www.ThePharmaJournal.com

The Pharma Innovation



ISSN (E): 2277-7695 ISSN (P): 2349-8242 NAAS Rating: 5.23 TPI 2023; SP-12(9): 873-877 © 2023 TPI

www.thepharmajournal.com Received: 18-06-2023 Accepted: 21-07-2023

Dr. S Shiva Kumar

Department of Veterinary Medicine, P.V. Narsimha Rao Telangana Veterinary University, Hyderabad, Telangana, India

Dr. K Satish Kumar

Professor and University Head, Department of Veterinary Medicine, College of Veterinary Science, Rajendranagar, Hyderabad, Telangana, India

Dr. P Nagaraj

Professor and HOD, Department of Veterinary Medicine, College of Veterinary Science, Korutla, Telangana, India

Corresponding Author: Dr. S Shiva Kumar Department of Veterinary Medicine, P.V. Narsimha Rao Telangana Veterinary University, Hyderabad, Telangana, India

Diagnosis and therapeutic management of staphylococcus associated bacterial dermatitis in dogs

Dr. S Shiva Kumar, Dr. K Satish Kumar and Dr. P Nagaraj

Abstract

In the current investigation, which involved screening 252 dogs for dermatological issues, 52 incidences of bacterial dermatitis were noted. There were 73.07% and 26.92%, respectively, of individuals with mild to moderate and severe bacterial dermatitis. Clinical symptoms of erythema (69.23%), alopecia (57.69%), pruritus (53.84%), crusts (50.00%), papules (34.61%), pustules (28.84%), scales (23.07%), and epidermal collarettes (15.38%) were found in 52 dogs with bacterial dermatitis. All 52 samples from the dogs with bacterial dermatitis that were collected and processed for a detailed culture analysis tested positive for 72 isolates. 52/72 of these were Staphylococci spp. (72.22%), with S. intermedius (65.38%), S. aureus (12.30%), and S. epidermidis (6.13%) making up the 34 isolates. However, 20 of these samples also showed the presence of E. coli in 3 (4.17%) of the samples, Klebsiella spp. in 9 (12.5%), and Pseudomonas spp. in 8 dogs (11.11%) that were mixed with staphylococci. Single and mixed bacterial infections occurred at rates of 61.53 and 38.47%, respectively.

Keywords: Bacterial dermatitis, occurence, symptoms, in vitro, sensitivity and efficacy

Introduction

In the realm of small animal medicine, pyoderma is one of the most typical causes of dermatitis. A bacterial infection of the skin that produces pus is known as pyoderma. Due to the distinct features of canine skin, including a thin stratum corneum, a lack of lipid plug in the hair follicles, and a high skin pH that increases the chance of bacterial invasion, subsequent colonization, and overgrowth, dogs are more susceptible to pyoderma. This could result in bacterial superficial folliculitis. Devriese *et al.* (2005) [12] are two examples. Lesions can be quite superficial, affecting simply the epidermis, or they might involve deeper dermal or subcutaneous tissue structures. Pyoderma is divided into three categories: surface, superficial, and deep pyoderma. According to Scott *et al.* (2003) [29], canine superficial pyoderma is characterized as a superficial bacterial infection of the epidermis and hair follicles that typically develops as a complication of allergy, parasite, endocrine, immune-mediated, conformational, or keratinization problems. Follicular papules, which may or may not be crusted, epidermal collarettes, erythema, hyperpigmentation, and alopecia are more prevalent lesions. The majority of the Staphylococcus intermediusa coagulase positive microorganisms were recovered from an affected dog.

Materials and Methods

The current study focused on clinical cases of dogs who had a history of chronic, recurrent, and persistent skin complaints, including alopecia, pruritus, scratching, and body rubbing along with erythema, papules, and pustules. These dogs were chosen for the study and underwent a thorough clinical examination. Using a sterile swab, various clinical samples, including secretions from the skin lesions, were obtained and then transferred to nutrient broth for analysis. Whole Blood was drawn, and it was examined hemologically. Two sets of sick dogs were created. Clindamycin was administered orally to Group I dogs at a dose rate of 6–11 mg/kg b.wt. once daily, while marbofloxacin was administered orally to Group II dogs at a dose rate of 2–5 mg/kg b.wt. once daily. Based on the resolution of clinical signs, clinical response score, change in haematological parameters, and length of recovery, the therapeutic efficacy was evaluated in both groups. The usage of Vitabest Derm syrup, oral cetrizine tablets, staphban F ointment, and chlorhexidine gluconate shampoo was done in both groups.

