www.ThePharmaJournal.com

The Pharma Innovation



ISSN (E): 2277-7695 ISSN (P): 2349-8242 NAAS Rating: 5.23 TPI 2023; 12(8): 2591-2594 © 2023 TPI

www.thepharmajournal.com Received: 01-05-2023 Accepted: 06-06-2023

Sujith Vaishnav K

PG Scholar, Department of Plantation, Spices, Medicinal and Aromatic Crops, HC & RI, Tamil Nadu Agricultural University, Coimbatore, Tamil Nadu, India

Aneesa Rani MS

Professor (Hort.) and Nodal Officer HC&RI, Jeenur, Paiyur Campus, Krishnagiri, Tamil Nadu, India

Irene Vethamoni P

Dean (Horticulture), Horticultural College and Research Institute, TNAU, Coimbatore, Tamil Nadu, India

Renukadevi P

Professor (Plant pathology), Department of Medicinal and Aromatic Crops, Horticultural College and Research Institute, TNAU, Coimbatore, Tamil Nadu, India

Saraswathi T

Professor, Department of Medicinal & Aromatic crops, HC &RI, Tamil Nadu Agricultural University, Coimbatore, Tamil Nadu, India

Dr. Kewat Sanjay Kumar

Department of Botany, University of Allahabad, Prayagraj, Uttar Pradesh, India

Dr. Awadhesh Kumar

Department of Horticulture, Aromatic and Medicinal Plants, Mizoram University, Aizawl, Mizoram, India

Anbu Megala M

Senior Research Fellow, Department of Plant Biotechnology, Centre for Plant Molecular Biology and Biotechnology, Tamil Nadu Agricultural University, Coimbatore, Tamil Nadu, India

Santhanakrishnan VP

Associate Professor, Department of Medicinal & Aromatic Crops, HC&RI, Tamil Nadu Agricultural University, Coimbatore, Tamil Nadu, India

Corresponding Author: Santhanakrishnan VP

Associate Professor, Department of Medicinal & Aromatic Crops, HC&RI, Tamil Nadu Agricultural University, Coimbatore, Tamil Nadu, India

In vitro anti-fungal and antioxidant studies on the essential oil: *Citrus macroptera* (Wild orange) of Northeast India

Sujith Vaishnav K, Aneesa Rani MS, Irene Vethamoni P, Renukadevi P, Saraswathi T, Dr. Kewat Sanjay Kumar, Dr. Awadhesh Kumar, Anbu Megala M and Santhanakrishnan VP

Abstract

Citrus macroptera, commonly known as 'Wild Lime,' is an underexplored fruit with promising medicinal properties. This study aimed to assess the antifungal and antioxidant activities of *Citrus macroptera* essential oil against three prevalent Plant fungal pathogens, namely *Macrophomina phaseolina*, *Fusarium oxysporum* f. sp. *cubense* and *Colletotrichum gloeosporioides*. *In vitro* assay results indicated that *Colletotrichum gloeosporioides* (98.88%) showed the highest percentage of inhibition, followed by *Macrophomina phaseolina* (92.96%) and *Fusarium oxysporum* (77.40%). The essential oil of *Citrus macroptera* exhibited good antioxidant potential with an IC₅₀ value of 195.96 µg/ml, which revealed its extent of application in both the pharmaceutical and agricultural sectors. Future research is likely to integrate and contrast the identification of the chemical composition of the essential oil, as well as the study of the active component responsible for its antifungal and antioxidant effects.

Keywords: Citrus macroptera, Lunglei-Mizoram, essential oil, antifungal, antioxidant activity

1. Introduction

Citrus is one of the most important horticultural crops grown and traded globally. Citrus has its basic origin in Southeast Asia and is extensively distributed throughout the tropical and subtropical parts of the world. (Moore 2001; National Horticulture Board, 2010) ^[11, 13]. Citrus fruits are beneficial with potential health-promoting chemical components and the key sources of vitamins and minerals (Teigiserova *et al.*, 2021) ^[21]. India is exceptionally rich in both cultivated and wild citrus genetic resources (Nair and Nayar 1997) ^[12].

