of Agronomy, College of Agriculture, Central Agricultural University, Imphal, India

B Keyeang

Scientist, Department of Agronomy, College of Agriculture, Central Agricultural University, Imphal, India

Thangneibong Serto

Scientist, Department of Agronomy, College of Agriculture, Central Agricultural University, Imphal, India

O Yaiphabee

Scientist, Department of Agronomy, College of Agriculture, Central Agricultural University, Imphal, India

M Puja Devi

Scientist, Department of Agronomy, College of Agriculture, Central Agricultural University, Imphal, India