Results and Discussion

The goal of the current investigation was to screen dogs for the presence of bacterial dermatitis. 2.94% (252/8576) of all cases of dermatological diseases were reported. The prevalence of dermatological affections in the current study was higher than that of Summers et al. (2014) [37] and lower than that of Sarma et al.'s (2013) [43] prior findings, at 5.6% and 1.3%, respectively. In this study, 52 dogs were discovered to have bacterial dermatitis, of which 38 (73.07%) tested positive for superficial lesions and 14 (26.92%) tested positive for more severe forms. According to the findings of Vasilescu and Togoe (2014) [41] and Kelany and Husein (2011) [17], superficial bacterial dermatitis occurs more frequently than deep bacterial dermatitis. In the current analysis, the prevalence of superficial and deep bacterial dermatitis was generally 15.07% (38/252), and 5.55% (14/252), respectively. This conclusion was very similar to those of Udayasree and Pillai (2006) [38, 40] and Shyma and Vijay Kumar (2011) [35], who similarly found that canine superficial pyoderma occurred at rates of 12.71 and 13.61 percent, respectively.

In the current study dogs suffering from bacterial dermatitis exhibited a wide spectrum of clinical manifestations which in their descending order of frequency were erythema (69.23%), alopecia (57.69%), pruritus (53.84%), crusts (50.00%), papules (34.61%), pustules (28.84%), scales (23.07%), epidermal collarettes (15.38%), hyper pigmentation (9.61%), moth eaten appearance and erosions (7.69% each), edema of toes (3.84%) and nodules (1.92%). Erythema, alopecia, pruritus, papules, and pustules were more prevalent among the clinical symptoms noted in the current study. These findings concurred with those of Kelany and Husein (2011) [17], Beigh *et al.* (2013) [4], Hillier *et al.* (2006) [16], Craig (2003) [10], and Hillier *et al.* 2006 [16].

The majority of the dogs (52) in the current study who had bacterial dermatitis were identified cytologically, and this was later validated by looking at Gram's stained smears of cultures taken from the lesions. 52 samples were used in the cultural examination, which revealed a total of 72 bacterial isolates. Staphylococci spp. were found in all 52 samples according to bacterial isolation investigations, and in 20 samples, they were mixed with gram negative bacteria. Based on cultural traits and biochemical characteristics, Staphylococcus' species identification was confirmed (Castellanos et al., 2011). Among 52 Staphylococci isolates, 34 (65.38%) were S. intermedius, 12 (23.77%) were S. aureus, and 6 (11.53%) were S. epidermidis. The results according to Cavalcanti et al. (2005) [9], Wilkoek et al. (2006) [42] and Bensignor and Germain (2004) [5], S. intermedius was also identified as the primary pathogen in dogs with pyoderma in the current investigation.

Even though an in-vitro antibiogram revealed that the bacteria isolated from the bacterial dermatitis cases were susceptible to a wide range of antibiotics, two formulations (Clindamycin and Marbofloxacin) were chosen to test for efficacy. Based on prior studies and the outcomes of an in-vitro sensitivity test in a pilot research, the antibiotics used in the current investigation were chosen. In the current investigation, the majority of the gram negative isolates and all Staphylococci isolates were in-vitro susceptible to enrofloxacin and clindamycin. Additionally, Kelany and Husein (2011) [17]

advised starting treatment with an antibiotic that is known to be effective against more than 90% of isolates of S.intermedius, the most common causal agent of canine bacterial dermatitis. It was recommended that the corresponding antibiotics be continued for one week after clinical cure to lessen the likelihood of reinfection in accordance with the advice of earlier researchers (Beale *et al.*, 2003; Beco *et al.*, 2013 and Reddy *et al.*, 2014) [2, 3, 27].