Citrus peels, the dominant waste in the citrus processing units, are normally discarded. However, recent studies indicated that citrus peels contain a great number of flavonoids, and glycosides, which have excellent anti-inflammatory, anti-carcinogenic, and anti-microbial properties. Apart from the fruit, the citrus peel essential oil contains terpene-based compounds with D– limonene as a major compound which is known for its aroma health benefits and nutritional content. (Han *et al.*, 2021)^[4]

The medicinal properties of citrus are attributed to the bio-active secondary metabolites found in the flavedo and albedo of the fruit peels, including monoterpenes (Limonene), citric acid, phenolic compounds, etc. (Lv *et al.*, 2015; Patil *et al.*, 2017; Zou *et al.*, 2016) ^[8, 15, 24]. The flavanoids and the phenolic compounds present in the citrus peel are responsible for the anti-inflammatory, anti-carcinogenic, antiviral, antibacterial, and anti-allergenic effects (Yashaswini *et al.*, 2018) ^[23].

Citrus macroptera belonging to the family of Rutaceae is commonly known as SatKara, a pharmacologically diverse medicinal plant. This plant's parts, especially the fruit, have a wide variety of traditional medical applications for a wide range of health problems. Numerous active phytochemical components of this plant, including D-limonene, beta-caryophyllene, beta-pinene, geranial edulinine, ribalinine, isoplatydesmine, and terpene-based compounds have been identified (Waikedre *et al.*, 2010)^[5]. Studies indicated that *C. macroptera* fruit, peel and leaves have anti-inflammatory, anti-tumour, anti-microbial, antioxidant, thrombolytic, hypoglycemic, anxiolytic, antidepressant, cardioprotective, and hepatoprotective properties. (Aktar *et al.*, 2017)^[1]

Plant fungi affect crop yield, quality, and profitability (Shuping *et al.*, 2017)^[19]. Synthetic chemical fungicides have damaged the condition of the environment and the health of the soil,

The Pharma Innovation Journal

and their residual toxicity has a substantial impact on nontarget creatures, people's health, and the global economy. (Mahmood et al., 2016; Tripathi et al., 2020)^[9, 22]. Synthetic fungicides can be replaced with plant-based compounds since they are more abundant and safer than their synthetic counterparts. The wide range of naturally occurring plantbased compounds has attracted the attention of researchers. (Shweta Singh et al., 2021) [20]. With this favourable perception towards natural compounds in pest control and for the efficient reuse of citrus peels, the current study was conducted to assess the activity of citrus peel essential oils against significant fungal species, including Macrophomina phaseolina, Fusarium oxysporum, and Colletotrichum gloeosporioides. As citrus peel has antioxidant activity and there is no report on the antioxidant activity of Citrus macroptera essential oil. Consequently, it was subjected to screen the Citrus macroptera essential oil for its antioxidant properties (Miguel, 2010)^[10].

2. Methods and Materials

2.1 Collection and preparation of plant material

Citrus macroptera fruits were collected from the Lunglei district of Mizoram, India (22° 79'51.02" N 92° 22' 47.38" E 1162 m). The fruit was washed thoroughly, peeled and the peels were shade dried for 4-5 days. The dried peels were cut into small pieces and stored for future use in airtight containers.

2.2 Extraction of Essential oils

Peels were powdered, weighed and then taken in a roundbottomed flask. The flask was then filled with twenty times the volume of distilled water as the weight of the peels (1:20 W/V) and placed over the heating mantle. The clevenger apparatus was attached to the round bottomed flask and was boiled at 70 °C for six hours. The essential oil, along with the steam, was distilled in a graduated cylinder. The condensed water that was accumulated in the graduated cylinder was collected at regular intervals. After six hours, the aqueous layer and the oil that had accumulated in the graduated cylinder were separated. The oil obtained was then refrigerated for further investigation.

2.3 Test fungi

The plant pathogenic fungi *Colletotrichum gloeosporioides, Fusarium oxysporum and Macrophomina phaseolina* were obtained from the Department of Plant Pathology, Tamil Nadu Agricultural University, Coimbatore. These cultures were sub-cultured at regular intervals for bioassay experiments.

2.4 Antifungal assay

The antifungal activity of C. macroptera essential oil was performed by the poisoned food technique against all the test fungi. Sterile PDA medium was mixed with the C. macroptera essential oil (CaMaLu) to prepare varying concentrations ranging from 250 ppm, 500 ppm, 750 ppm, and 1000 ppm, which were then equally distributed to all the Petri plates. The media without treatment was taken as the negative control. All the treatments were done in triplicate. After the media had solidified, an actively growing fungal disc of 4 mm-diameter was removed from the maintained mother culture and placed in the middle of the Petri plates containing PDA medium that had been enriched with C. macroptera essential oil. All Petri plates were subjected to an incubation period of their respective growth periods (till the hyphae reaches the edge of the petri plates in the control concentration) at 28±2 °C. After incubation, the fungal mycelial growth was measured. The following formula was used for calculating the percentage of mycelial growth that was inhibited compared to the control (Gopalakrishnan et al., 2014) [3].