Clindamycin hydrochloride was administered orally to Group I dogs once daily at a dose rate of 6–11 mg/kg b.wt. Previous reports on the use of clindamycin in canine bacterial dermatitis came from Bloom and Rosser (2001) [6]. Based on the remission of clinical symptoms, which was consistent with Bloom and Rosser (2001) [6], the therapy had an excellent response in the present study dogs in this group. By the third day, three of the dogs had a day wise percentage of clinical cure, but by the end of the treatment period, every dog had responded to therapy, with 10 of them having excellent clinical responses and the other two having good ones. Bloom and Rosser (2001) [6] also reported that clindamycin had an excellent clinical response. The average number of days it took for the lesions on the dogs in this group to regress and show clinical improvement was 5.0 +/- 0.46 days. According to Udayasree and Pillai (2006) [38, 40] and Reddy et al. (2014) [27], the majority of dogs with bacterial dermatitis needed antibiotic treatment for at least three weeks. This finding was somewhat in agreement with their findings.

The treatment for the dogs in Group II began with the oral administration of marbofloxacin at a dose rate of 2 to 5 mg/kg body weight each day. Regular evaluation of the dogs based on the absence of clinical indicators showed complete clinical cure in five dogs by the third day, although response to therapy (with grades ranging from fair to outstanding) was seen in all the dogs by seven days after therapy started. However, all of the dogs had seen complete therapeutic efficacy and outstanding clinical improvement by day 7. For all 12 dogs in this group II of cases, the average time required for lesions to regress and clinical improvement was 4.66 +/-0.48 days

Although the two medications used in Groups I and II were equally efficient in curing canine bacterial dermatitis, significant variations in the course of recovery were seen. Within 7 days following the start of therapy in Groups I and II, all of the dogs had a response (with grades ranging from fair to excellent). By day 5, 75% of the dogs in Group II showed outstanding clinical recovery, compared to just 66.66% of the dogs in Group I instances, according to the recovery path. However, all of the dogs had seen complete therapeutic efficacy and outstanding clinical improvement by day 7. In Group I and Group II dogs treated with clindamycin and marbofloxacin, respectively, the average time required for clinical recovery with the resolution of symptoms was 5.0+/-0.46 days and 4.66 +/-0.48 days, respectively. The results of the current study are consistent with those of Bloom and Roser (2011) [6] and Paradis, Abbey, and Baker (2001) [25], who believed that clindamycin hydrochloride is superior to marbofloxacin for treating bacterial dermatitis over a 14-day period with an excellent recovery path. Therefore, it may be inferred from the current study that marbofloxacin is a more effective treatment for bacterial dermatitis.

Table 1: The course of recovery in dogs with bacterial dermatitis

Group	Percent of dogs recovered (day - wise)			A viguage days taken for massyons
(n=12)	Day 3	Day 5	Day 7	Average days taken for recovery
I	3 (25%)	5 (66.66%)	4 (100%)	5.16 ± 0.45
II	5 (41.66%)	4 (75%)	3 (100%)	4.66 ± 0.48



Fig 1: In Group I and Group II dogs, the path to recovery

Reference

- 1. Aujla RS, Singh N, Sood N, Gupta PP, Sodhi S. Bacterial dermatitis in dogs in Punjab prevalence and clinico pathological studies. Indian Veterinary Journal. 1997;74:837-840.
- Beale KM. Choosing antimicrobials for infections of the skin. The North American Veterinary Conference Proceedings, Small animal Dermatology; c2003. p. 199-200.
- Beco L, Guaguere E, Mendez C L, Noli C, Nuttall T, Vroom M. Suggested guidelines for using systemic antimicrobials in skin infections: part 2- antimicrobial choice, treatment regimens and compliance. Veterinary Record. 2013;10:1-6.
- 4. Beigh SA, Soodan JS, Tantary H, Tikoo A. Comparative evaluation of antibacterial alone and antibacterial along with zinc in management of pyoderma in canines. Intas polivet. 2013;14(II):388-390.
- 5. Bensignor E, Germain PA. Canine recurrent pyoderma: a multicenter prospective study. Veterinary Dermatology. 2004;15(S1):40-42.
- 6. Bloom PB, Rosser EJ. Efficacy of once daily clindamycin hydrochloride in the treatment of superficial bacterial pyoderma in dogs. Journal of the American Animal Hospital Association. 2011;37:537-542.
- 7. Bond R, Loffler A. What's happened to *Staphylococcus intermedius*, Taxonomic revision and emergence of multi drug resistance. Journal of Small animal practice. 2012;53:147.
- Castellanos L, Rodriguez MG, Santos AR. Isolation and biochemical identification of bacterial organism from infection in dogs, Universidad deli Salle, Bogota, Columbia. Revista de Medicina Veterinaria. 2011;22:21-30.
- 9. Cavalcanti S, Das N, Coutinho SD. Identification and sensitivity of *Staphylococcus* spp. isolates from skin of healthy dogs and from dogs with pyoderma. Clinica Veterinaria. 2005;58(10):60-66.
- 10. Craig M. Diagnosis and management of pyoderma in the dog. In practice. 2003;25(7):421-425.
- 11. Curtseit S, Ciobotaru E, Militaru M, Soare T, Dinescu G. Diagnosis of pyoderma in dogs. Scientific works, C series. 2009;3:80-87.
- 12. Devriese LA, Vancanneyt M, Baele M, Vaneechoutte M, Graef DE, Snauwaert C, *et al. Staphylococcus pseudintermedius spp. nov*; a coagulase positive species from animals; c2005. http://ijs.sgmjournals.org/content/55/4/1569.
- 13. Devriese L, Coll E *Staphylococcus pseudintermedius* versus *Staphylococcus intermedius*. Veterinary Microbiology. 2009;133:206-207.
- 14. Feijo FMC, Souza NFD, Ramadinha RHR. A study of the yeast *Malassezia pachydermatis* by examination of skin cytology in the dog. Revista Brasileira de Medicina Veterinaire. 1998;20:66-68.
- 15. Gera S, Khurana R, Jakhar KK, Garg SL, Arya S. Blood-Biochemical studies in skin affections in dogs. Indian Journal of Veterinary Research. 2009;18(1):23-26.
- Hillier A, Pinchbeck LR, Cole LK. Efficacy of cefpodoxime proxetil in the treatment of canine superficial pyoderma. European Society of Veterinary Dermatology; c2006. p. 209.
- 17. Kelany Wael M, Husein M Galal. Diagnosis of recurrent pyoderma in dogs by traditional and molecular based