Inhibition(%) = $\frac{\text{Growth of pathogen mycelium in control} - \text{Growth of pathogen mycelium in treatment}}{\text{Growth of pathogen mycelium in control}} \times 100$

2.5 DPPH antioxidant activity

DPPH antioxidant activity was calculated using a method adapted by Shimada *et al.* (1992)^[17]. Different concentrations (20-100 µg/ml) of ascorbic acid were prepared with methanol and are used as standards. 50µl of different concentrations (20-100 µg/ml) of *C. macroptera* essential oil were added and mixed with 450 µl of 50 mM Tris HCl (50 mmol/L, pH 7.4). 50 µl of methanol in place of the test sample was used as the blank. To this, 1 ml of 0.1 mM DPPH (0.1 mmol/L in methanol), was added, mixed thoroughly, and incubated for 30 minutes at dark (RT). After incubation, the Free Radical Scavenging activity of essential oil (E-CMaLu) against stable DPPH was determined spectrophotometrically and the UV absorbance was read at 517 nm.

2.5.1 The scavenging effect was calculated using the following equation

Scavenging effect (%) = $\frac{(A0 - A1)}{A0} \times 100$

where A_0 was the absorbance of the control (blank, without essential oil) and A_1 was the absorbance in the presence of the essential oil. Experiments were done in triplicate and IC₅₀ was

measured based on the inhibition percentage of DPPH radicals scavenged.

2.6 Statistical analysis

Three replications of the bioassay were performed using a completely randomized design. Using one-way analysis of variance (ANOVA), the results of several treatments on mycelial growth were investigated. To compare the treatment means at a 5% significance level, Duncan's multiple range test (DMRT) was employed in the Statistical Package for Social Sciences (SPSS version 16.0. Chicago, SPSS Inc. USA).

3. Results

3.1 In vitro Antifungal assay

The antifungal activity of *C. macroptera* essential oil indicated that the growth of the agriculturally important soilborne pathogens *Fusarium oxysporum* f. sp. *cubense, Macrophomina phaseolina,* and *Colletotrichum gloeosporioides* were significantly inhibited in a dose dependent manner at different concentrations, namely 250 ppm, 500 ppm, 750 ppm and 1000 ppm by poisoned food technique Complete growth of fungal mycelium was observed in all the control plates. The results indicated that *Citrus*

The Pharma Innovation Journal

macroptera essential oil peel extract exerted strong antifungal activity in a dose-dependent manner, where percentage inhibition increased with the increase in concentration of the doses.

In vitro antifungal assay of the essential oil of E-CMaLu hindered the growth of *Colletotrichum gloeosporioides* and suppressed the mycelial growth of *Col* by 98.88% at 1000 ppm, which is higher than the other fungal species. The minimum inhibition percentage observed in *Fusarium oxysporum* f. sp. *cubense* 77.4% at 1000 ppm concentration of E-CMaLu and *Macrophomina phaseolina* growth was inhibited at a rate of 92.96%. (Fig 1, Table 1) C. MaLu peel essential oil has a high impact on *C. gloeosporioides*, when compared with other fungi, namely *M. Phaseolina* and *F. oxysporum* f. sp. *Cubense*. The results clearly indicated that, *Citrus macroptera* essential oil has got highest inhibitory activity against C. *gloeosporioides* and *M. phaseolina* and the lowest activity against *F. oxysporum*.

3.2 DPPH antioxidant activity assay

The antioxidant activity of E-CMaLu improved as the volume of essential oil used for the assay was increased from 20 to 100 μ l. In the DPPH assay, E-CMaLu essential oil (20, 40, 60, 80 and 100 μ l) showed an IC₅₀ value of 195.96 μ g/ml. Furthermore, the essential oil of E-CMaLu was able to scavenge DPPH radicals in a concentration dependent manner.



Fig 1: Antifungal activity of *C. macroptera lunglei* essential oil against A) *Fusarium oxysporum* f. sp. *cubense* B) *Colletotrichum gloeosporioides* C) *Macrophomina* phaseolina

 Table 1: Effect of Citrus macroptera essential oil at different concentrations against F. oxysporum, M. phaseolina and C. gloeosporioides

Concentration of	Per cent inhibition over control (%)		
<i>Citrus macroptera</i> Essential oil (ppm)	F. oxysporum	M. phaseolina	C. gloeosporioides
250	26.85±0.14 ^d	12.5±0.58 ^d	10±0.15 ^b
500	42.2±0.1 ^b	31.38±0.03 ^a	32.27±0.67 ^d
750	50.92±0.11°	48.61±0.15°	87.28±0.38°
1000	77.40±0.03ª	92.98±0.11b	98.88±0.09 ^a
Control	0	0	0

Data represented as mean percentage \pm SD and values followed by the same letter along the column are not significantly different (*p*<0.05) from each other.

 Table 2: Evaluation of DPPH antioxidant activity of the Citrus macroptera lunglei ESO

Sample	Concentration (µg/ml)	IC ₅₀ (µg/ml)
E-CMaLu	100 ppm	195.96

4. Discussion

D-Limonene is a prominent constituent found in the essential oils of citrus plants and recognized as one of the most abundant naturally occurring monocyclic monoterpenes (Kim et al., 2013)^[7]. In this study, the inhibition of the mycelial growth of fungi is attributed to the phytochemicals present in the essential oil of E-CMaLu. The antifungal efficacy of Colletotrichum gloeosporioides, Fusarium oxysporum and Macrophomina phaseolina on E-CMaLu is primarily reported in this study. Our findings showed that 1000 ppm of E-CMaLu essential oil extract reduced C. gloeosporioides growth by 98.88 percent and M. phaseolina growth by 92.96 percent compared to systemic (Saravani et al., 2021; Pallavi et., al. 2022) ^[17, 14]. The earlier literature reports suggest that the efficacy of C. maxima, C. sinensis essential oil as free radical scavengers may be due to the antioxidant activity of DL-limonene, the oil's principal component (Priyanka et al., 2010; Junior et al., 2009)^[16, 6]. The essential oil from Citrus macroptera peel extract showed more potential antioxidant activity than the pulp (Sadia et al., (2008)^[2].

5. Conclusion

The findings of the present study demonstrated that the oil isolated from the peel of Citrus essential *macroptera* exhibits *vitro* fungicidal effective in and antioxidant activities and can be used in crop protection due to its wide availability, safety, resistance to pests, benevolent nature towards non-target species, less adverse impact on plant growth, and low cost. In addition, the antioxidant property of the C. macroptera essential oil can be utilized as a natural preservative in the agrofood and in the cosmetic industries.

6. Acknowledgement

Sujith Vaishnav acknowledges Tamil Nadu Agricultural University, Coimbatore. for providing the students fellowship, the authors are thankful for NER Citrus Programme, Department of Biotechnology (BT/PR40087/NER/95/1656/2020) and Vegetable Research Station, Palur (V60ER VCS- Jack) for providing financial support for carrying out the project. The authors also thank Department of Plant Biotechnology, Centre for Plant Molecular Biology and Biotechnology and Department of Medicinal and Aromatic crops, Horticulture College and Research Institute Tamil Nadu Agricultural University, Coimbatore for providing infrastructure facilities to carry out the above research work.

7. References

- 1. Aktar Koly, Tahira Foyzun. Phytochemistry and pharmacological studies of *Citrus macroptera:* A medicinal plant review. Evidence-Based Complementary and Alternative Medicine; c2017.
- Chowdhury, Sadia A, Hossain Sohrab M, Bidyut K Datta, Choudhury M Hasan. Chemical and antioxidant studies of *Citrus macroptera*. Bangladesh Journal of Scientific and Industrial Research. 2008;43(4):449-454.
- 3. Gopalakrishnan, Subarayan Bothi, Thangaraj Kalaiarasi.

Screening of various extracts of the fruits of *Cucumis sativus* Linn. for antimicrobial activity. International Journal of Research and Development in Pharmacy & Life Sciences. 2014;3(5):1200-1205.