- diagnostic assays and its therapeutic approach. Journal of American Science, 2011, 7(3).
- 18. King MD, Humphrey BJ, Wang YF, *et al.* Emerge of community –acquired methicillin- resistant *S. aureus* USA 300 clone as the predominant cause of skin and soft tissue infections. Annual Internal Medicine. 2006;144:309.
- 19. Koshnegah J, Movassaghi AR, Merman R. Survey of dermatological Conditions in a population of domestic dogs in Mashhad North East of Iran (2007-2011). Veterinary Research Forum. 2013;4(2):99-103.
- 20. Leib ME, Monroe WE. Diseases of the integument. Text book of Practical Small; c1997.
- 21. Loeffler A, Baines SJ, Toleman MS. *In-vitro* activity of fusidic acid and mupirocin against coagulase positive *Staphylococci* from pets. Journal of Antimicrobial Chemotherapy. 2008;62:1301-1346.
- Veeranki MB. Clinico-diagnostic and therapeutic studies on superficial pyoderma in dogs. M.V.Sc Research work; c2015.
- 23. Muller GH, Kirk RW, Scott DW. Small animal dermatology 3rd edition W.B. Saunders Co, Philadelphia, USA; c1983.
- 24. Nair SS, Nauriyae. Diagnostic significance of haematological changes associated with various canine dermatoses. Intas polivet. 2007;8(I):68-72.
- 25. Paradis M, Abbey L, Baker B. Evaluation of the Clinical efficacy of marbofloxacin tablets for the treatment of canine pyoderma: An open clinical trial. Veterinary Dermatology. 2001;12:163-169.
- 26. Reddy SB, Nalini Kumari K, Vaikuntarao V, Rayulu VC. Cultural isolates and the pattern of antimicrobial sensitivity of whole cultures from recurrent pyoderma in dogs. Indian Journal of Field Veterinarians. 2011;7(1):40-43.
- 27. Reddy BS, Nalinikumari K, Vaikuntarao V, Rayulu VC, Sivajothi S. Efficacy of enrofloxacin in the treatment of recurrent pyoderma in dogs. Journal of Advanced veterinary Research. 2014;4(3):108-112.
- 28. Rosenkrantz W. Cutaneous cytology a quick review of an indispensable test a supplement to Veterinary Medicine; 2008. p. 20-21.
- 29. Scott D. Antibiotic use in canine pyoderma. Australian College of Veterinary Scientists Science Week Small Animal Medicine Chapter meeting; c2003. p. 20-23.
- 30. Scott DW, Peters J, Miller WH. efficacy of Orbifloxacin tablets for the treatment of superficial and deep pyoderma due to *S. intermedius* infection in dogs. Canadian Veterinary Journal. 2006;47(10):999-1002.
- 31. Senapati SK, Patra RC, Panda HK. Prevalence and antibiogram of bacterial pathogens isolated from canine pyoderma. Indian Journal of Field Veterinarians. 2014;9(3):41-45.
- 32. Sharma J, Gupta GC. Serum protein profiles in naturally occurring dermatological disorders in dogs. Indian Journal of Veterinary Medicine. 2005;25:121-122.
- 33. Sharma SK, Soodan JS, Dutta TK, Rama BB, Tikoo AA. Occurrence of bacterial dermatitis in canines and their antibiogram. Indian Journal of Veterinary medicine. 2008;27:126-127.
- 34. Sharma SK, Soodan JS, Hussain K, Tikoo A. Clinical management of canine bacterial dermatitis. Intas polivet. 2013;14(II):381-384.
- 35. Shyma VH, Vijay Kumar K. Haemato biochemical

- studies in dogs affected with bacterial dermatitis. Journal of Veterinary Animal Sciences. 2011;42:20-22.
- 36. Siugzdaite J, Zamokas G, Grigonis A, Maeijauskas V, Lasys V. Antimicrobial susceptibility of *Staphylococcal species* isolated from dogs with pyoderma. Medycyna Veterinaria. 2008;64(8):991-994.
- 37. Summers JF, Hendricks A, Brodbelt DC. Prescribing practices of primary- care Veterinary Practitioners in dogs diagnosed with bacterial pyoderma. Biomed Central Veterinary Research. 2014;1:240-245.
- 38. Udayasree V, Pillai UN. Treatment of canine pyoderma. Indian journal of Veterinary Medicine. 2006;26(1):75-76.
- 39. Udayasree VJ, Pillai UN. Antibiotic sensitivity patterns of bacterial isolates from canine pyoderma. Indian veterinary journal. 2007;84(5):523-524.
- 40. Udayasree VJ, Pillai UN, Baby PG. Canine pyoderma and its management with cynodon dactylon. Indian Veterinary Journal. 2006;83(12):1274-1276.
- 41. Vasilescu CA, Togoe I. The Morphological properties of the *Staphylococcus species*, strains isolated in canine pyodermas. Bulletin of University of Agriculture Sciences and Veterinary medicine. 2014;71(1):250-255.
- 42. Wilkoek P, Szczepanik M, Blimke Z, Nowak M, Pomorska D. Identification of bacteria from canine pyoderma and their susceptibility to the most commonly used antibiotics in veterinary Dermatology. Medicina Veterinaria. 2006;61:135-141.
- Rao GD, Sarma VV. Contribution of N2O emissions to the atmosphere from Indian monsoonal estuaries. Tellus B: Chemical and Physical Meteorology. 2013 Dec 1;65(1):19660.