- 4. Han Yingjie, Wenxue Chen, Zhichang Sun. Antimicrobial activity and mechanism of limonene against *Staphylococcus aureus*. Journal of Food Safety. 2021;41(5):e12918.
- 5. Waikedre Jean, Annabelle Dugay, Isabel Barrachina, Christine Herrenknecht, Pierre Cabalion, Alain Fournet. Chemical composition and antimicrobial activity of the essential oils from New Caledonian *Citrus macroptera* and *Citrus hystrix*. Chemistry & biodiversity. 2010;7(4):871-877.
- Junior, Mário Maróstica R, Thomaz AA, Rocha Silva E, Gilberto Franchi C, Alexandre Nowill, *et al.* Antioxidant potential of aroma compounds obtained by limonene biotransformation of orange essential oil. Food Chemistry. 2009;116(1):8-12.
- 7. Kim, Young Woo, Min Ji Kim, Bu Young Chung, Du Yeon Bang, Seong Kwang Lim, *et al.* Safety evaluation and risk assessment of d-limonene. Journal of Toxicology and Environmental Health, Part B. 2013;16(1):17-38.
- 8. Lv Xinmiao, Siyu Zhao, Zhangchi Ning, Honglian Zeng, Yisong Shu, Ou Tao, *et al.* Citrus fruits as a treasure trove of active natural metabolites that potentially provide benefits for human health. Chemistry Central Journal. 2015;9:1-14.
- 9. Mahmood Isra, Sameen Ruqia Imadi, Kanwal Shazadi, Alvina Gul, Khalid Rehman Hakeem. Effects of pesticides on environment. Plant, soil and microbes: implications in crop science. 2016;1:253-269.
- Miguel, Maria Graça. Antioxidant and anti-inflammatory activities of essential oils: A short review. Molecules. 2010;15(12):9252-9287.
- 11. Moore, Gloria A. Oranges and lemons: clues to the taxonomy of Citrus from molecular markers. TRENDS in Genetics. 2001;17(9):536-540.
- Nair KN, Nayar MP. Rutaceae in Hajra PK, Nair VJ, Daniel P. eds. The Flora of British India. 1997;4:229-407.
- 13. National Horticulture Board. National Horticulture Database. Ministry of Agriculture, Gurgaon, 2010. http://nhb.gov.in.
- 14. Pallavi HU, Navinraj S, Manikanda Boopathi N, Uma D, Nakkeeran S. *In vitro* evaluation of antifungal activity of lemon peels (*Citrus limon* L. Osbeck) extract against agriculturally important soil-borne plant pathogenic fungi. 2022.
- 15. Patil, Bhimanagouda S, Guddadarangavvanahally Jayaprakasha K, Kotamballi Chidambara Murthy N. Beyond vitamin C: The diverse, complex healthpromoting properties of citrus fruits. Citrus Research & Technology. 2017;38(1):107-121.
- 16. Priyanka Singh, Shukla Ravindra, Prakash Bhanu, Kumar Ashok, Singh Shubhra, Mishra PK, *et al.* Chemical profile, antifungal, anti-aflatoxigenic and antioxidant activity of *Citrus maxima* Burm. and *Citrus sinensis* (L.) Osbeck essential oils and their cyclic monoterpene, DLlimonene. Food and Chemical Toxicology. 2010;48(6):1734-1740.
- 17. Saravani B, Chandra R. *In vitro* and *in vivo* Evaluation of Chemical Fungicides against *Sclerotium rolfsii* causing

Collar Rot of Chickpea. Biological Forum-an international journal. 2021;13(2):10-16.

- Shimada, Kazuko, Kuniko Fujikawa, Keiko Yahara, Takashi Nakamura. Antioxidative properties of xanthan on the autoxidation of soybean oil in cyclodextrin emulsion. Journal of agricultural and food chemistry. 1992;40(6):945-948.
- 19. Shuping DSS, Jacobus Nicolaas Eloff. The use of plants to protect plants and food against fungal pathogens: A review. African Journal of Traditional, Complementary and Alternative Medicines. 2017;14(4):120-127.
- 20. Shweta Singh, Jaiganesh Rengarajan, Iyappan Sellamuthu. Screening plants extracts for Antifungal activity against *Rhizoctonia solani*. Research Journal of Pharmacy and Technology. 2021;14(12):6545-8.
- 21. Teigiserova, Dominika Alexa, Ligia Tiruta-Barna, Aras Ahmadi, Lorie Hamelin, Marianne Thomsen. A step closer to circular bioeconomy for citrus peel waste: A review of yields and technologies for sustainable management of essential oils. Journal of Environmental Management. 2021;280:111832.
- 22. Tripathi Sachchidanand, Pratap Srivastava, Rajkumari Devi S, Rahul Bhadouria. Influence of synthetic fertilizers and pesticides on soil health and soil microbiology. In Agrochemicals detection, treatment and remediation. Butterworth-Heinemann; c2020. p. 25-54.
- 23. Yashaswini P, Arvind. Antimicrobial properties of orange (citrus reticulata var. Kinnow) peel extracts against pathogenic bacteria. Int J Curr Microbiol App Sci. 2018;7(3):737-746.
- 24. Zou Zhuo, Wanpeng Xi, Yan Hu, Chao Nie, Zhiqin Zhou. Antioxidant activity of Citrus fruits. Food chemistry. 2016;196:885-896.